

Effect of the nature of protecting group at O-4 on stereoselectivity of glycosylation by 4-O-substituted 2,3-di-O-benzylfucosyl bromides

Alexey G. Gerbst,^a Nadezhda E. Ustuzhanina,^a Alexey A. Grachev,^a Dmitry E. Tsvetkov,^b Elena A. Khatuntseva^b and Nikolay E. Nifant'ev^{*b}

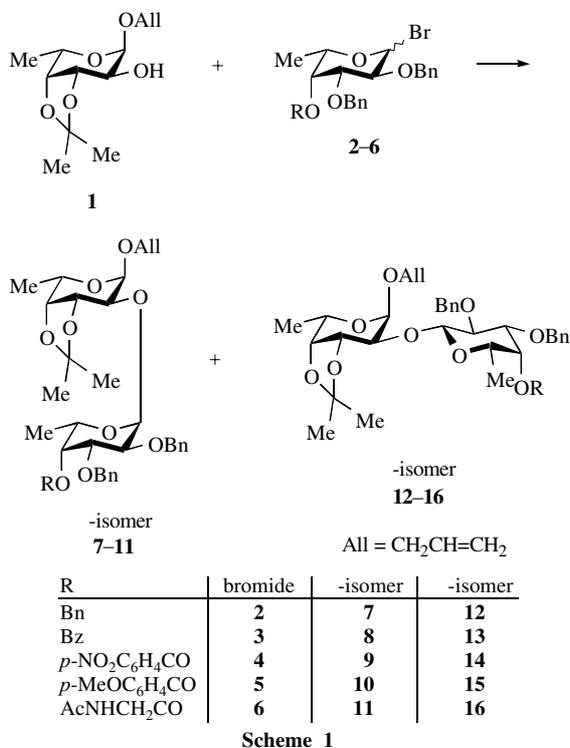
^a Higher Chemical College, Russian Academy of Sciences, 125047 Moscow, Russian Federation

^b N. D. Zelinsky Institute of Organic Chemistry, Russian Academy of Sciences, 117913 Moscow, Russian Federation.

Fax: +7 095 135 8784; e-mail: nen@ioc.ac.ru

The effect of the nature of the substituent at O-4 on the stereoselectivity of glycosylation by 2,3-di-O-benzylfucosyl bromides was studied by direct chemical experiments and computer modelling.

Within the synthesis of fucoidan fragments¹ we performed glycosylation of acetonide **1** by 2,3-di-O-benzylated L-fucosyl bromides **2** and **3** with benzyl and benzoyl protecting groups at O-4 (Scheme 1). In case of 4-O-benzoylated bromide **3** glycosylation was more stereoselective than in case of **2** (Table 1). Similar data on the stereoselectivity of fucosylation were reported before,^{2,3} but the origin of the dependence of the stereoselectivity of fucosylation on the structure of fucosyl donor remains unclear.



To explain the predominance of the α -product in case of glycosylation by **3** we supposed the formation of intermediate cation II (Scheme 2), in which the carbonyl group of benzoate provides intramolecular 'stabilisation' of the cationic centre. Cation II is hindered from the β -side for a nucleophilic attack leading to the formation of the α -glycoside product.

To evaluate the ability of the substituent at O-4 in fucosyl bromide **3** to 'stabilise' the cationic centre at C-1, the difference (E) between the total energy of 'non-stabilised' glycosyl cation I and 'stabilised' glycosyl cation II was calculated using the MM+ force field.⁴ Partial charges were calculated on the AM1 level⁵ of approximation. Both molecular-mechanics and semi-empirical calculations were performed using the HyperChem software[†] (version 5.02). The starting conformations of cations I and II were built using standard geometric parameters and setting the torsion angle H(4)-C(4)-O(4)-C equal to 0° for

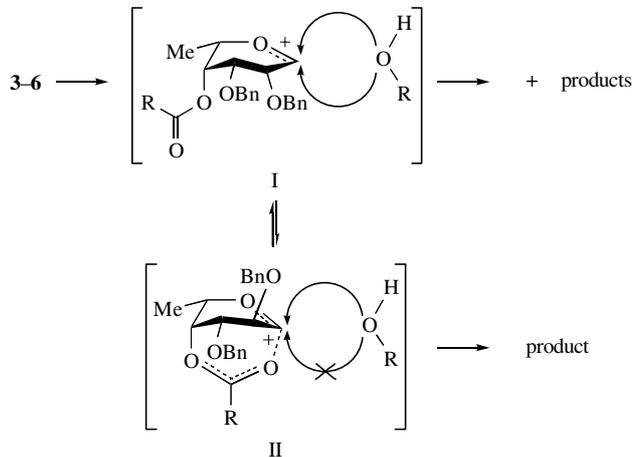
Table 1 The ratios between α - and β -disaccharide products in the glycosylation of acceptor **1** with bromides **2–6** (Scheme 2) and the E values for corresponding cations of type II.

Fucosyl bromide	Substituent at O-4	E /kcal mol ⁻¹	α - and β -disaccharide products	Ratio between α - and β -products
2	Bn	-0.1	7 and 12	1:1 ^a
3	Bz	3.6	8 and 13	3.5:1 ^a
4	<i>p</i> -NO ₂ C ₆ H ₄ CO	2.1	9 and 14	2:1
5	<i>p</i> -MeOC ₆ H ₄ CO	4.7	10 and 15	5:1
6	AcNHCH ₂ CO	1.6 (19.3 ^b)	11 and 16	2:1

^a Experimental ratios of α : β isomers were determined by integration of respective ¹H signals of Fuc residues at the 'non-reducing' end. ^b E value for intermediate III.

cation I and to 180° for cation II. The total geometry optimisation was performed using the Polak-Ribie re conjugate gradient algorithm until the gradient value reached 0.1 kcal mol⁻¹ Å⁻¹. The E value for 4-O-benzoylated compound **2** (Table 1) was close to zero, but it was positive for 4-benzoate **3**, thus confirming the stabilisation hypothesis and explaining the difference in the stereoselectivities of fucosylation by bromides **2** and **3**. Note that all glycosylation reactions were performed in CH₂Cl₂,[‡] which solvates all mentioned cations in a similar manner. This permitted us to neglect solvation effects in the calculations.

To elucidate in more details the stabilising effect of protecting group at O-4, which favours the α -selectivity of fucosylation, we also calculated E for cations with *p*-nitrobenzoyl, *p*-methoxybenzoyl and *N*-acetylaminoacetyl groups at O-4. The E values for *p*-nitrobenzoate **4** and 4-*O*-(*N*-acetylaminoacetyl) derivative **6** (Table 1) were lower than that for benzoylated compound **3**. On the contrary, the E value for *p*-methoxybenzoate **5** was higher than that for benzoate **3**. According to these calculation data, we expected that the α -selectivity of fucosylation should decrease in the order **5** > **3** > **4** > **6** > **2**. These results were later



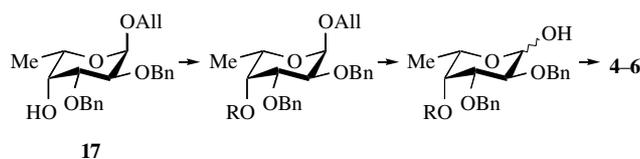
Scheme 2

[†] HyperChem™, Hypercube, Inc., 1115 NW 4th Street, Gainesville, Florida 32601, USA.

Table 2 ¹H NMR data^a for monosaccharides **17–23** and disaccharides **9–11** and **14–16**.

Compound	Fucose residue ^b	Chemical shifts, δ/ppm						Coupling constants, ^c J/Hz			
		1-H	2-H	3-H	4-H	5-H	6-H	J _{1,2}	J _{2,3}	J _{3,4}	J _{5,6}
9	R	4.99	3.87	4.39	4.09	n/d ^c	1.05–1.50 ^d	3.5	7.9	5.7	n/d
	N	5.02	3.86	4.28	5.68	4.53	1.05–1.50 ^d	3.5	9.9	3.1	n/d
10	R	4.98	3.86	4.38	4.09	n/d	1.05–1.50 ^d	3.1	7.9	5.5	n/d
	N	5.01	3.91	4.25	5.66	4.48	1.05–1.50 ^d	3.5	10.0	3.2	n/d
11	R	4.94	3.82	4.34	4.03	n/d	1.05–1.50 ^d	2.9	7.7	5.2	n/d
	N	5.01	3.71	4.12	5.31	4.39	1.05–1.50 ^d	3.2	10.0	3.0	n/d
14	R	5.09	3.91	4.45	4.09	n/d	1.05–1.50 ^d	3.2	8.0	5.1	n/d
	N	4.81	3.69–3.74	4.54	3.76	1.05–1.50 ^d	6.5	n/d	2.9	n/d	
15	R	5.11	3.90	4.45	4.09	n/d	1.05–1.50 ^d	3.9	7.5	5.1	n/d
	N	4.75	3.65	3.75	5.58	3.72	1.05–1.50 ^d	7.1	7.1	3.2	n/d
16	R	5.08	3.88	4.40	4.03	n/d	1.05–1.50 ^d	3.1	8.5	4.9	n/d
	N	4.74	3.60–3.63	5.46	3.62	1.05–1.50 ^d	6.9	n/d	3.5	n/d	
17		4.85	3.81	4.05	3.79	n/d	1.20	3.9	9.5	3.1	6.0
18		4.95	3.89	4.13	5.65	4.25	1.19	3.5	9.5	3.5	7.5
19		4.95	3.93	4.12	5.62	4.21	1.20	3.9	9.1	3.0	6.0
20		n/d	3.83	4.07	5.52	4.16	1.20	4.0	10.0	3.9	6.8
21		5.35	3.87	4.10	5.65	4.42	1.22	3.2	10.0	3.4	6.9
		n/d	3.65	3.71	5.59	3.82	1.31	7.4	9.8	3.1	6.3
22		5.28	3.89	4.01	5.58	4.31	1.18	3.6	10.0	2.9	6.5
		5.24	4.09	3.63	5.51	3.73	1.21	n/d	7.8	2.8	6.0
23		5.23	4.06	3.76	5.48	4.33	1.15	3.5	9.0	4.0	6.7
		n/d	3.48	3.73	5.43	3.84	1.25	8.0	9.5	3.1	6.7

^aNMR spectra were recorded on a Bruker AM-300 instrument (300 MHz) in CDCl₃ at 303 K. Assignment was performed by 2D ¹H–¹H correlation spectroscopy. ^bR is the ‘reducing’ end (*i.e.*, a fucose residue attached to allyl aglycon), N is the ‘non-reducing’ end (*i.e.*, a fucose residue attached to fucose aglycon). ^cn/d = not determined. ^dSignals of 6-H in all Fuc residues in the spectra of mixtures of disaccharide pairs (**9,14**), (**10,15**) and (**11,16**) were not assigned because of overlapping. ^eFor all compounds, J_{4,5} < 1 Hz.

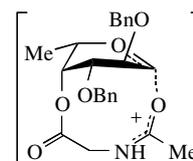
**17**All = CH₂CH=CH₂

R		
<i>p</i> -NO ₂ C ₆ H ₄ CO	18	21
<i>p</i> -MeOC ₆ H ₄ CO	19	22
AcNHCH ₂ CO	20	23

Scheme 3

[‡] Preparation of bromides **4–6**. Esters **18** {[α]_D –1 34° (*c* 2, CHCl₃)} and **19** {[α]_D –1 62° (*c* 1, CHCl₃)} were prepared in 80–90% yields by acylation of compound **7** (1 mmol) with a corresponding acylchloride (4 mmol) in 40 mmol of pyridine in the presence of a catalytic amount of *N,N*-dimethylaminopyridine. Amide **20** {[α]_D –6 2° (*c* 1, CHCl₃)} was obtained in 75% yield by reaction of **7** with equimolar amounts of *sym* anhydride of *N*-acetylglycine and *N,N*-dimethylaminopyridine. Deallylation of esters **18–20** in the presence of PdCl₂ (0.4 mmol) in methanol gave semiacetals **21–23** with 75–80% yields. The bromination with CBr₄ and Ph₃P (1.1 mmol each) in 5 ml of boiling methylene chloride resulted in formation of bromides **4–6** in almost quantitative yields. Fucosyl bromides were used directly in glycosylation reactions without special purification.

Glycosylation with bromides 2–6 (typical procedure). A solution of 1 mmol of acetonide **1**, 1.5 mmol of Hg(CN)₂, 10–20 mg of HgBr₂ and 1.4 g of molecular sieves 4 Å were stirred for 1 h at room temperature under Ar, and a solution of 1.5 mmol of a corresponding fucosyl bromide was added portionwise within 1 h at room temperature. The mixture was additionally stirred for 24 h at room temperature, then filtered through Celite, diluted with CH₂Cl₂, washed with saturated aqueous KBr and NaHCO₃ solutions, filtered through cotton wool and concentrated *in vacuo*. The residue was subjected to flash column chromatography to separate a mixed fraction of - and -disaccharide products. The ratio between the products was determined from the ¹H NMR spectra (Tables 1 and 2). The anomeric configurations of Fuc residues in disaccharides **7–16** at the ‘non-reduced’ end were confirmed by characteristic values of J_{1,2}, which were 3–3.5 Hz for -anomers and 7–7.5 Hz for -anomers.

**III**

proven experimentally by chemical glycosylations (Table 1), except for the coincidence of stereochemical outcomes in glycosylations with compounds **4** and **6**.

For compound **6** which comprises a carbonyl of the amido group along with the ester carbonyl, the hypothetical intermediate III can also be expected in addition to cation II. The *E* value for III is higher (Table 1) than that for ester-stabilised cations of the type II.

Taking into account too high *E* value for intermediate III, we can expect high -stereoselectivity of fucosylation with bromide **6**. However, the ratio between - and -disaccharides in the glycosylation with bromide **6** was as low as 2:1. This result argues that the glycosylation proceeds preferentially *via* cation II rather than cation III.

In conclusion, the data obtained show the mechanism of the influence of the substituent at O-4 on the -stereoselectivity of glycosylation by 2,3-di-*O*-benzylfucosyl donors. Molecular-mechanics calculations according to the described procedure can be successfully applied to the estimation of the stereoselectivity of glycosylation.

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