

# One-step synthesis of shidasterone 22*S*-analogue from ecdysterone

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Shidasterone 22*S*-epimer was synthesised by the interaction of ecdysterone and trifluoroacetic anhydride in chloroform.

Ecdysteroids control the moulting and metamorphosis processes in insects and crustaceans.<sup>1</sup> At the same time, ecdysteroids and their analogues are of great interest for medicine. Derivatives of ecdysterone and -ecdysone with a tetrahydrofurane ring in a side chain were tested for antitumour activity.<sup>2</sup> Shidasterone isolated from the plant *Blechnum niponicum*<sup>3</sup> is known to contain an ether linkage between C-22 and C-25, as was confirmed by mass and <sup>13</sup>C NMR spectra.<sup>4</sup> Shidasterone was synthesised by six-step transformation of ecdysterone, and the C-22 chiral centre in natural shidasterone was determined to be of the (*R*)-configuration.<sup>5</sup>

The paper deals with the one-step transformation from ecdysterone **1** to shidasterone 22*S*-analogue **2**, which proceeds under the action of a two-fold molar amount of trifluoroacetic anhydride.<sup>†</sup>

The dehydration of **1** into **2** was confirmed by the mass spectrum (MS) of the latter. The MS of **2**‡ contained the ions with *m/z* of 444, 426 and 408 corresponding to the fragmentation of M<sup>+</sup> with the elimination of one, two and three H<sub>2</sub>O molecules, respectively, while that of initial compound **1** contained the ions M<sup>+</sup> (*m/z* 480) and 462, 444, 426 and 408 corresponding to the sequential elimination of four H<sub>2</sub>O molecules from M<sup>+</sup>.<sup>6</sup> Just as in the case of the known shidasterone, a downfield shift of the C-25 signal in the <sup>13</sup>C NMR spectrum of epimer **2** (δ 81.3 ppm) respecting the corresponding signal of **1** (δ 71.4 ppm) confirmed the formation of a C-22-C-25 ether

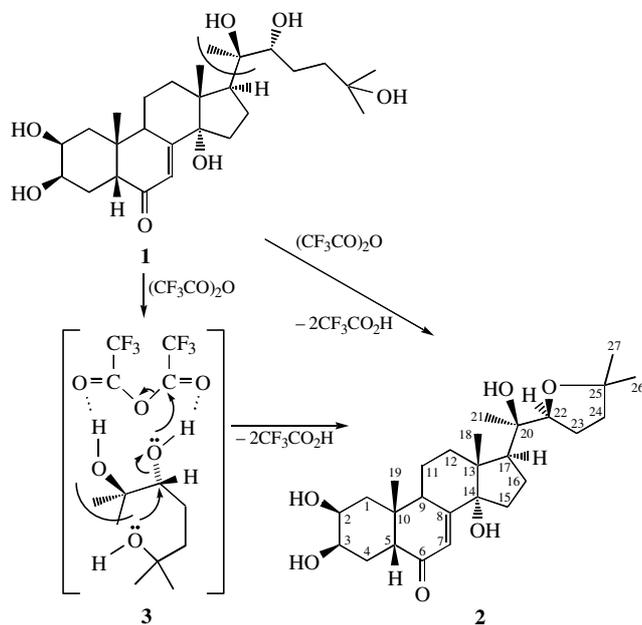
linkage. On the other hand, an upfield signal of the chiral C-22 atom of **2** was observed with reference to the corresponding signal of shidasterone (δ 7.5 ppm). This fact evidenced the formation of the shidasterone 22*S*-analogue in our case. The <sup>1</sup>H NMR spectrum of 22*S*-epimer **2** differed from that of 22*R*-shidasterone by the position of two singlets of the geminal methyl groups of tetrahydrofurane ring, 26-Me and 27-Me (δ 1.56 and 1.58 ppm for epimer **2** and δ 1.24 and 1.25 ppm for shidasterone).<sup>5</sup>

Thus, the reaction changes the configuration of the C-22 atom by intramolecular S<sub>N</sub>2 reaction with an attack of the 25-hydroxyl at C-22 in intermediate complex **3**.

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† A mixture of trifluoroacetic anhydride (0.35 g, 166.4 mmol) and ecdysterone **1** (0.4 g, 83.2 mmol) in 5 ml of chloroform was stirred for 15 min at room temperature. After homogenisation of the reaction mixture, TLC analysis (Silufol, CHCl<sub>3</sub>-MeOH, 5:1) has evidenced the presence of **1** (*R<sub>f</sub>* 0.36) and **2** (*R<sub>f</sub>* 0.48) in a ~1:1 ratio. The product was chromatographed on a silica gel column (eluent CHCl<sub>3</sub>-MeOH, 5:1) to give 0.16 g of **1** and 0.15 g (37.5%) of **2**, mp 180–182 °C (EtOAc), [α]<sub>D</sub><sup>25</sup> +34.5° (c 0.17, MeOH), [α]<sub>D</sub><sup>25</sup> +23.5° (c 0.08, CHCl<sub>3</sub>); {for 22*R*-shidasterone [α]<sub>D</sub><sup>25</sup> +65.0° (c 0.18, CHCl<sub>3</sub>)<sup>5</sup>}, IR (KBr, ν/cm<sup>-1</sup>): 3400 (*w<sub>h/2</sub>* 305), 1635 (*w<sub>h/2</sub>* 80). UV (λ<sub>max</sub>/nm): 242. Found (%): C, 70.10; H, 9.15. Calc. for C<sub>27</sub>H<sub>42</sub>O<sub>6</sub> (%): C, 69.68; H, 9.07.

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‡ Spectral data for **2** [the signals in the <sup>13</sup>C NMR spectrum were assigned using a pulse sequence of *J*-modulated spin echo (JMOD)]: <sup>1</sup>H NMR (300 MHz, CD<sub>3</sub>OD) δ: 5.80 (d, 1H, 7-H, *J* 2.1 Hz), 3.94 (m, 2H, 3-H and 22-H, *w<sub>h/2</sub>* 20.0 Hz), 3.85 (dm, 1H, 2-H, *J* 12.0 Hz), 3.15 (ddd, 1H, 9-H, *J* 12.0, 7.5 and 2.1 Hz), 2.36 (m, 2H, 5-H and 17-H, *w<sub>h/2</sub>* 25.0 Hz), 2.15 (m, 1H, 12-H<sub>ax</sub>), 2.0–1.6 (m, 14H, 6CH<sub>2</sub>, 1-H<sub>eq</sub> and 12-H<sub>eq</sub>), 1.58 (s, 3H, 27-Me), 1.56 (s, 3H, 26-Me), 1.42 (dd, 1H, 1-H<sub>ax</sub>, *J* 12.0 and 13.0 Hz), 1.21 (s, 3H, 21-Me), 0.99 (s, 3H, 19-Me), 0.90 (s, 3H, 18-Me); <sup>13</sup>C NMR (75.5 MHz, CD<sub>3</sub>OD) δ: 206.5 (C-6), 168.0 (C-8), 122.2 (C-7), 85.2 (C-14), 81.8 (C-25), 77.9 (C-22), 77.1 (C-20), 68.7 (C-2), 68.5 (C-3), 51.8 (C-5), 50.6 (C-17), 48.6 (C-13), 39.3 (C-10), 37.4 (C-1, C-24), 35.1 (C-9), 32.9 (C-4), 32.3 (C-12), 31.8 (C-15), 26.9 (C-23), 26.0 (C-26), 25.7 (C-27), 24.4 (C-19), 21.8 and 21.5 (C-11 and/or C-16), 20.7 (C-21), 18.1 (C-18). MS, *m/z*: 444 (21, [M - H<sub>2</sub>O]<sup>+</sup>), 426 (100, [M - 2H<sub>2</sub>O]<sup>+</sup>), 411 (22, [M - Me - 2H<sub>2</sub>O]<sup>+</sup>), 408 (28, [M - 3H<sub>2</sub>O]<sup>+</sup>), 393 (16, [M - Me - 3H<sub>2</sub>O]<sup>+</sup>), 345 (52, [M - 99 - H<sub>2</sub>O]<sup>+</sup>), 327 (69, [M - 99 - 2H<sub>2</sub>O]<sup>+</sup>), 309 (24, [M - 99 - 3H<sub>2</sub>O]<sup>+</sup>), 300 (64).