

Catalytic Hydrogenation/Enzymic Hydrolysis as a Route from Alkyl Alka-2,4-dienoates to (Z)-Alk-3-enoic Acids

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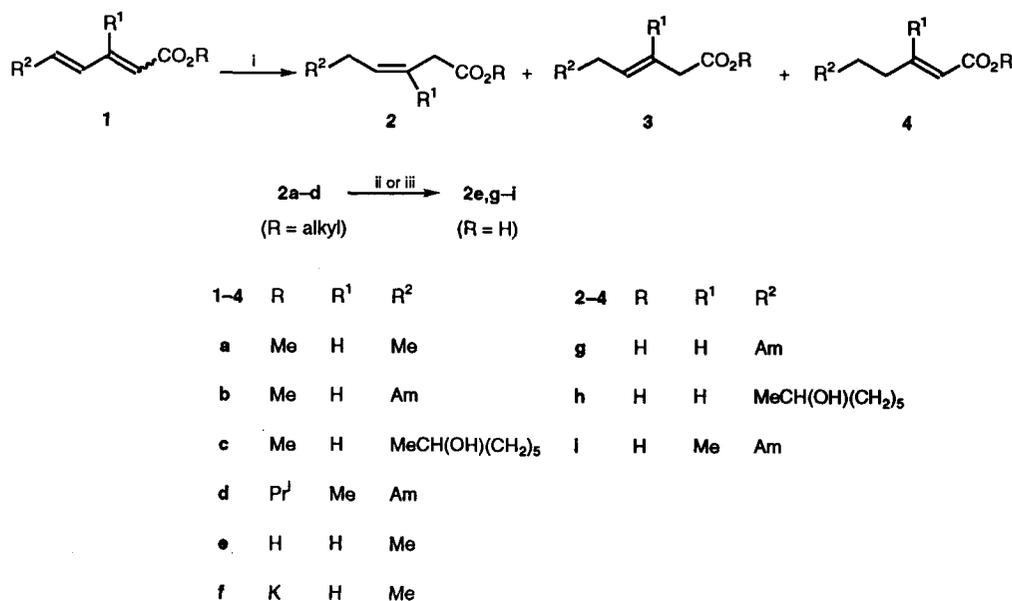
(Z)-Alk-3-enoic acids of no less than 94% geometrical purity have been obtained from the respective alkyl alka-2,4-dienoates after hydrogenation over chromium hexacarbonyl followed by hydrolysis of the resulting products using porcine pancreatic lipase or KOH in H₂O/MeOH.

(Z)-Alk-3-enoic acids, some of which are the pheromones of certain coleopterous insects or intermediates used in their synthesis,¹⁻³ are most often obtained either by the partial reduction of the triple bond in alk-3-ynoic acids (see, e.g., refs. 3-5) or by Wittig *cis*-olefination of alkanals (see, e.g., refs. 6-7). We have studied the feasibility of preparing (Z)-alk-3-enoic acids from the derivatives of respective alka-2,4-dienoic acids using hydrogenation over chromium hexacarbonyl as the key step.

The 1,4-addition of H₂ to the complexes formed on interaction of conjugated dienes with Cr(CO)₆ occurs with high *cis* stereoselectivity,^{8,9} and in recent years has successfully been applied to the synthesis of fragrance substances.^{10,11} However, only one case of the desired transformation has been reported, namely, the reduction of alkyl sorbates, such as **1a**, to the respective (Z)-hex-3-enoates of type **2a** (refs. 10 and 11).

We have found that the above procedure can be successfully extended to the higher dienoates **1b-d** (Table 1). Just as in the case of methyl sorbate (**1a**), the 1,4-hydrogenation of **1b** and **1c** affords, in addition to the target methyl (Z)-alk-3-enoates **2a-c**, minor quantities of their 3(*E*)- and 2(*E*)-isomers (**3a-c** and **4a-c**, respectively). Hydrogenation of the dienoate **1d**, the molecule of which contains a methyl group at C(3), proceeds with practically 100% selectivity (Scheme 1).

It is to be noted that the starting dienoates **1b-d**, prepared by Horner-Emmons olefination of aldehydes R²CHO with allylic phosphonates (EtO)₂P(O)CH₂C(R¹)=CHCO₂R, contained the 2(*E*), 4(*E*)- and 2(*Z*), 4(*E*)-isomers in the ratios 90:10 (for **1b,c**) or 65:35 (**1d**). However, in each case both of these stereoisomers on hydrogenation over Cr(CO)₆ gave the respective (Z)-alk-3-enoates as the main reaction product.



Scheme 1 Reagents and conditions: i, H₂ (50 atm)/Cr(CO)₆ (ca. 10 mol.%), 160-180 °C, 2-4 h, for solvents, see Table 1; ii, KOH (3 equiv.)/H₂O-MeOH (1:3, v/v), r.t., then HCl to pH 2; iii, PPL (25% w/w)/0.05 M phosphate buffer (pH 7.0)-Et₂O (1:5, v/v), r.t., with gradual addition of 1 M aqueous NaOH to maintain the pH at 7.0.

Table 1 The products of hydrogenation of alka-2,4-dienoic acid derivatives over chromium hexacarbonyl.

Substrate (final product)	Solvent	Yield ^{a,b} (%)	Isomer composition ^{b,c} (%)			B.p. /°C (Torr)
			2	3	4	
1a (2e)	Hexane	59	87	2	11	75(20)
		(75) ^d (80) ^e	(84) ^d (95) ^e	(2) ^d (2) ^e	(14) ^d (3) ^e	
1b (2g)	Hexane	70	96	1	3	106(10)
		(78) ^d (50) ^e	(93) ^d (99) ^e	(1) ^d (1) ^e	(6) ^d (0) ^e	
1c (2h)	Benzene	60	96	2	2	140 (3)
		(72) ^d	(94) ^d	(2) ^d	(4) ^d	
1d (2i)	Benzene	62	100	0	0	90 (2)
		(74) ^d	(95) ^d	(0) ^d	(5) ^d	
1e(2e)	Benzene	40 ^f	12	11	77	125-130(40)
1f(2e^g)	H ₂ O	70 ^g	36 ^g	8 ^g	56 ^g	125-130(40) ^g

^aYield of the isolated mixture of isomers. ^bData pertaining to the mixture of isomeric acids obtained after hydrolysis of the hydrogenation products are given in parentheses. ^cDetermined from GLC analyses and/or ¹H NMR spectra. ^dAlkaline hydrolysis. ^eEnzymic hydrolysis, at 50-80% conversion. ^fWith extensive resinification. ^gFor the mixture of acids **2e**, **3e** and **4e** upon acidification of the crude hydrogenation product.

Mild alkaline hydrolysis of esters **2a-d** liberated the corresponding target acids **2e, g-i**. Unfortunately, the hydrolysis was accompanied by partial isomerisation, mainly to the 2(*E*)-counterparts (**4e, g-i**), to the extent of ca. 5%.

In order to avoid this isomerisation, which was obviously due

to the effect of the alkaline medium, we made use of an early observation¹² that porcine pancreatic lipase (PPL) hydrolyses the esters of (*E*)-alk-2-enoic acids much slower than their unconjugated isomers. In fact, the PPL-catalysed hydrolysis of esters **2a** and **2b** at pH 7.0 afforded samples of acids **2e** and **2g** of higher geometrical purity with respect to the starting esters (Table 1) whereas admixtures of the 2(*E*)-isomers accumulated in unconverted material. Interestingly, the enzymic hydrolysis in the biphasic system phosphate buffer–Et₂O proceeds much slower for **2b** than for **2a**; this may be due to their different lipophilicity.

We have also tried to obtain the (*Z*)-alk-3-enoic acids by circumventing the hydrolysis step. To this end, the hydrogenation of the free sorbic acid (**1e**) and its potassium salt (**1f**) was studied. To our surprise, the hydrogenation of **1e** and **1f** proceeded with low selectivity (Table 1); in the former case it was accompanied by extensive resinification. Attempts to hydrogenate the derivatives of **1e**, in which the carboxyl group was protected by groups easily removable under neutral or mild acidic conditions (R = SiMe₃, CH₂OMe, CH₂CCl₃), were equally unsuccessful.

The preparative value of the two-step synthetic sequence developed depends on the accessibility of the starting dienoates and is illustrated by the transformation of **1b** to (*Z*)-dec-3-enoic acid **2g**, the aggregation pheromone of the furniture carpet beetle *Anthrenus flavipes* (*Dermestidae*),¹ and the formal synthesis of (±)-ferrulactone II, the pheromone of the rusty grain beetle *Cryptolestes ferrugineus* (*Cucujidae*), for which the hydroxy acid **2h** is the nearest precursor (see refs. 2–7). Our results also allow for the prospect of employing the lipase-catalysed hydrolysis for a clean separation of alkyl (*E*)-alk-2-enoates from their non-conjugated isomers in otherwise inseparable mixtures.

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