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**In-depth ^{27}Al NMR investigation of $\text{Al}(\text{C}_6\text{F}_5)_3$ and its complexes
with Lewis bases**

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1. General information

All reagents were purchased from Acros, Aldrich or ABCR and used without further purification. Hexane, toluene, benzene were distilled over Na-benzophenone ketyl anion-radical and stored over 4Å Linde type molecular sieves. C₆D₆ was vacuum transferred from potassium metal and stored over 4Å Linde type molecular sieves in a high-quality glovebox (O₂, H₂O < 1 ppm) free of donor solvent vapors. All reactions outside a glovebox were carried out under argon atmosphere using standard Schlenk technique. ¹H, ¹⁹F, ¹³C, ³¹P, ²⁷Al NMR were recorded on Bruker AVANCE III HD 300 spectrometer (300.1, 282.4, 75.5, 121.5, 78.2 MHz, respectively) in dry C₆D₆. Chemical shifts in C₆D₆ were measured relative to residual C₆D₅H (δ = 7.16 ppm) for ¹H or C₆D₆ (δ = 128.06 ppm) for ¹³C. Chemical shifts for ¹⁹F are relative to external CFCl₃ in CDCl₃ sample (δ = 0 ppm).

2. Experimental procedures

2.1. Synthesis of Al(C₆F₅)₃·0.5toluene

Published procedure of E. Y.-X. Chen *et al.* (S. Feng, G. R. Roof and E. Y.-X. Chen, *Organometallics*, 2002, **21**, 832–839) was used. In a high-quality glovebox (O₂, H₂O < 1 ppm) free of donor solvent vapors, a Schlenk tube was charged with B(C₆F₅)₃ (491 mg, 0.96 mmol), sealed with glass stopper, and removed outside the glovebox. On a Schlenk line, absolute toluene (1.0 mL) and absolute *n*-hexane (3.0 mL) were added. To the resulted suspension, a 2.0 M solution of AlMe₃ in *n*-hexane (0.48 mL, 0.96 mmol) was added dropwise. During AlMe₃ addition all solids gradually dissolved. The obtained solution was left without stirring while being opened to an argon line. White crystalline solids precipitated out in 2–3 hours. After that the Schlenk tube was placed to a freezer (–18°C) and left there overnight, then connected to a Schlenk line, and the mother liquor was decanted by Pasteur pipette. The residual solids were washed with absolute *n*-hexane 3 times, dried under high vacuum, and transferred to a glovebox. Product Al(C₆F₅)₃·0.5toluene was obtained as a white crystalline solid (325 mg, 64% yield).

¹H NMR (300.1 MHz, C₆D₆) δ: 7.16–7.10 (m, ca. 2H), 7.09–7.03 (m, ca. 1H), 7.03–6.97 (m, ca. 1H) (a mixture of partially deuterated toluenes), 2.10 (s, 3H, CH₃). **¹⁹F NMR (282.4 MHz, C₆D₆) δ:** –122.8 to –123.1 (m, 2F), –150.8 (tt, *J* = 20.2, 3.0 Hz, 1F), –160.5 to –160.8 (m, 2F). **¹³C{¹H} NMR (75.4 MHz, C₆D₆) δ:** 151.8–147.8 (m, CF in C₆F₅), 144.4–140.3 (m, CF in C₆F₅), 138.4–138.0 (m, C in partially deuterated toluenes), 139.3–135.1 (m, CF in C₆F₅), 129.7–129.3 (m, CH/CD in partially deuterated toluenes), 128.9–128.5 (m, CH/CD in partially deuterated toluenes, overlapped with C₆D₆), 125.6–125.1 (m, CH/CD in partially deuterated

toluenes), 111.5 (br. t, $J = 45$ Hz, Al-C), 21.5–21.1 (m, CH₃ in partially deuterated toluenes). ²⁷Al NMR (78.2 MHz, C₆D₆) δ : 134 \pm 2 ($W_{1/2}$ 12 kHz).

2.2. NMR investigations of Al(C₆F₅)₃·complexes with Lewis bases

2.2.1. Preparation of Al(C₆F₅)₃·complexes

In a high-quality glovebox (O₂, H₂O < 1 ppm) free of donor solvent vapors, NMR tube was charged with Al(C₆F₅)₃ (20–25 mg), dissolved in absolute C₆D₆ (0.50 mL) and sealed with septum. Outside the glovebox, appropriate Lewis base (3 equiv. of pyridine, 2,6-lutidine, P(OEt)₃, MeCN, Me₂S or 30 equiv. of THF) was added *via* microsyringe.

2.2.2. High-quality ²⁷Al NMR spectra acquisition and processing

High-quality ²⁷Al NMR spectra were recorded on "Bruker AVANCE III HD 300 MHz" spectrometer (²⁷Al, 78.2 MHz), and "Bruker AVANCE III WB 400 MHz" spectrometers (²⁷Al, 104.3 MHz), equipped with solid-state 1 kW AVANCE III electronic unit with high-speed response (<0.1 μ sec) for the possibility of recording wideline signals >10–20 kHz with extremely short T₂ relaxation times to avoid signal loss in solution. ²⁷Al chemical shift scale sets by Bruker instrument automatically using ²H lock channel (or the field was set manually using the same ¹H solvent signal) with compensation of magnetic susceptibility for different samples, normalized to external aluminum aqua cation water solution standard ([Al(H₂O)₆]³⁺, 0 ppm). ²⁷Al spectra were recorded using single-pulse sequence, with a dwell time of not more than 1.0–1.5 μ sec (corresponding to the spectral width ~4000–4500 ppm or ~350–450 kHz), with long acquisition time (typical 0.8 sec) and long recycle delay (typical 0.7 sec) to get the clearest signal without any losses, including for narrow signals of high-symmetrical aluminum and aluminum glass signal with slow T₁ relaxation in solid state. Typical T₂ relaxation times (equivalent to FID length) of studied aluminum samples in solution are very short (0.2–1.0 msec), while T₁ relaxation times can be much longer, especially for solid state aluminum signal from glass. T₁ relaxation times can reach more than 200–300 msec, and therefore require long delays in the sum of more than a 1 sec in order to avoid signal losses during the accumulation process. Correct accumulation of the solid-state glass signal is very important for further processing of the spectra and for obtaining the final clear difference spectra. Since aluminum ²⁷Al has a very good sensitivity in NMR, typical summary recording time of ²⁷Al spectra is from 4.5 min (176 scans) to 33 min (1280 scans), which is sufficient to obtain a clear signal from ~10 mg (or more) of the aluminum complex in 0.6 mL solution. If necessary, the recording time can be increased for more complex cases. The signal width of the target ²⁷Al signals ~10–20 kHz does not require high power radiofrequency field, so

standard hard 90° ^{27}Al pulses of 8–12 μsec long (30–50 W) were used. Further processing of the spectra involves subtracting the glass spectrum from the raw data of the sample to obtain difference aluminum ^{27}Al spectra containing a pure target ^{27}Al signal, and this can be done both with the direct FID processing and with the processed spectra after the Fourier transform (the result will be identical), using the standard Bruker Topspin software (usually 3 and 4 versions). The required subtraction accuracy and the quality of signal acquisition from the glass depend on the intensity and width of the aluminum ^{27}Al target signal. However, it is very important to take into account that in many cases, in order to obtain a pure aluminum ^{27}Al signal from the target complex, which does not contain any perturbations from the glass, all manipulations should be carried out with high accuracy. So, both ^{27}Al raw data sets for processing (sample and glass without sample) should be received from exactly the same NMR tube under exactly the same acquisition parameters, with very precise setting of shimming to compensate for magnetic inhomogeneity, temperature control, compensation of magnetic susceptibility of the sample, and chemical shift scale calibration, that in sum afforded a very good and clear spectrum. Ideally, the same solvent should be used for both target sample and blank glass sample. Determination of chemical shift and signal width ($W_{1/2}$) has some peculiarities compared to usual NMR, the most correct chemical shift and width parameters of the recorded signal will be obtained by line shape simulation that could be easily performed using standard TopSpin software package ("solid line shape analysis" program package ("SOLA") in Bruker TopSpin software, usually 3 and 4 versions), using "Gauss/Lorentz" model for simulation. Other details and a detailed description of the ^{27}Al approach can be also found in the main text of the article.

2.2.3. NMR data of $\text{Al}(\text{C}_6\text{F}_5)_3$ complexes

$\text{Al}(\text{C}_6\text{F}_5)_3$ ·Pyridine complex

^1H NMR (300.1 MHz, C_6D_6) δ : 8.43 (dt, $J = 4.6, 1.7$ Hz, 1H, CH in Pyridine), 6.98 (tt, $J = 7.9, 1.8$ Hz, 1H, CH in Pyridine), 6.65 (ddd, $J = 7.6, 4.5, 1.5$ Hz, 1H, CH in Pyridine), 2.12 (s, 3H, CH_3 in partially deuterated toluenes) [*Coordinated and non-coordinated pyridines are in a fast exchange and give one set of signals*]. **^{19}F NMR (282.4 MHz, C_6D_6) δ :** –121.7 to –121.9 (m, 2F), –151.6 (tt, $J = 20.0, 2.7$ Hz, 1F), –160.8 to –161.1 (m, 2F). **$^{13}\text{C}\{^1\text{H}\}$ NMR (75.4 MHz, C_6D_6) δ :** 152.8–148.9 (m, CF in C_6F_5), 149.8 (s, CH in Pyridine), 144.8–140.7 (m, CF in C_6F_5), 139.8–135.7 (m, CF in C_6F_5), 137.6 (s, CH in Pyridine), 124.5 (s, CH in Pyridine), 114.4–112.6 (m, Al–C), 21.8–21.5 (m, CH_3 in partially deuterated toluenes), (C,CH/CD in partially deuterated toluenes are overlapped with the other signals) [*Coordinated and non-coordinated*

pyridines are in a fast exchange and give one set of signals]. **²⁷Al NMR (78.2 MHz, C₆D₆) δ:** 127±1 (*W*_{1/2} 4.0 kHz).

Al(C₆F₅)₃·2,6-Lutidine complex

¹H NMR (300.1 MHz, C₆D₆) δ: 7.14–7.09 (m, partially deuterated toluenes, overlapped with the other signals), 7.07–6.98 (m, 1H, free Lutidine), 6.73–6.64 (m, 1H, coordinated Lutidine), 6.61–6.55 (m, 2H, free Lutidine), 6.21–6.14 (m, 2H, coordinated Lutidine), 2.41 (s, 3H, CH₃ in free Lutidine), 2.25 (s, 3H, CH₃ in coordinated Lutidine), 2.12 (s, 3H, CH₃ in partially deuterated toluenes). **¹⁹F NMR (282.4 MHz, C₆D₆) δ:** –122.6 to –122.9 (m, 2F), –152.1 to –152.6 (m, 1F), –160.7 to –161.2 (m, 2F). **¹³C{¹H} NMR (75.4 MHz, C₆D₆) δ:** 160.2 (s, C in Lutidine), 158.0 (s, C in Lutidine), 151.9–147.9 (m, CF in C₆F₅), 144.1–140.0 (m, CF in C₆F₅), 142.1 (s, CH in coordinated Lutidine), 139.5–135.3 (m, CF in C₆F₅), 136.1 (s, CH in free Lutidine), 129.3 (s, CH in toluene), 128.6 (s, CH in toluene), 125.7 (s, CH in toluene), 125.4 (s, CH in free Lutidine), 119.9 (s, Ch in coordinated Lutidine), 24.5 (s, CH₃ in free Lutidine), 24.2 (s, CH₃ in coordinated Lutidine), 21.4 (s, CH₃ in partially deuterated toluenes) (Al-C missed due to low intensity). **²⁷Al NMR (78.2 MHz, C₆D₆) δ:** 117±1 (*W*_{1/2} 3.6 kHz).

Al(C₆F₅)₃·MeCN complex

¹H NMR (300.1 MHz, C₆D₆) δ: 7.13–.08 (m, CH in partially deuterated toluene), 7.07–6.97 (m, CH in partially deuterated toluene), 2.11 (s, 3H, CH₃ in partially deuterated toluenes), 0.56–0.49 (m, 3H, CH₃CN, mixture of coordinated and free). **¹⁹F NMR (282.4 MHz, C₆D₆) δ:** –123.2 to –123.6 (m, 2F), –152.9 to –153.6 (m), 1F, –161.4 to –162.1 (m, 2F). **²⁷Al NMR (78.2 MHz, C₆D₆) δ:** 86±5 (*W*_{1/2} 12 kHz).

Al(C₆F₅)₃·THF complex

¹H NMR (300.1 MHz, C₆D₆) δ: 7.14–7.06 (m, CH in partially deuterated toluene), 7.06–6.96 (m, CH in partially deuterated toluene), 3.59–3.49 (m, 4H, THF), 2.11 (s, 3H, CH₃ in partially deuterated toluenes), 1.50–1.39 (m, 4H, THF) [*Coordinated and non-coordinated THF are in a fast exchange and give one set of signals*]. **¹⁹F NMR (282.4 MHz, C₆D₆) δ:** –122.8 to –123.1 (m, 2F), –151.9 (t, *J* = 19.5 Hz, 1F), –161.1 to –161.4 (m, 2F). **²⁷Al NMR (78.2 MHz, C₆D₆) δ:** 129±2 (*W*_{1/2} 4.5 kHz).

Al(C₆F₅)₃·Me₂S complex

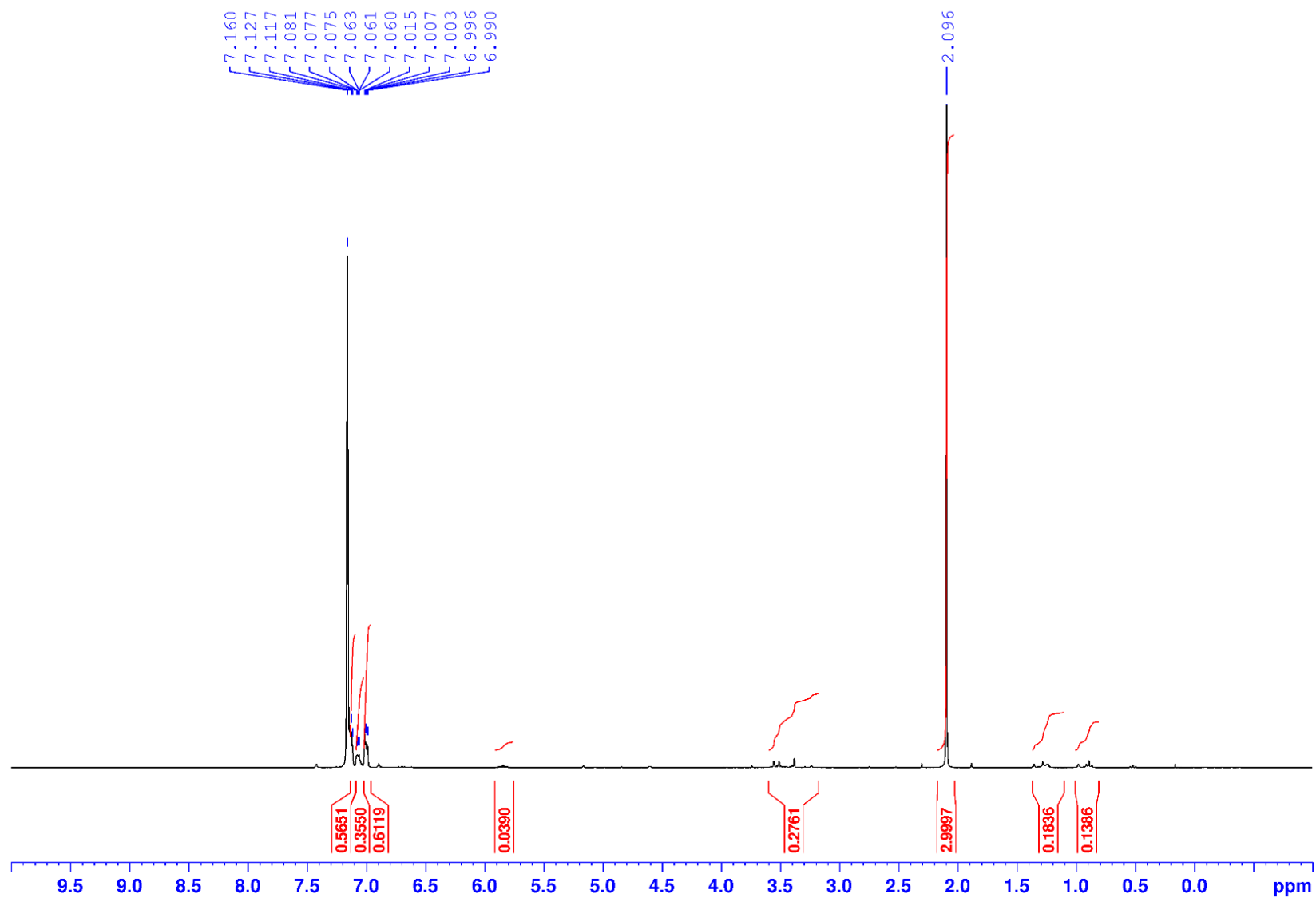
¹H NMR (300.1 MHz, C₆D₆) δ: 7.14–7.08 (m, CH in partially deuterated toluene), 7.07–6.97 (m, CH in partially deuterated toluene), 2.11 (s, 3H, CH₃ in partially deuterated toluenes), 1.57–1.51 (m, 6H, Me₂S, mixture of coordinated and free). **¹⁹F NMR (282.4 MHz, C₆D₆) δ:** –121.5 to –121.7 (m, 2F), –150.2 (t, *J* = 19.8 Hz, 1F), –160.0 to –160.4 (m, 2F). **²⁷Al NMR (78.2 MHz, C₆D₆) δ:** 135±2 (*W*_{1/2} 5.5 kHz).

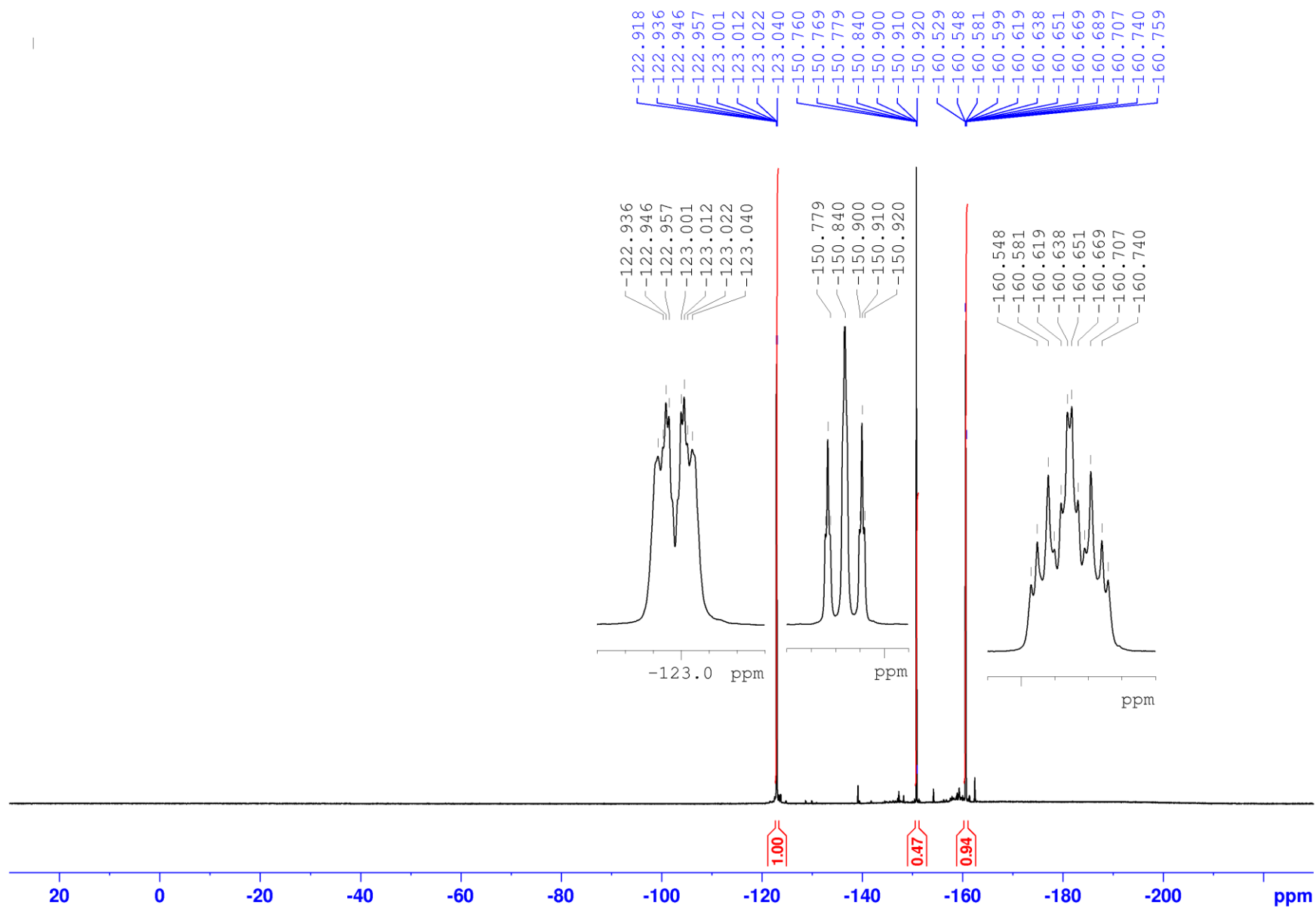
Al(C₆F₅)₃·P(OEt)₃ complex

¹H NMR (300.1 MHz, C₆D₆) δ: 7.14–7.07 (m, CH in partially deuterated toluene), 7.07–6.96 (m, CH in partially deuterated toluene), 3.78 (dq, *J* = 8.0, 7.1 Hz, 6H, CH₂ in P(OEt)₃), 2.11 (s, 3H, CH₃ in partially deuterated toluenes), 1.09 (t, *J* = 7.0 Hz, 9H, CH₃ in P(OEt)₃) [*Coordinated and non-coordinated P(OEt)₃ are in a fast exchange and give one set of signals*]. **¹⁹F NMR (282.4 MHz, C₆D₆) δ:** [*in ¹⁹F NMR spectra several types of C₆F₅-groups are presented that could be attributed to different complexes existing in dynamic equilibrium*] –123.4 to –123.7 (m, 2F, complex 1), –123.7 to –123.9 (m, 2F, complex 2), –153.0 (t, *J* = 19.7 Hz, complex 2), –153.2 (t, *J* = 19.6 Hz, complex 1), –161.4 – -161.9 (m, 2F, both complexes). **³¹P{¹H} NMR (121.5 MHz, C₆D₆) δ:** 138.6 (s) [*Coordinated and non-coordinated P(OEt)₃ are in a fast exchange and give one set of signals*]. **²⁷Al NMR (78.2 MHz, C₆D₆) δ:** 110±2 (*W*_{1/2} 4.0 kHz).

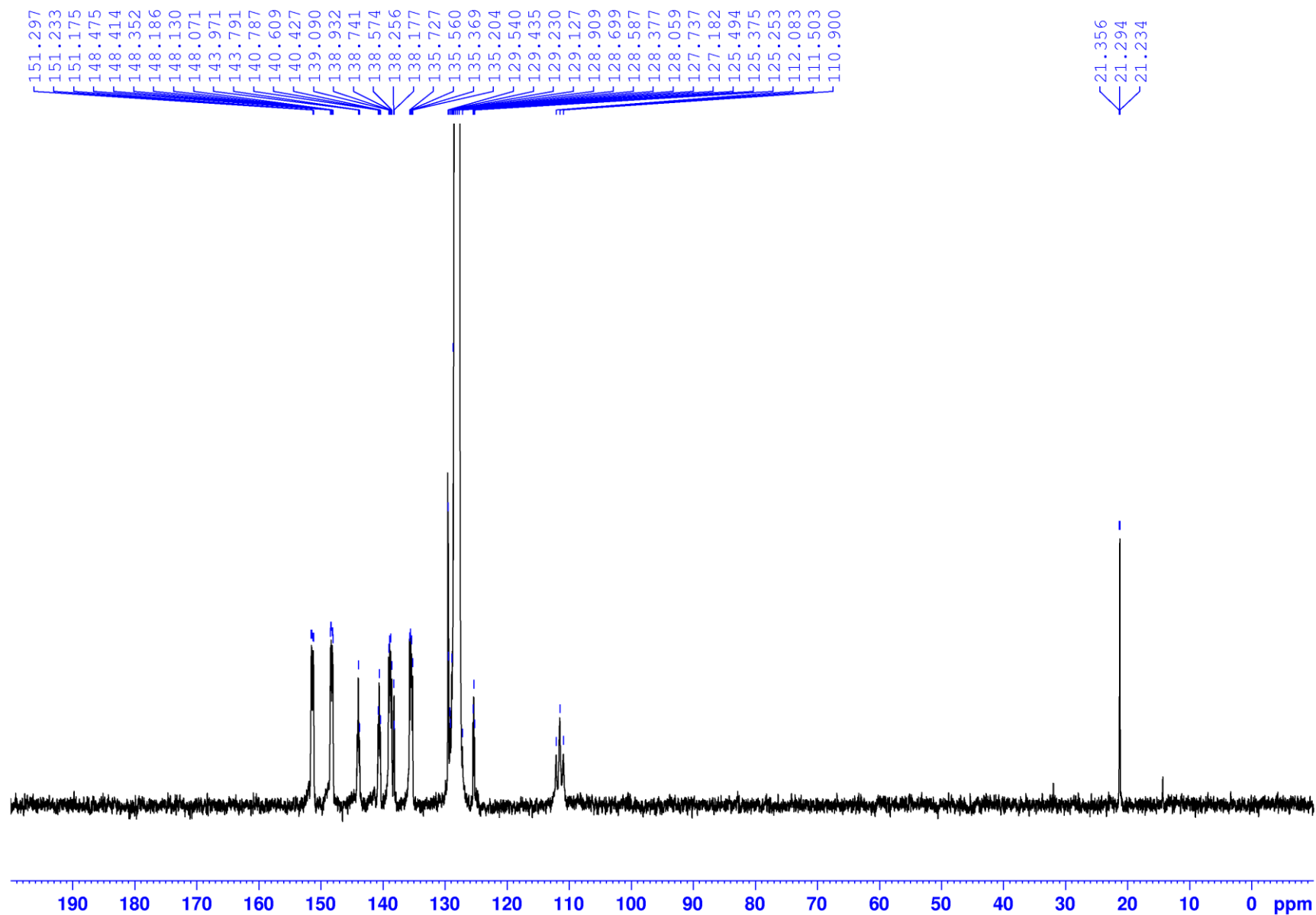
3. Copies of ^1H , ^{19}F , ^{13}C , ^{27}Al -NMR spectra and 2D correlations

^1H NMR (300.1 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 0.5\text{toluene}$

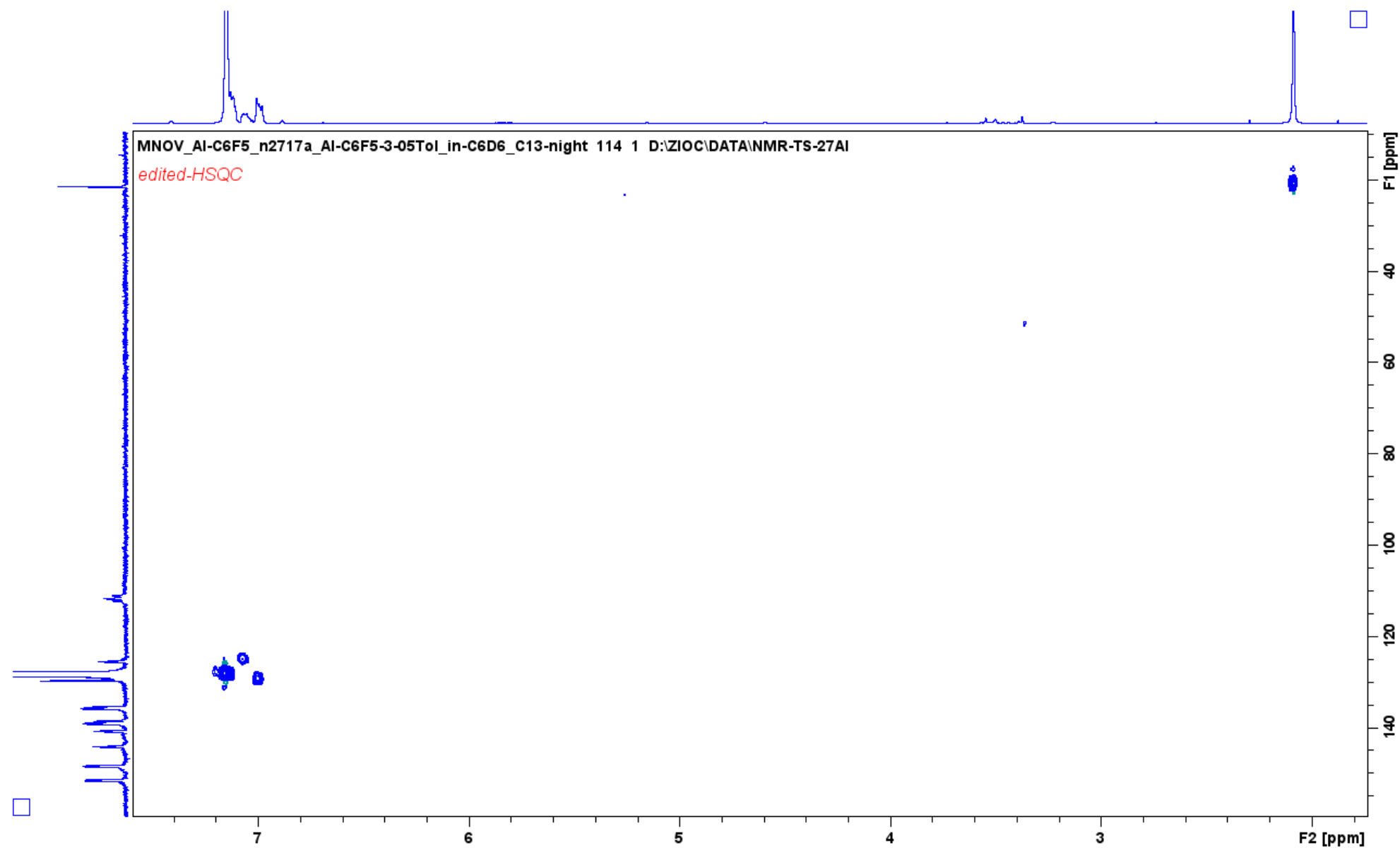


^{19}F NMR (282.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 0.5\text{toluene}$ 

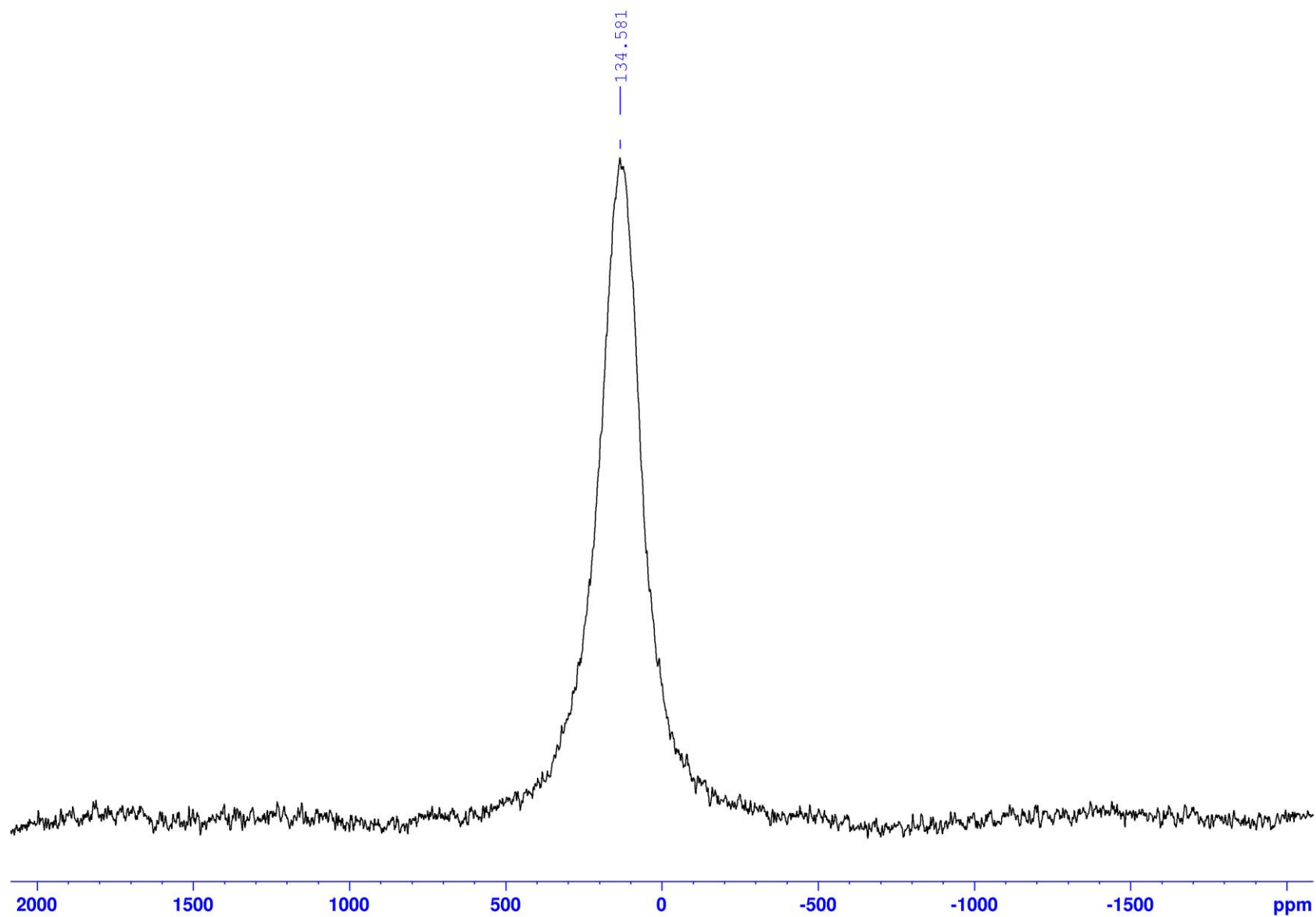
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 0.5\text{toluene}$

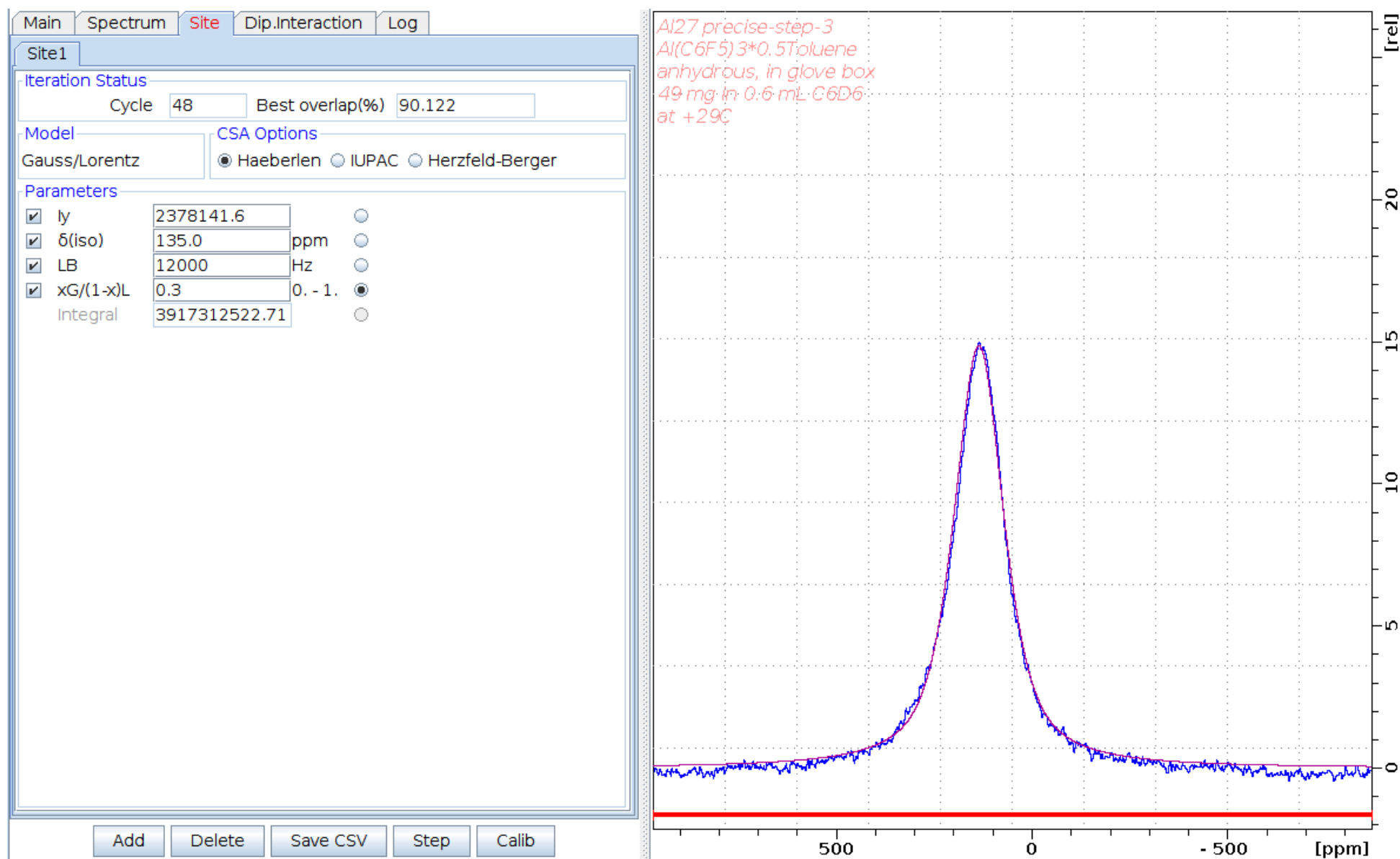


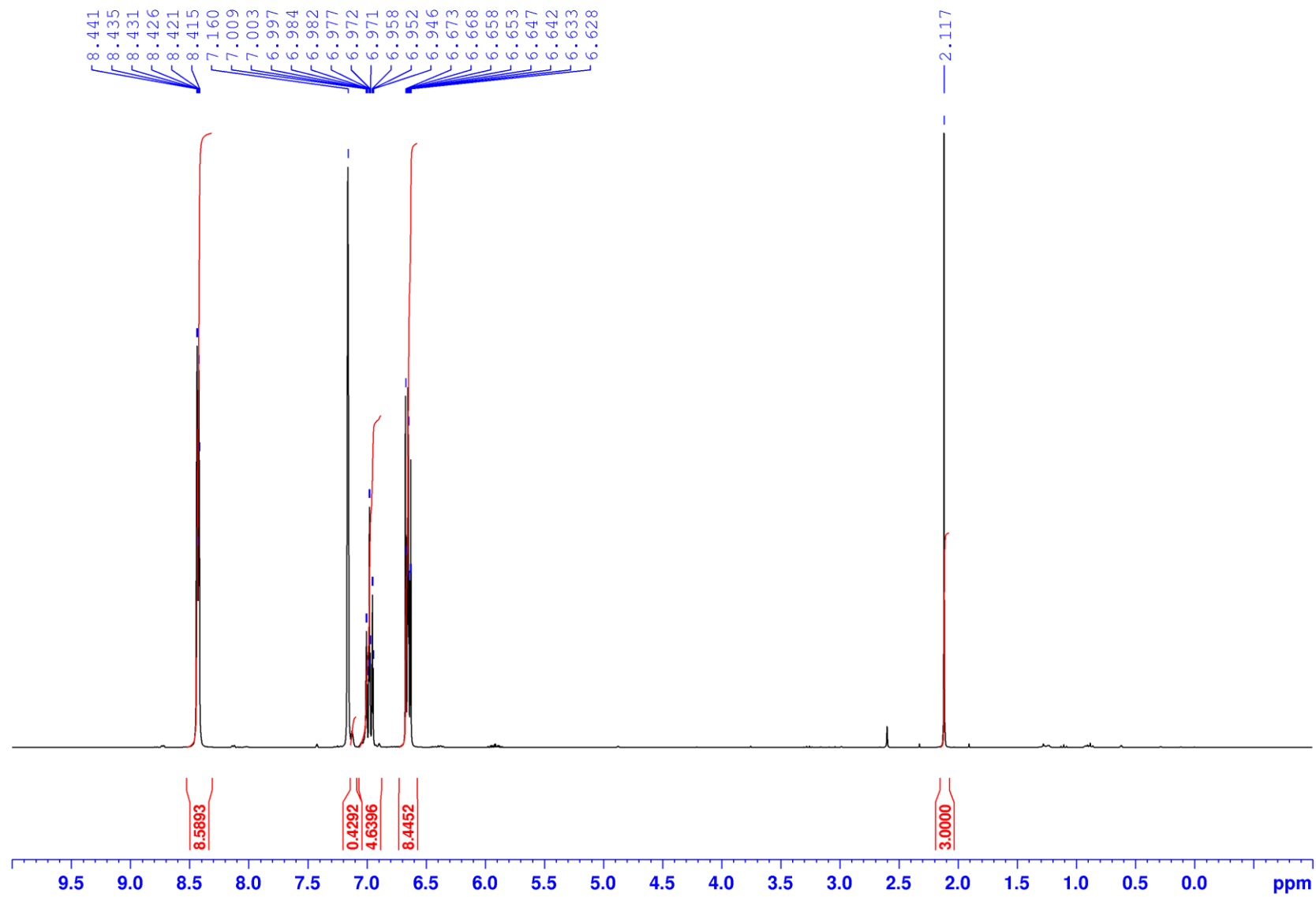
$\{^1\text{H}, ^{13}\text{C}\}$ -edited HSQC (C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 0.5\text{toluene}$

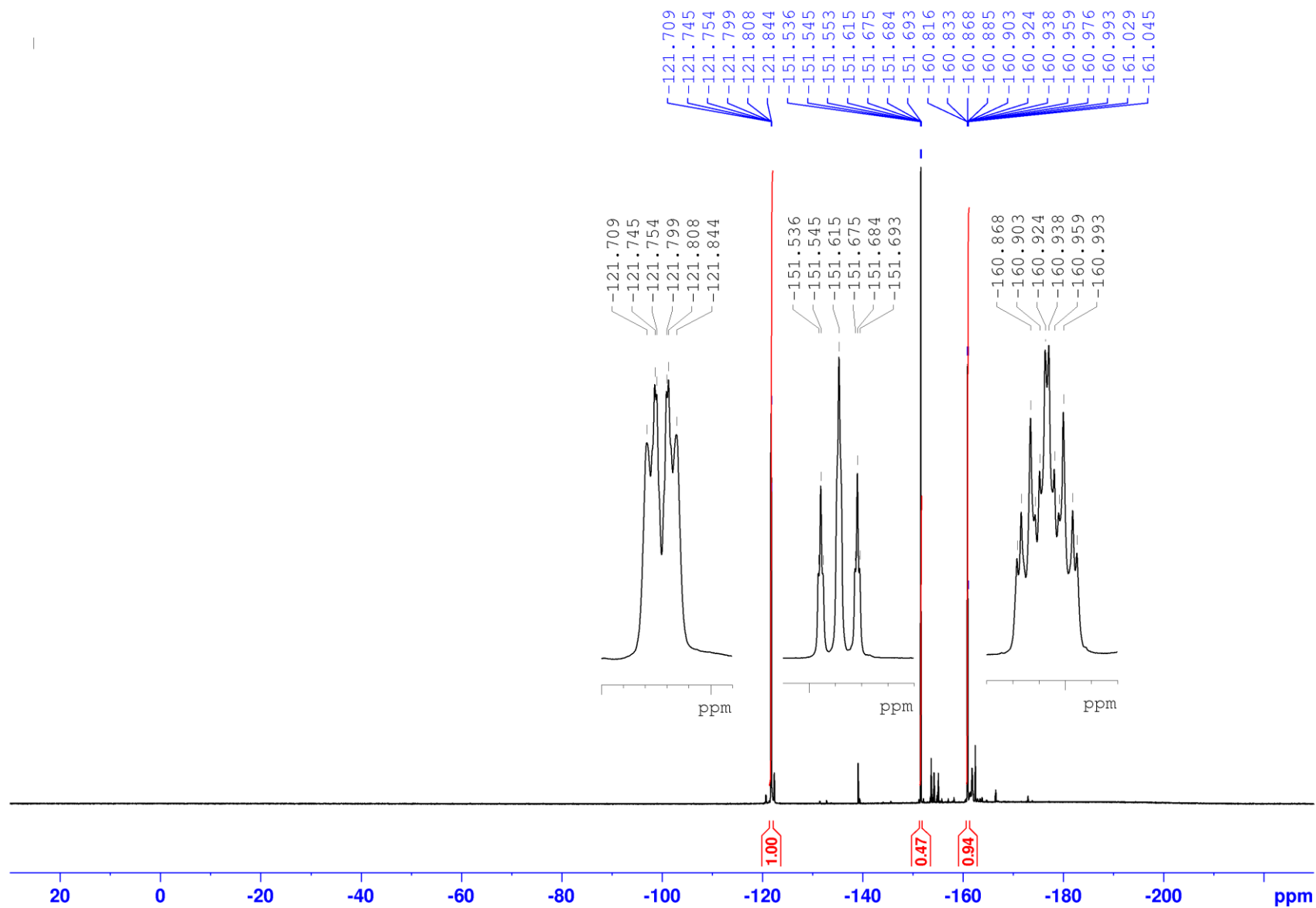


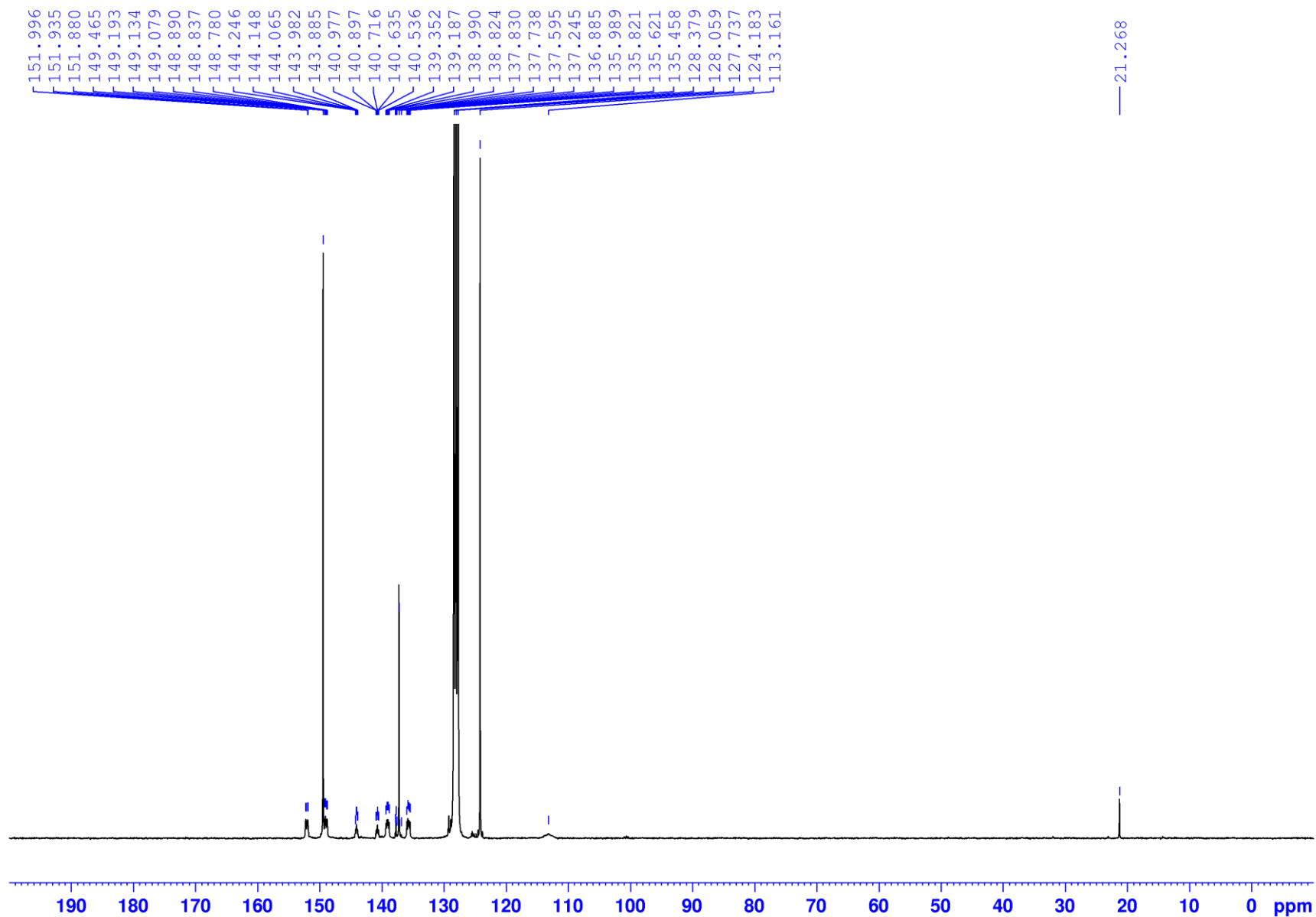
^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 0.5\text{toluene}$



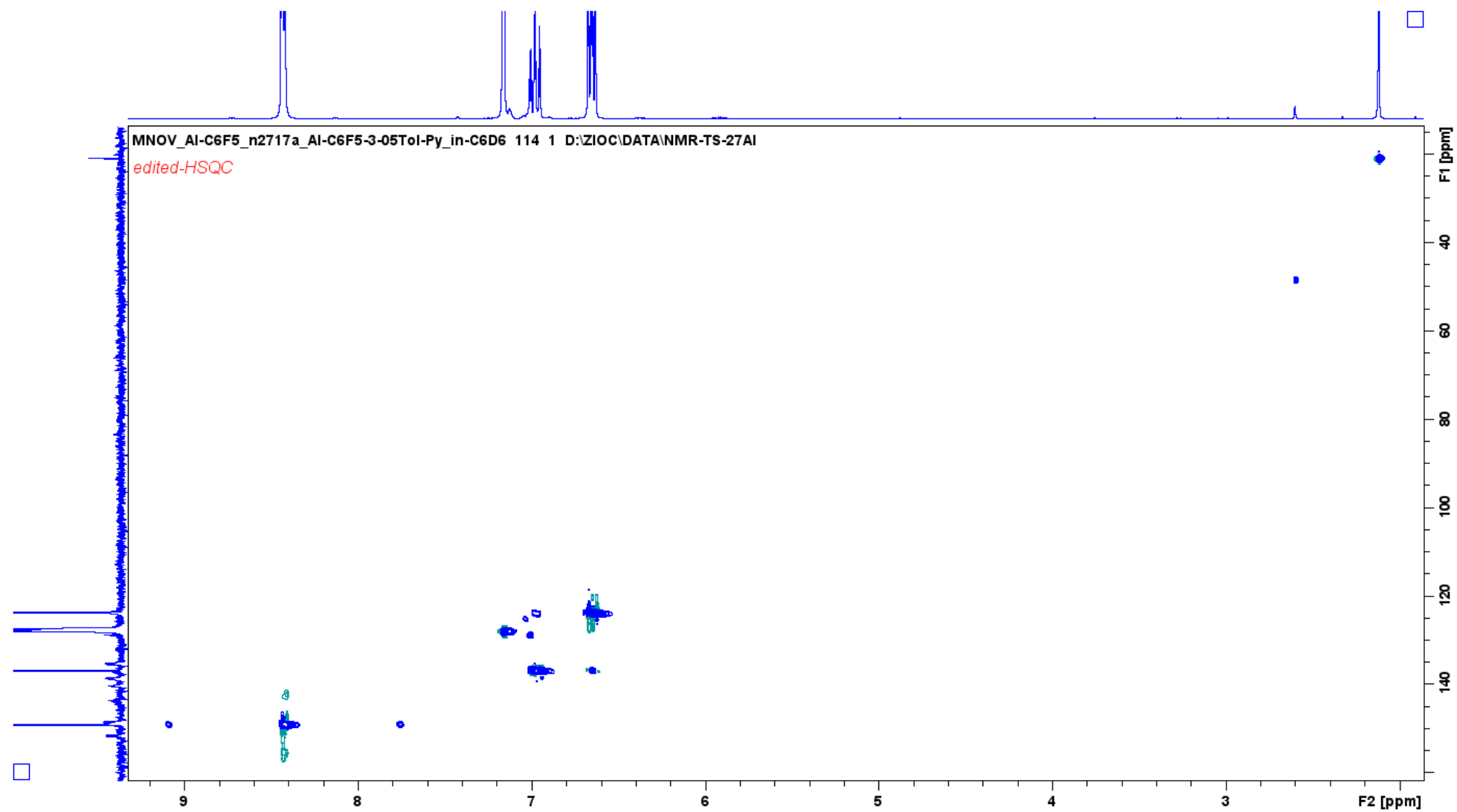
^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 0.5\text{toluene}$ — Line shape analysis

^1H NMR (300.1 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{Pyridine}$ complex

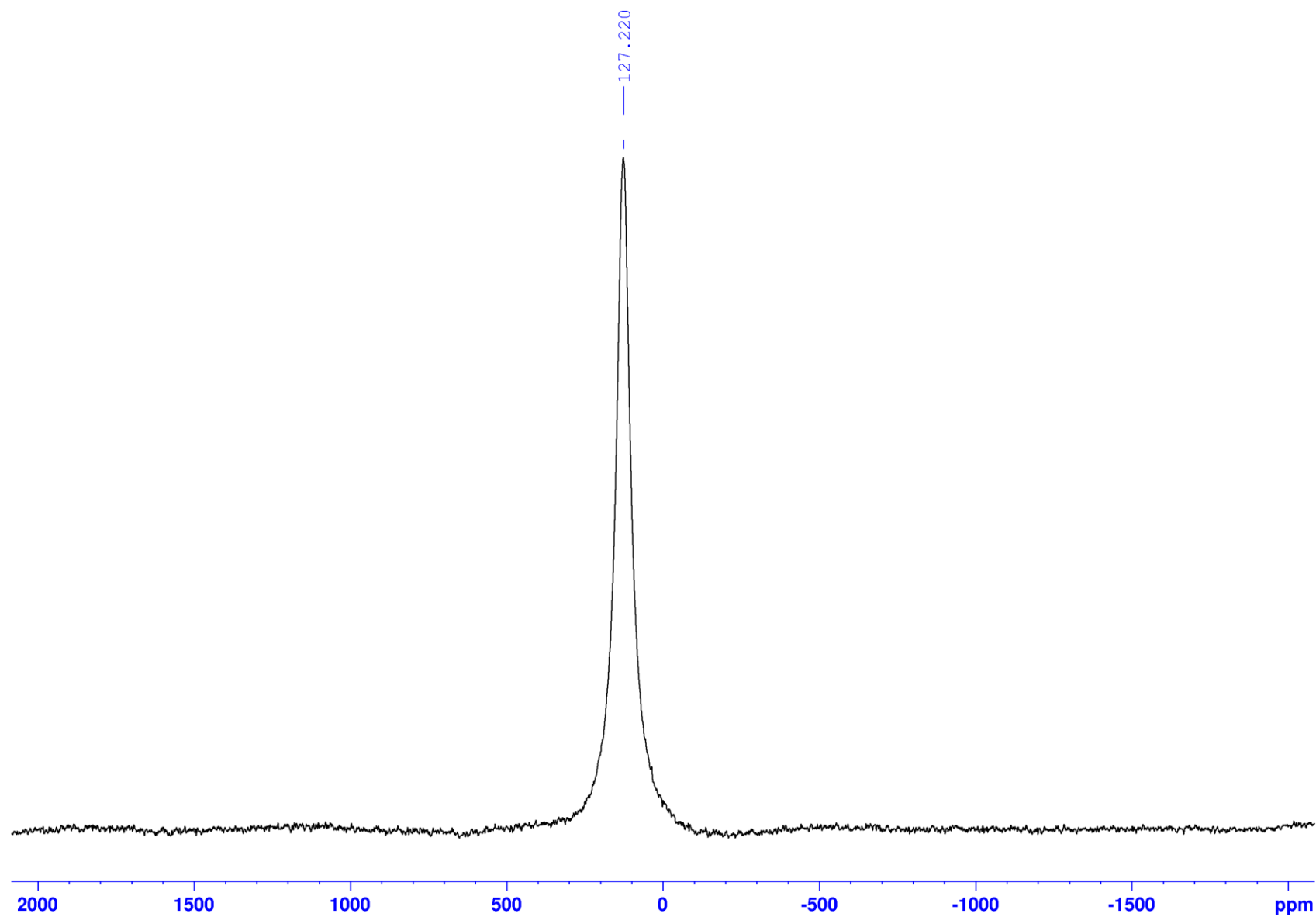
^{19}F NMR (282.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{Pyridine}$ complex

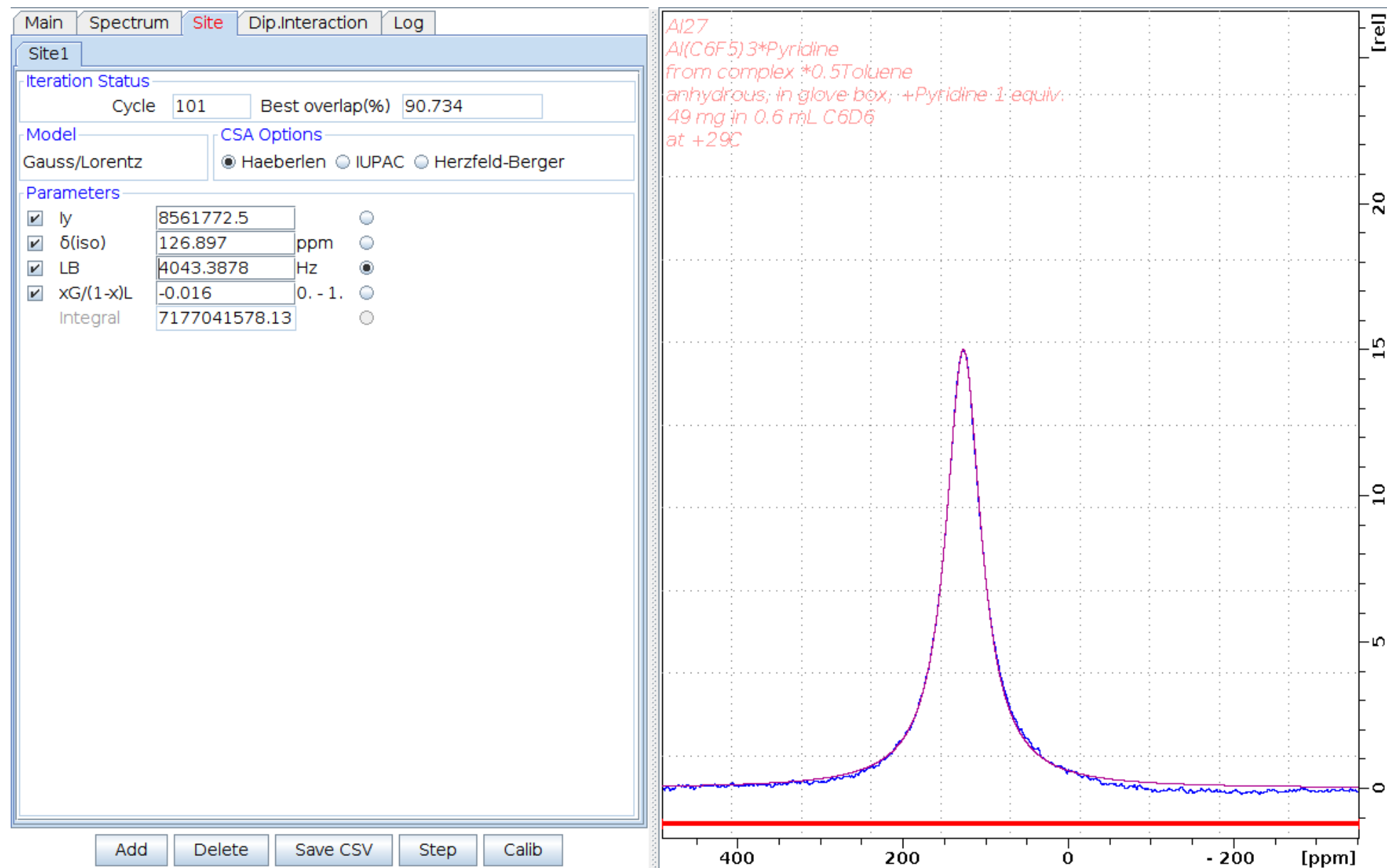
$^{13}\text{C}\{^1\text{H}\}$ NMR (75.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{Pyridine}$ complex

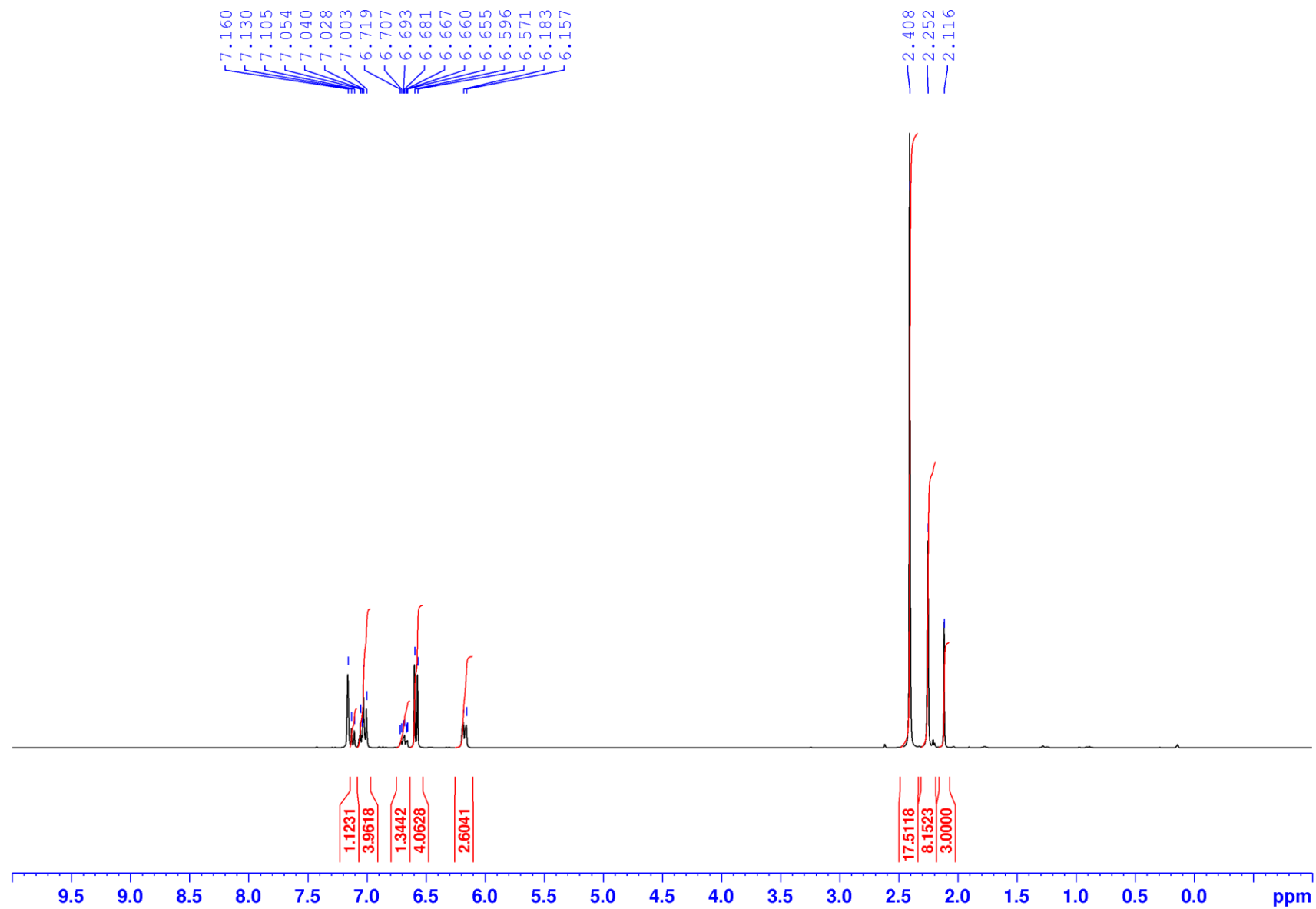
$\{^1\text{H}, ^{13}\text{C}\}$ -edited HSQC (C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{Pyridine}$ complex

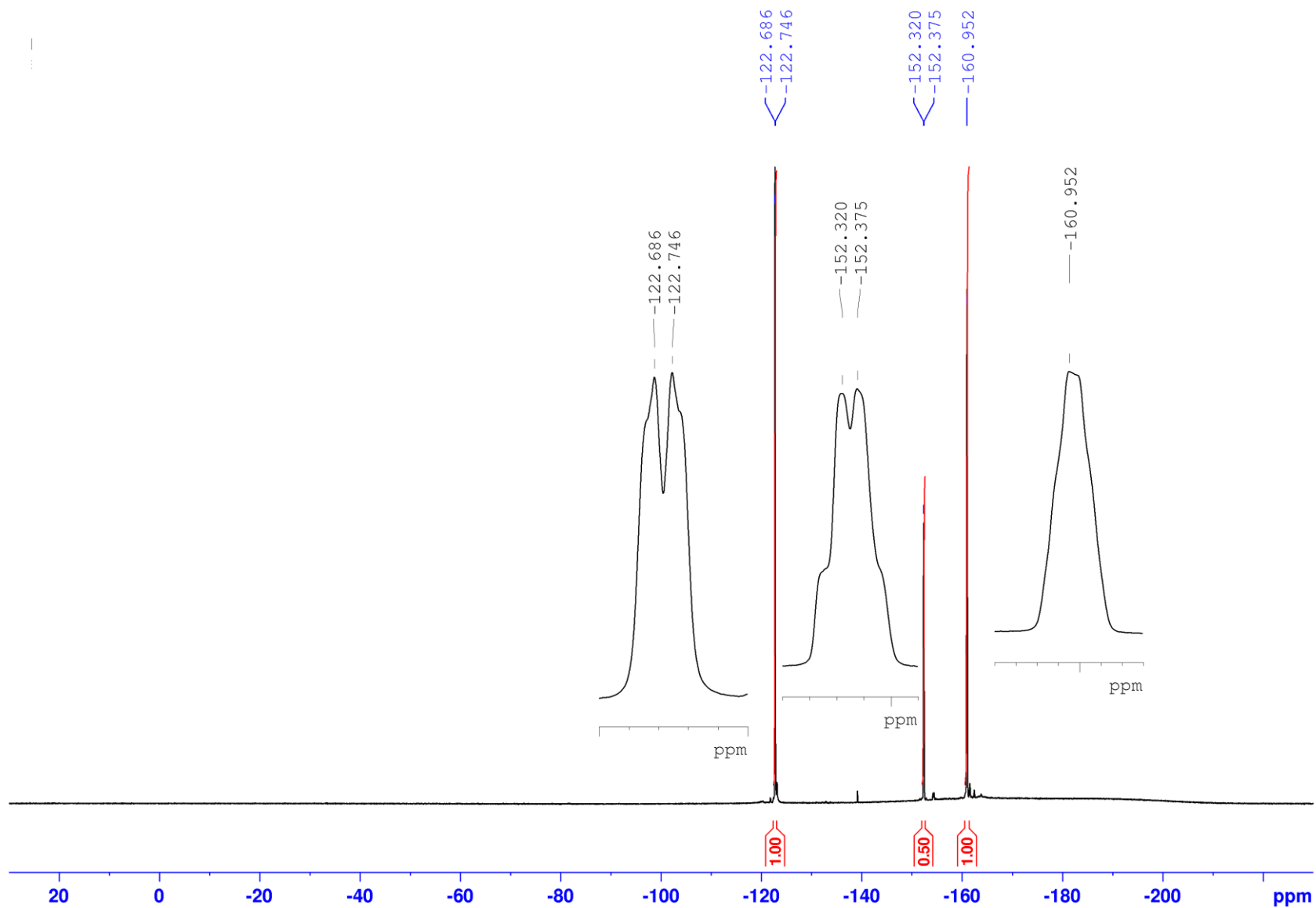


^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{Pyridine}$ complex

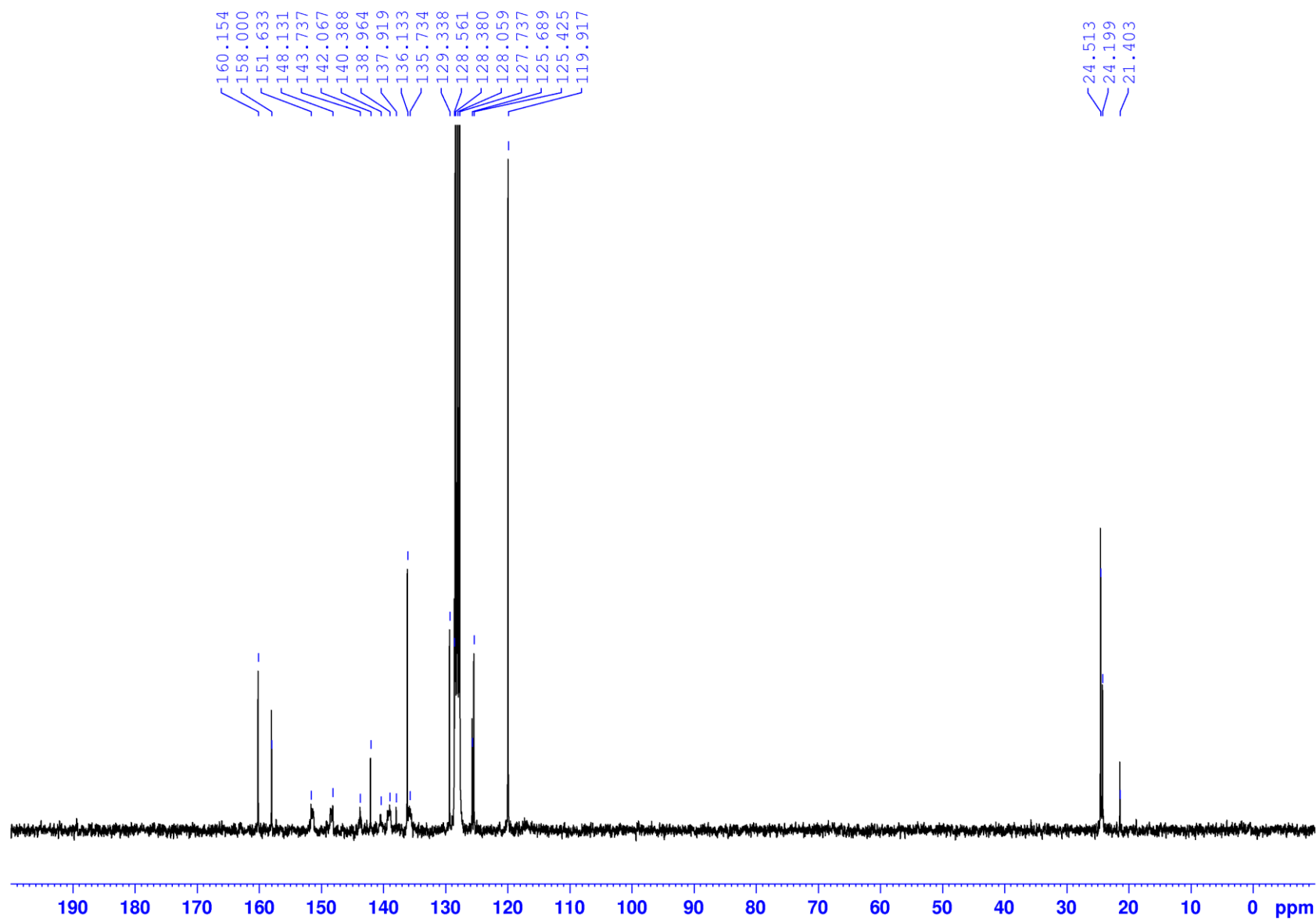


^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3\cdot\text{Pyridine}$ complex — Line shape analysis

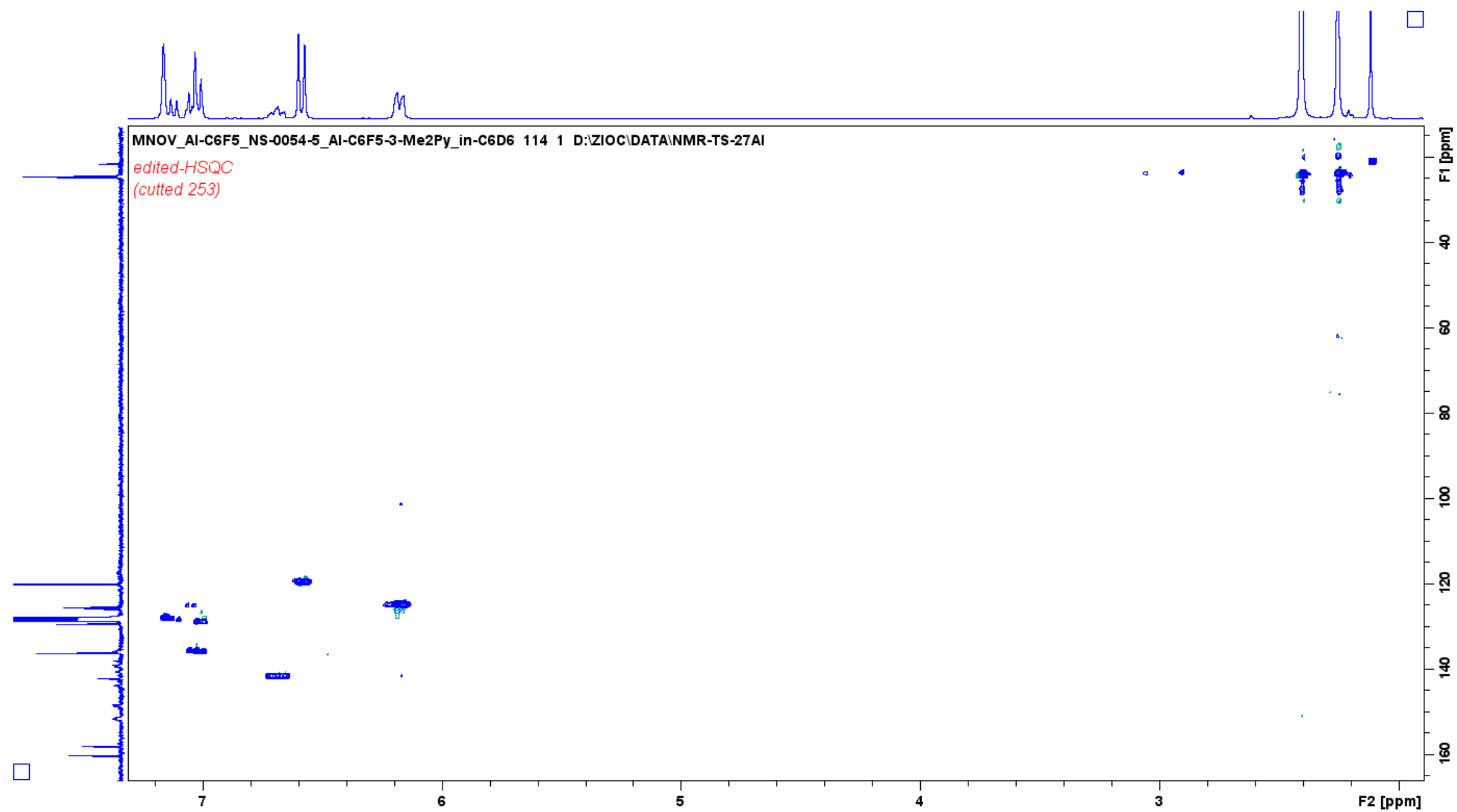
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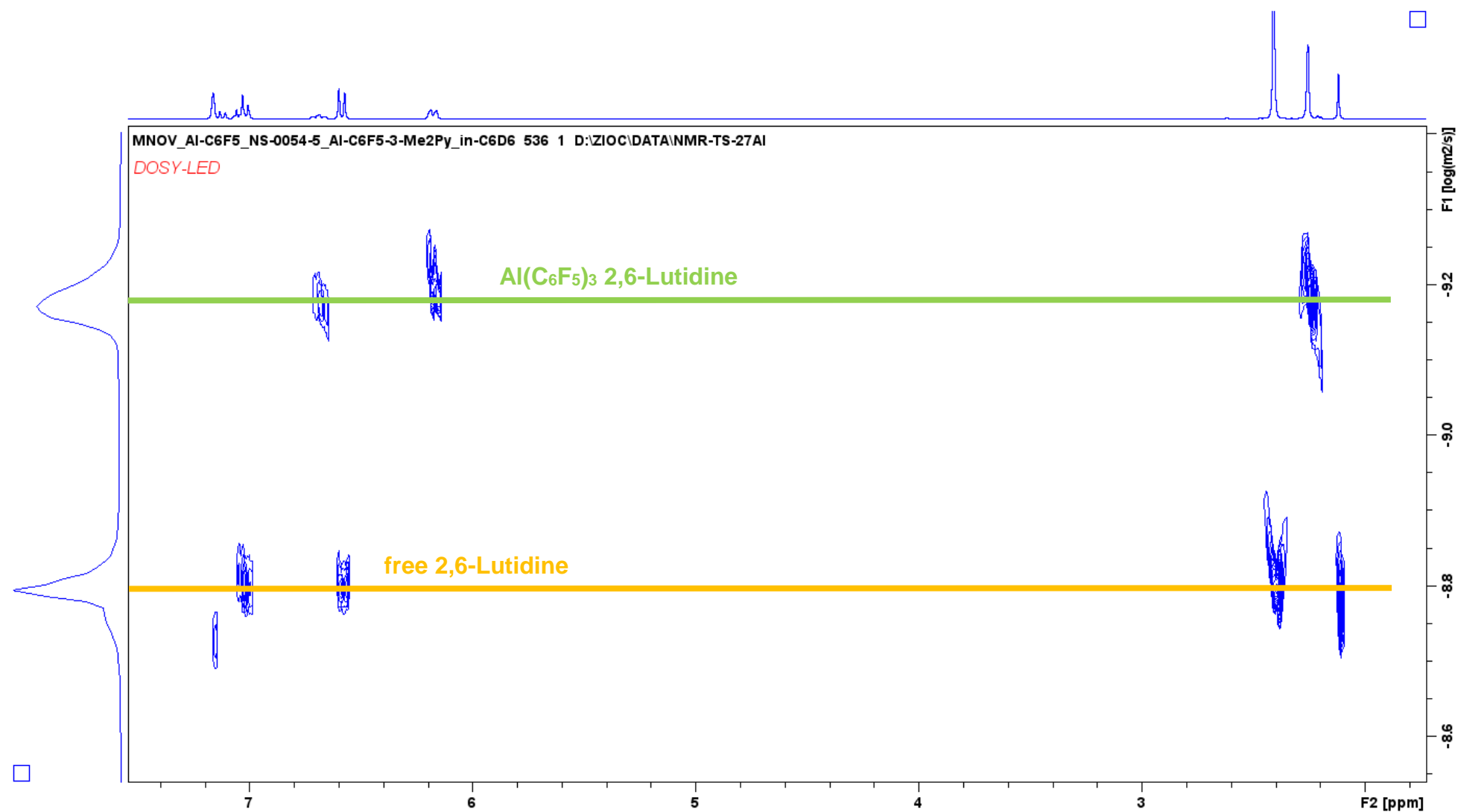
^{19}F NMR (282.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 2,6\text{-Lutidine}$ complex

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 2,6\text{-Lutidine}$ complex

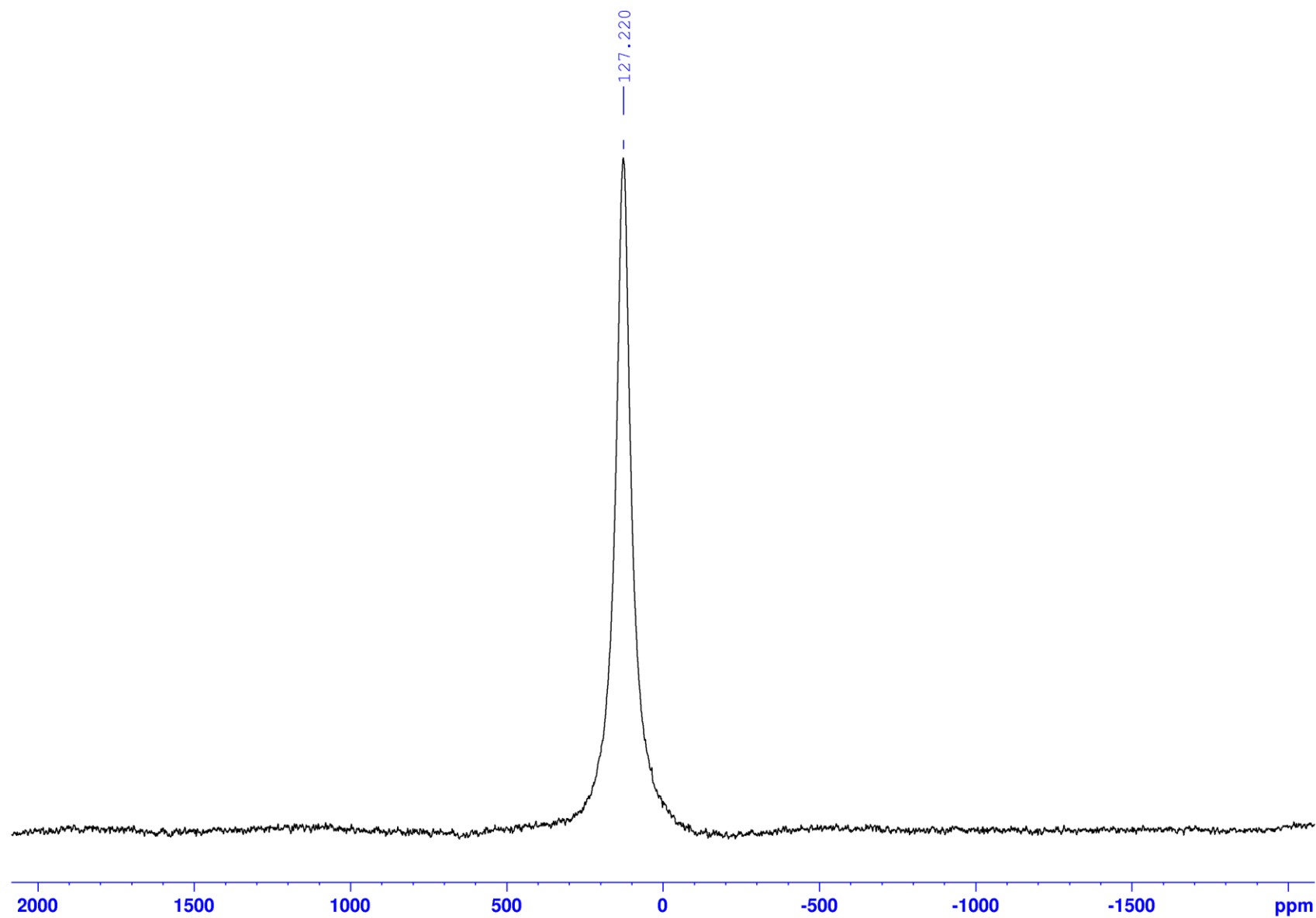


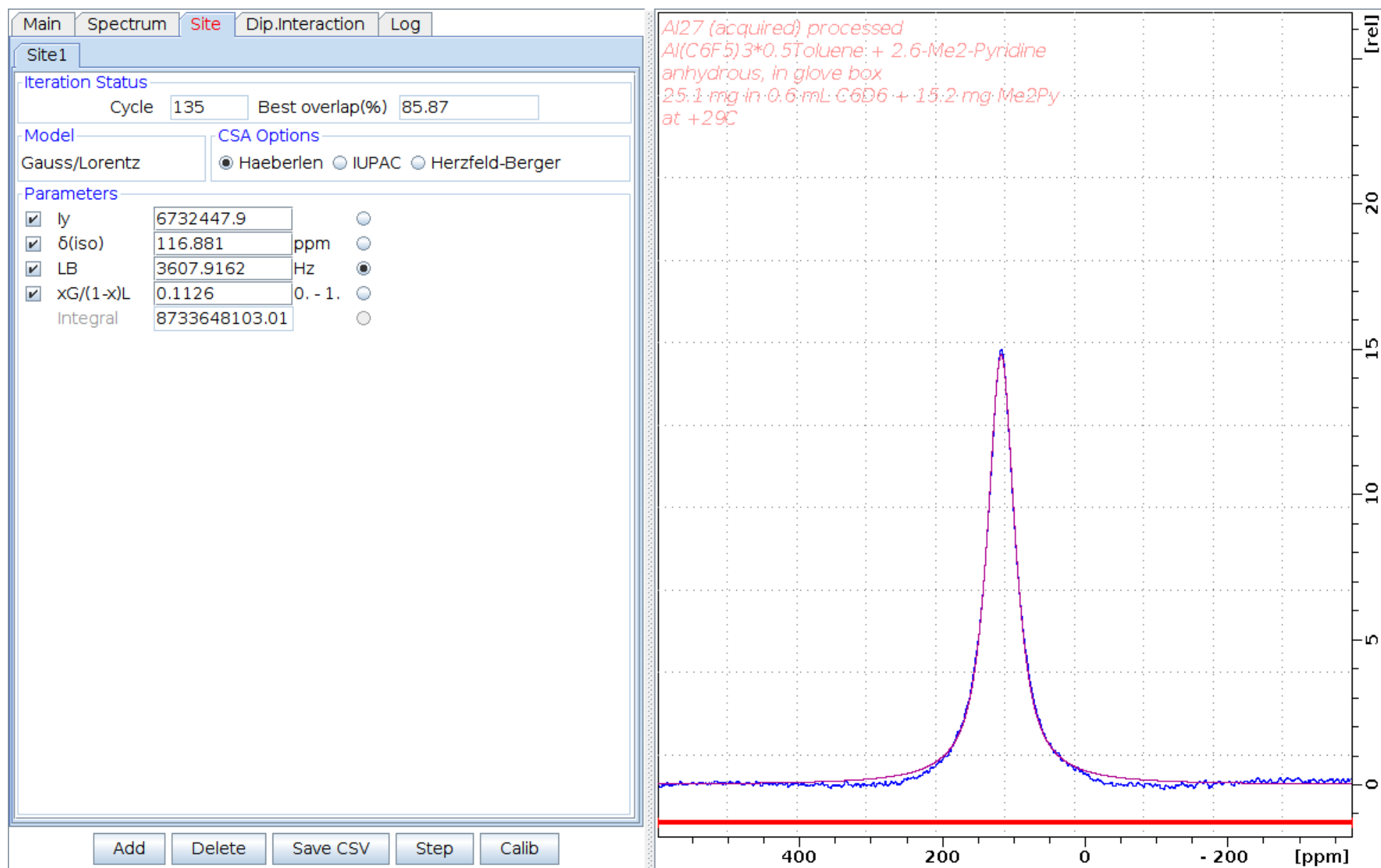
$\{^1\text{H}, ^{13}\text{C}\}$ -edited HSQC (C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 2,6\text{-Lutidine}$ complex

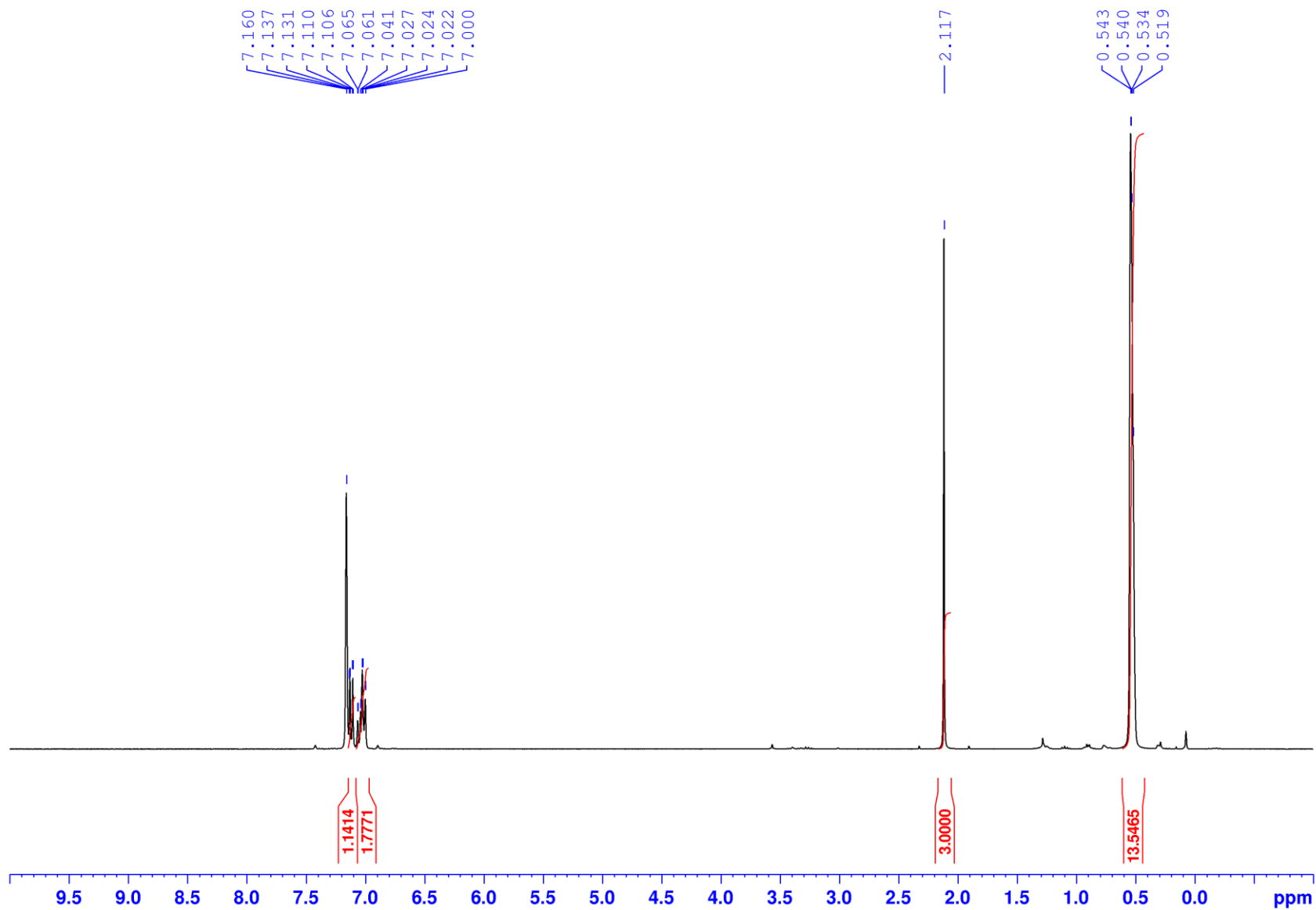


$\{^1\text{H}\}$ -DOSY (C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 2,6\text{-Lutidine}$ complex

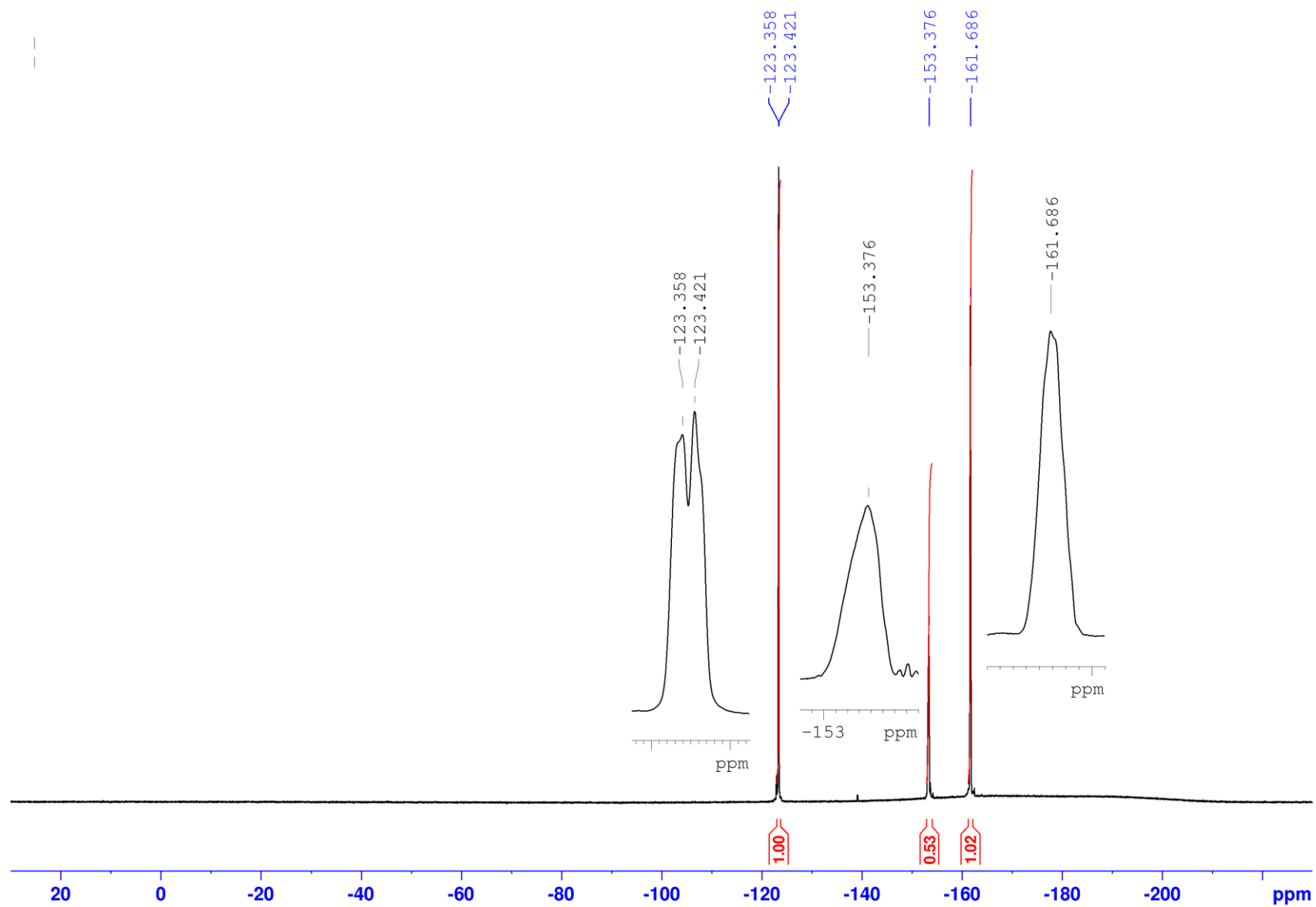
^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 2,6\text{-Lutidine}$ complex



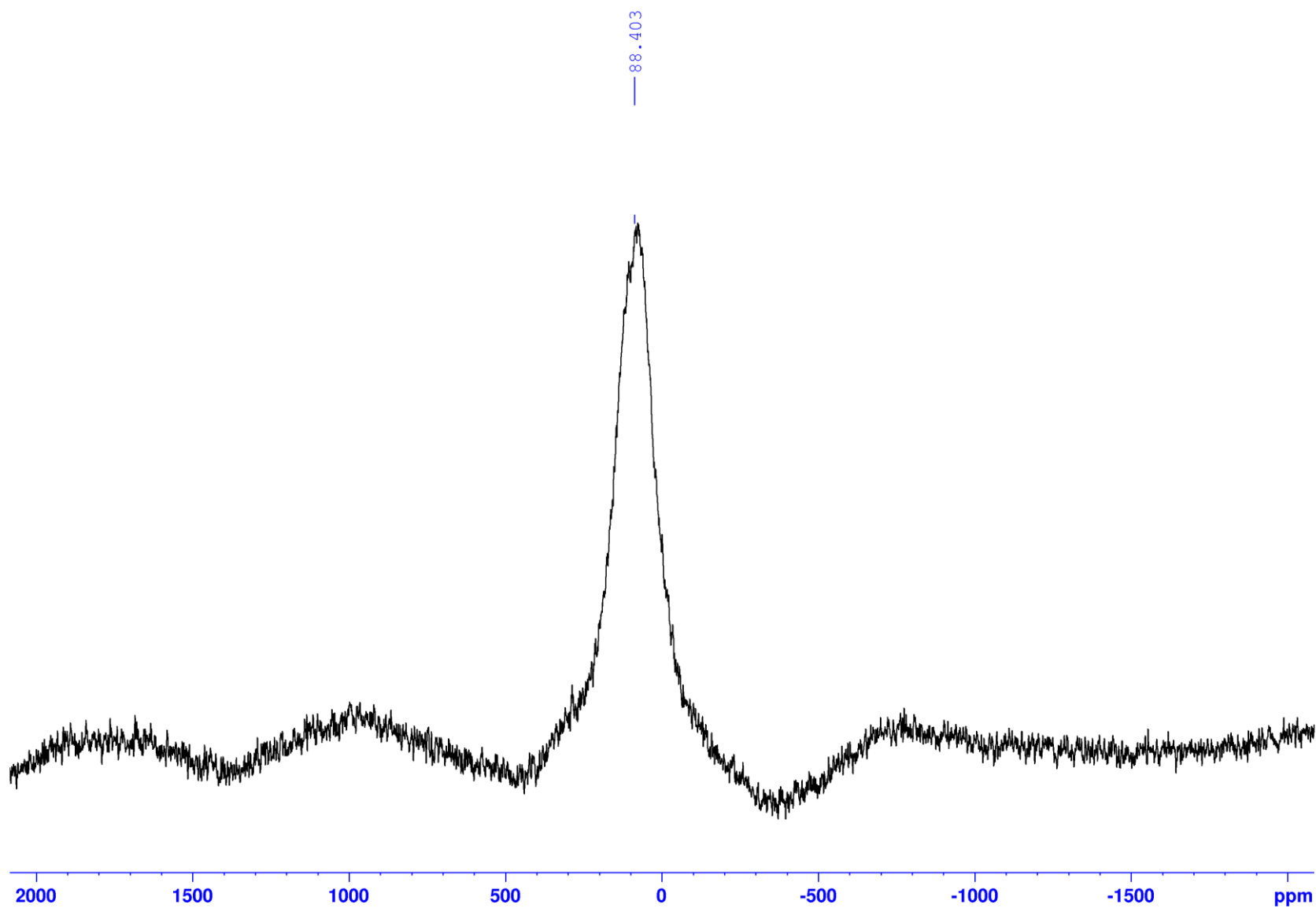
^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot 2,6\text{-LuTidine}$ complex — Line shape analysis

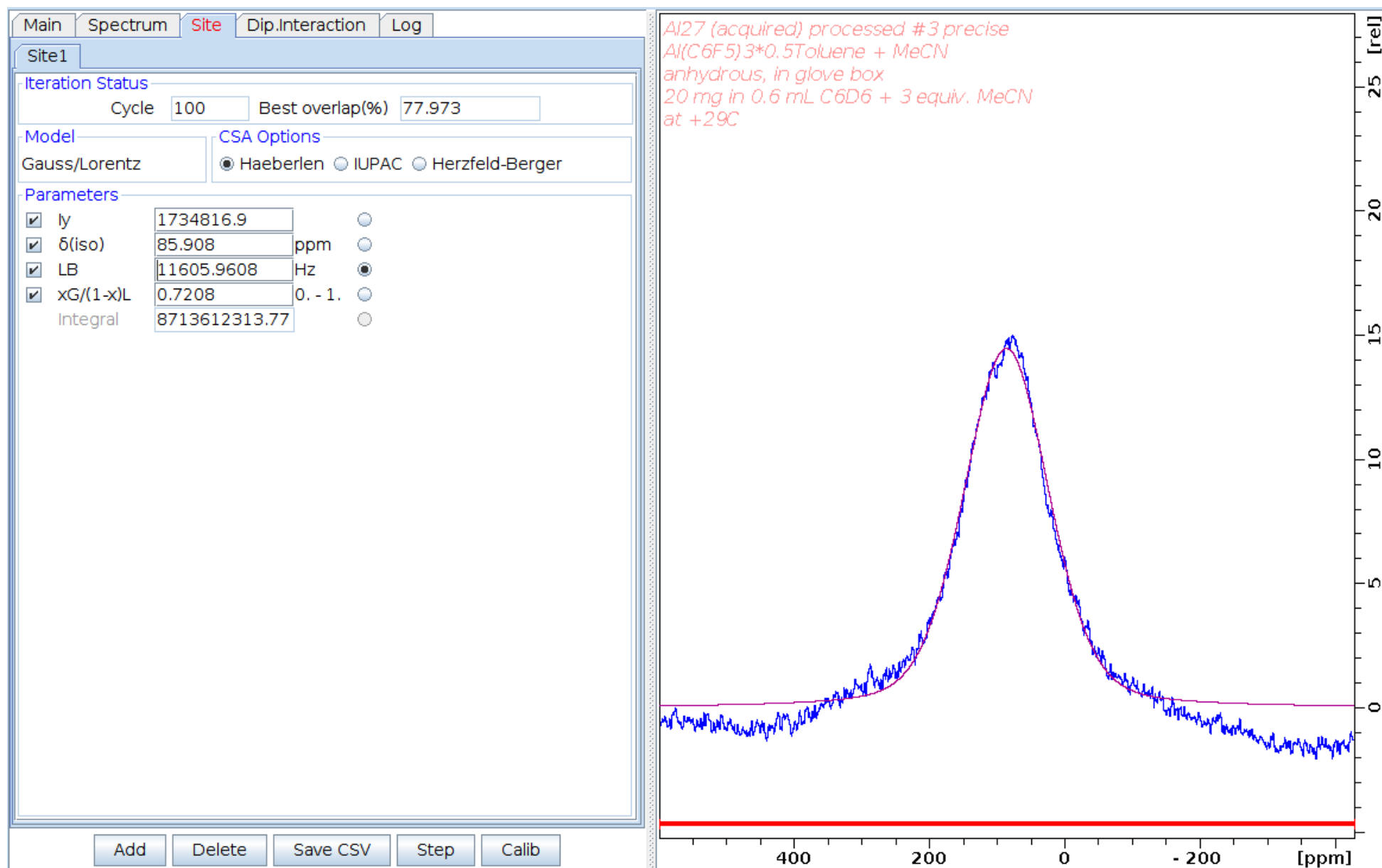
^1H NMR (300.1 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{MeCN}$ complex

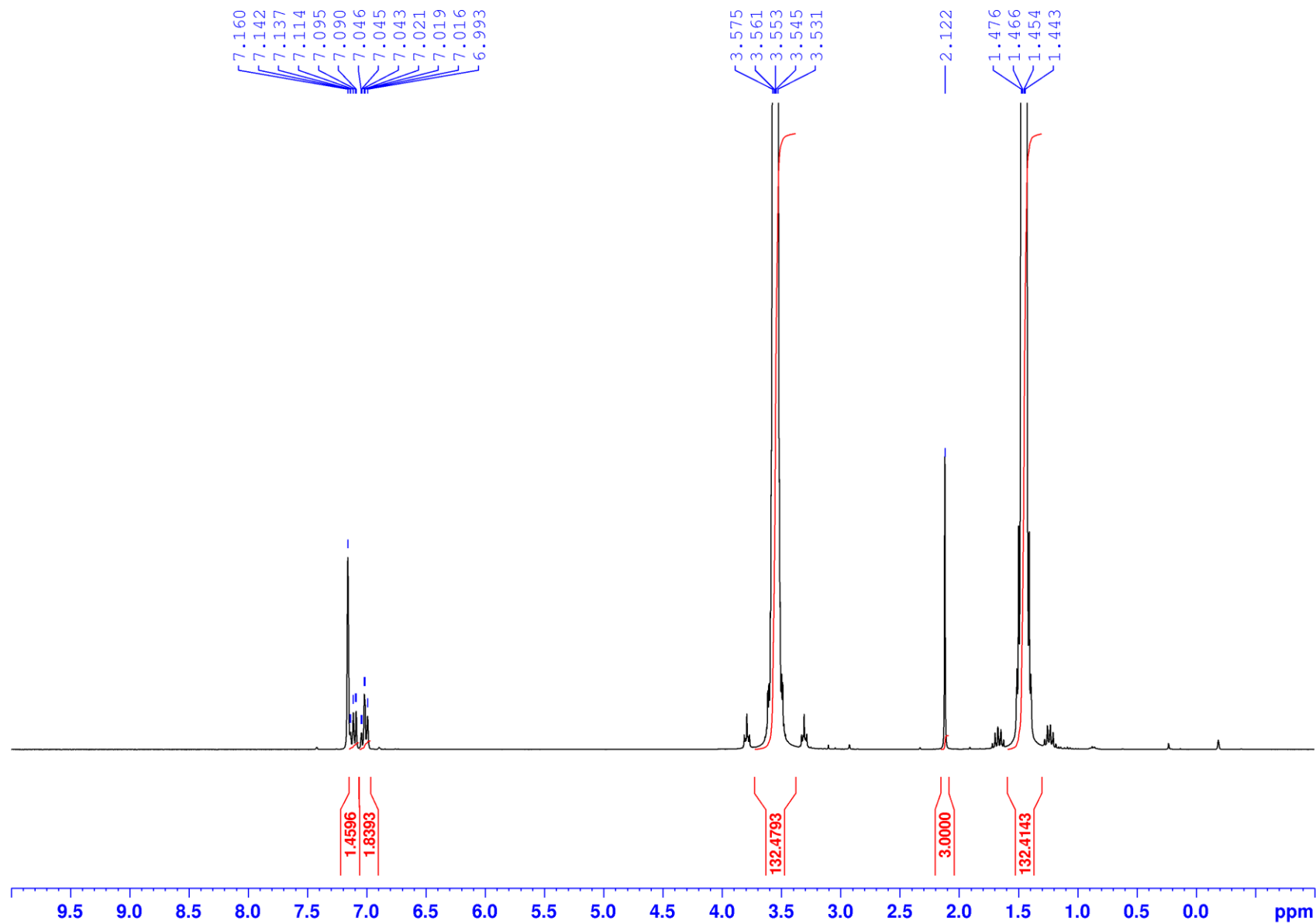
^{19}F NMR (282.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{MeCN}$ complex

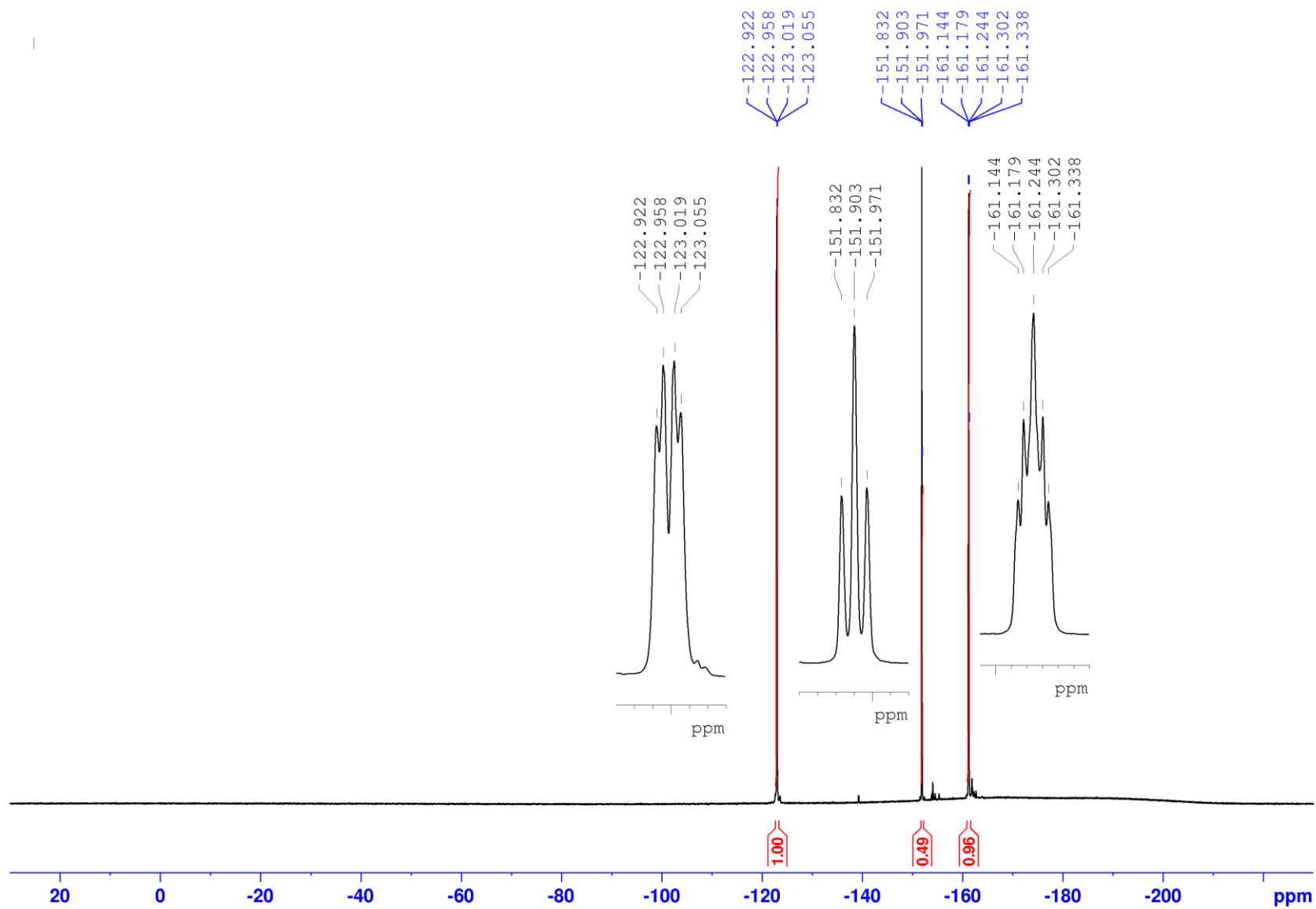


^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{MeCN}$ complex

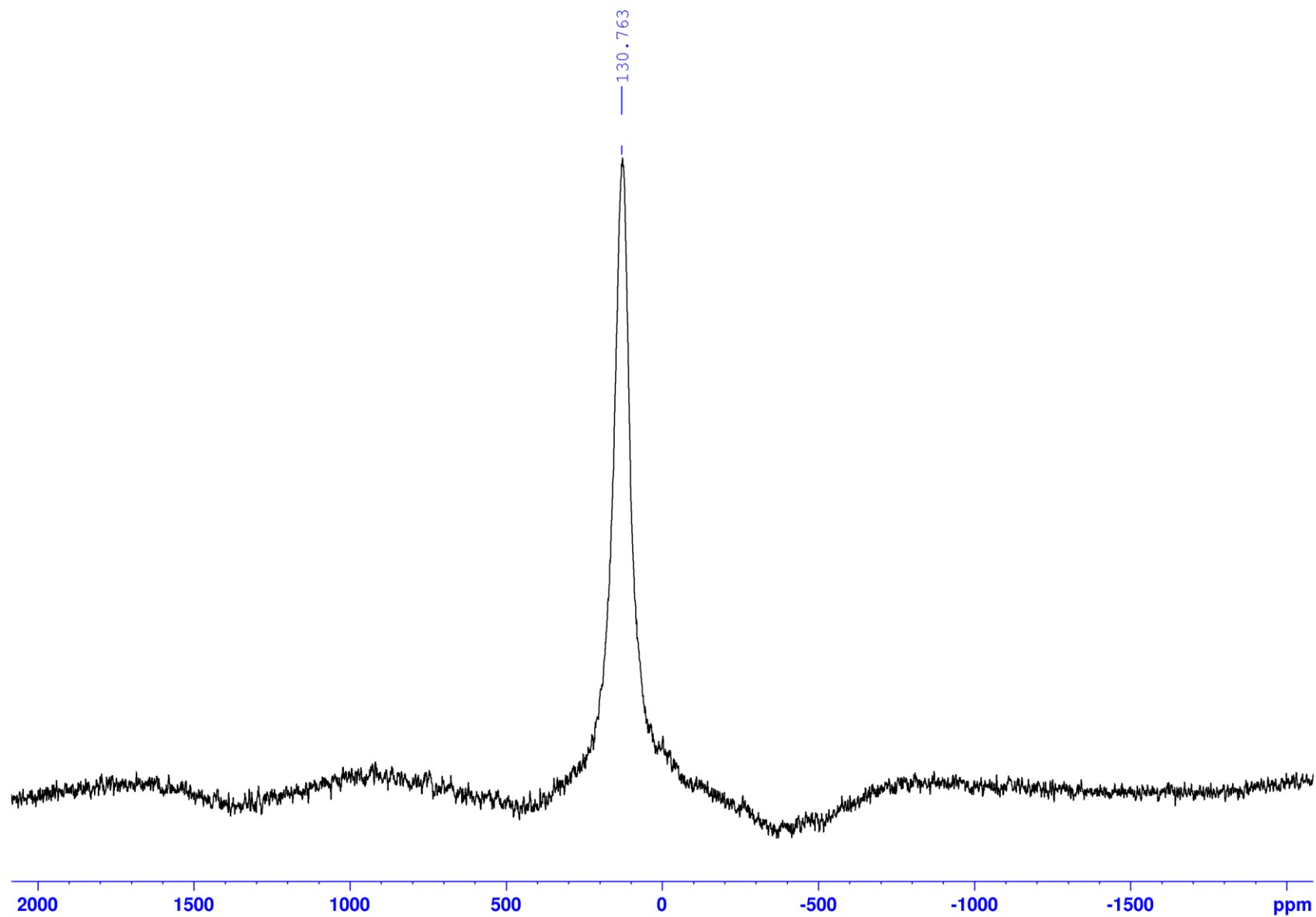


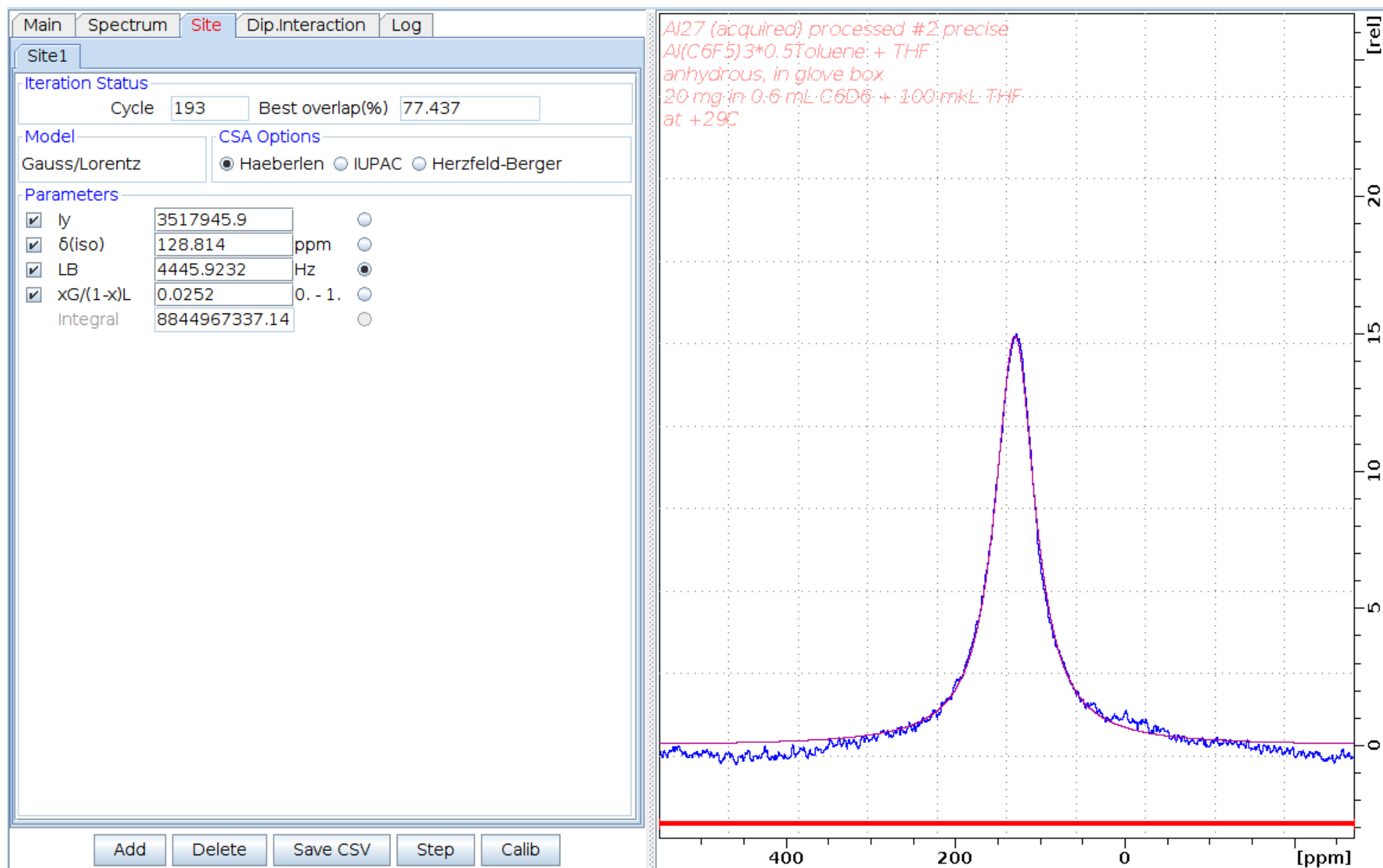
^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{MeCN}$ complex — Line shape analysis

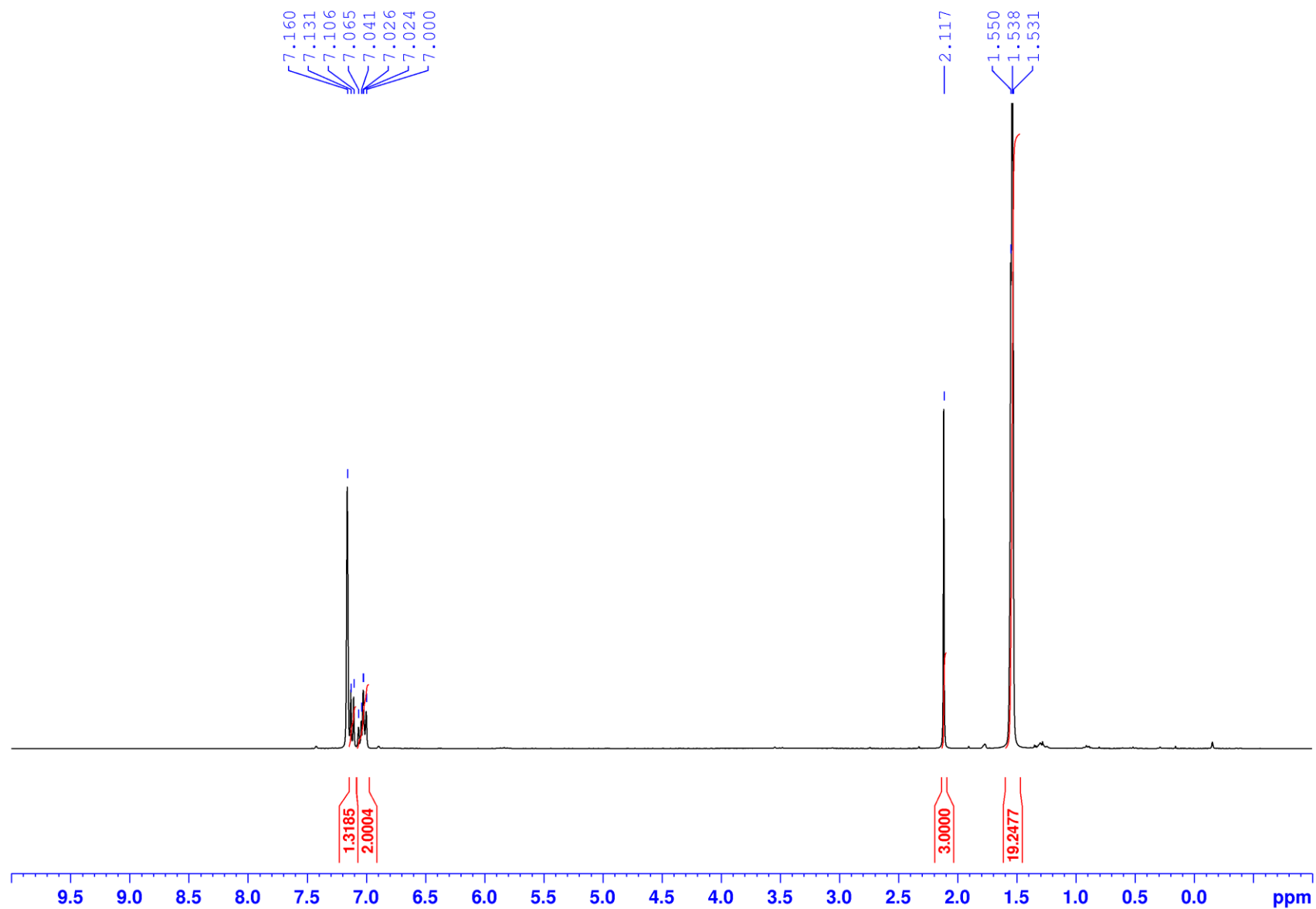
^1H NMR (300.1 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{THF}$ complex

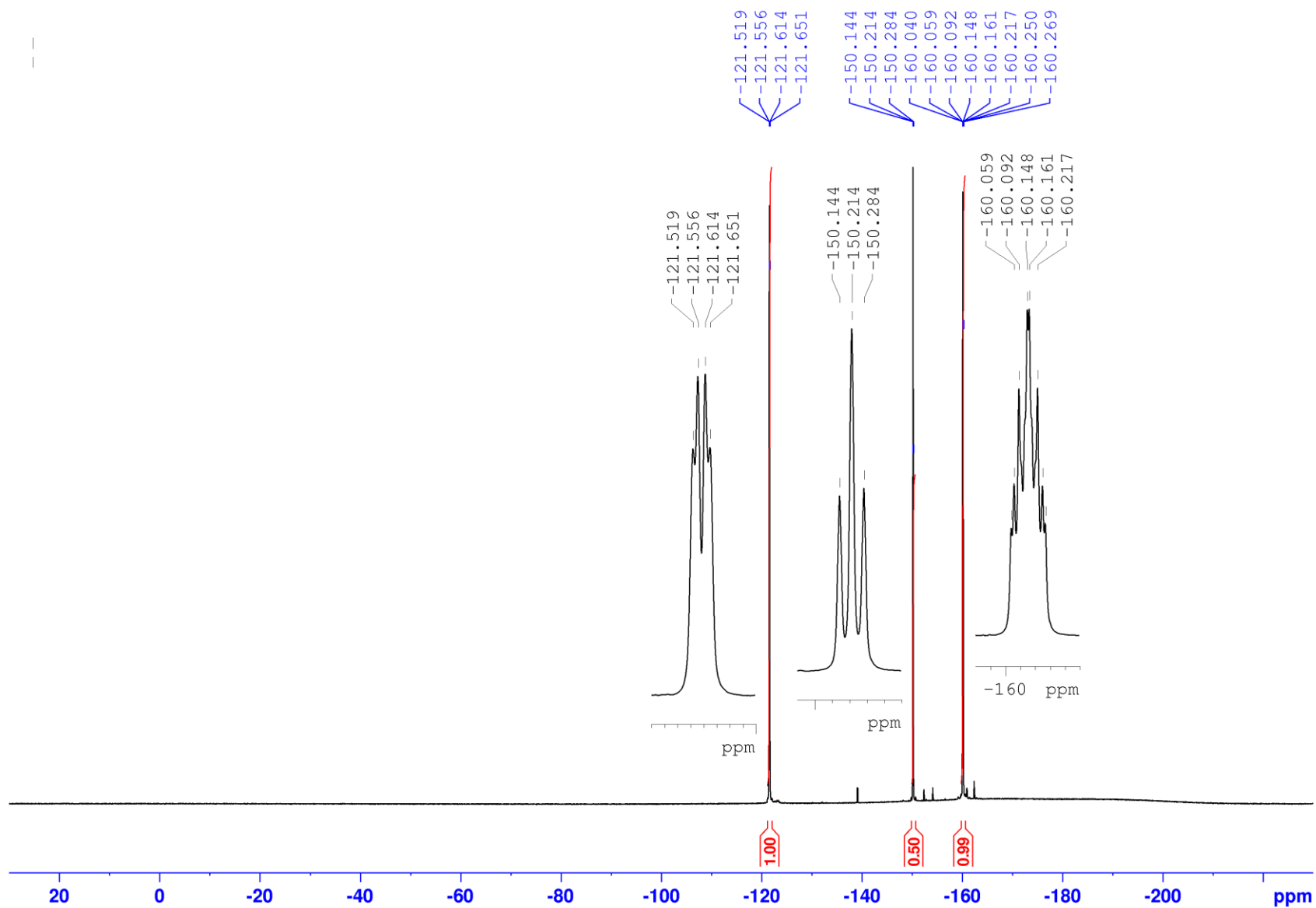
^{19}F NMR (282.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{THF}$ complex

^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{THF}$ complex

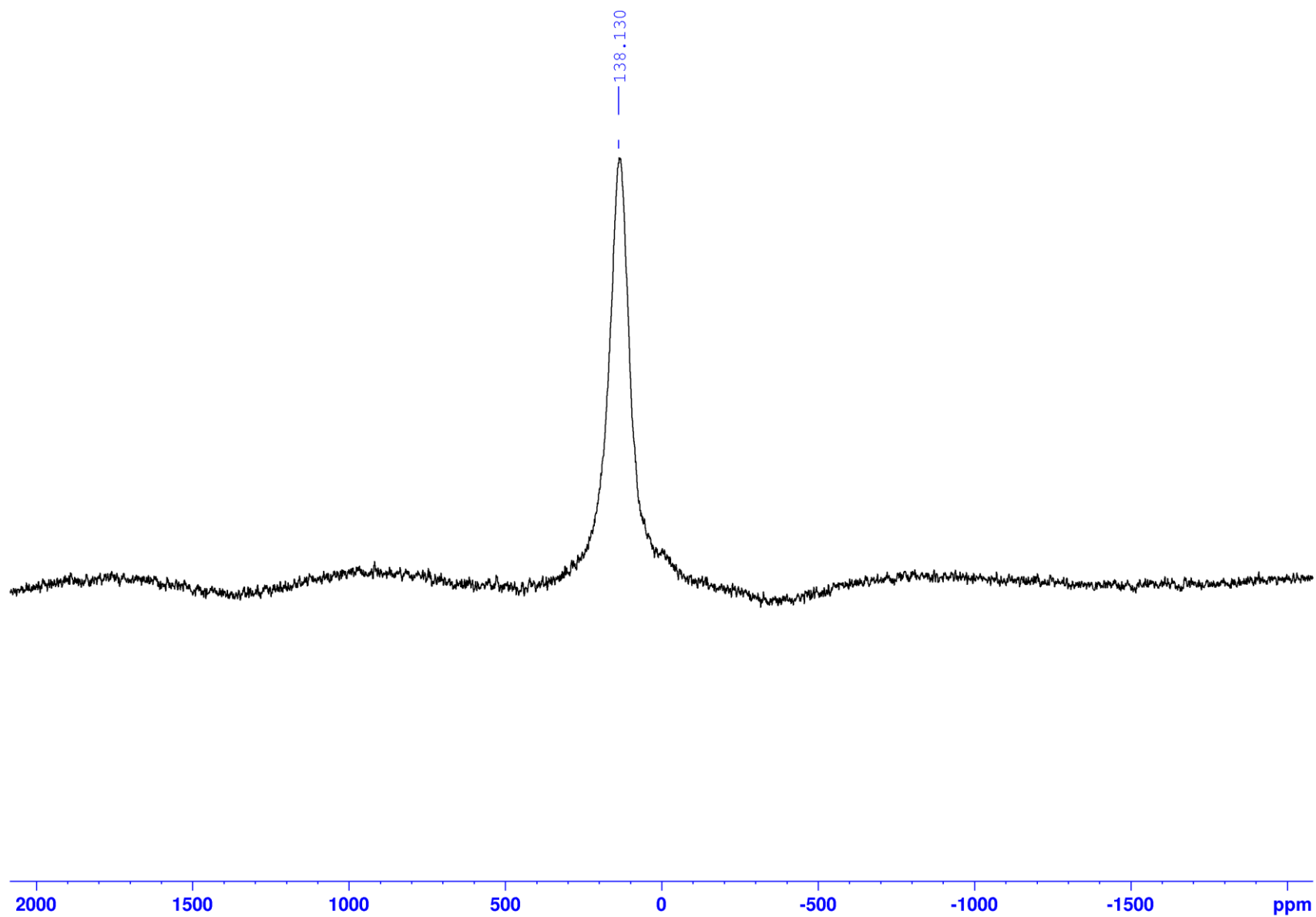


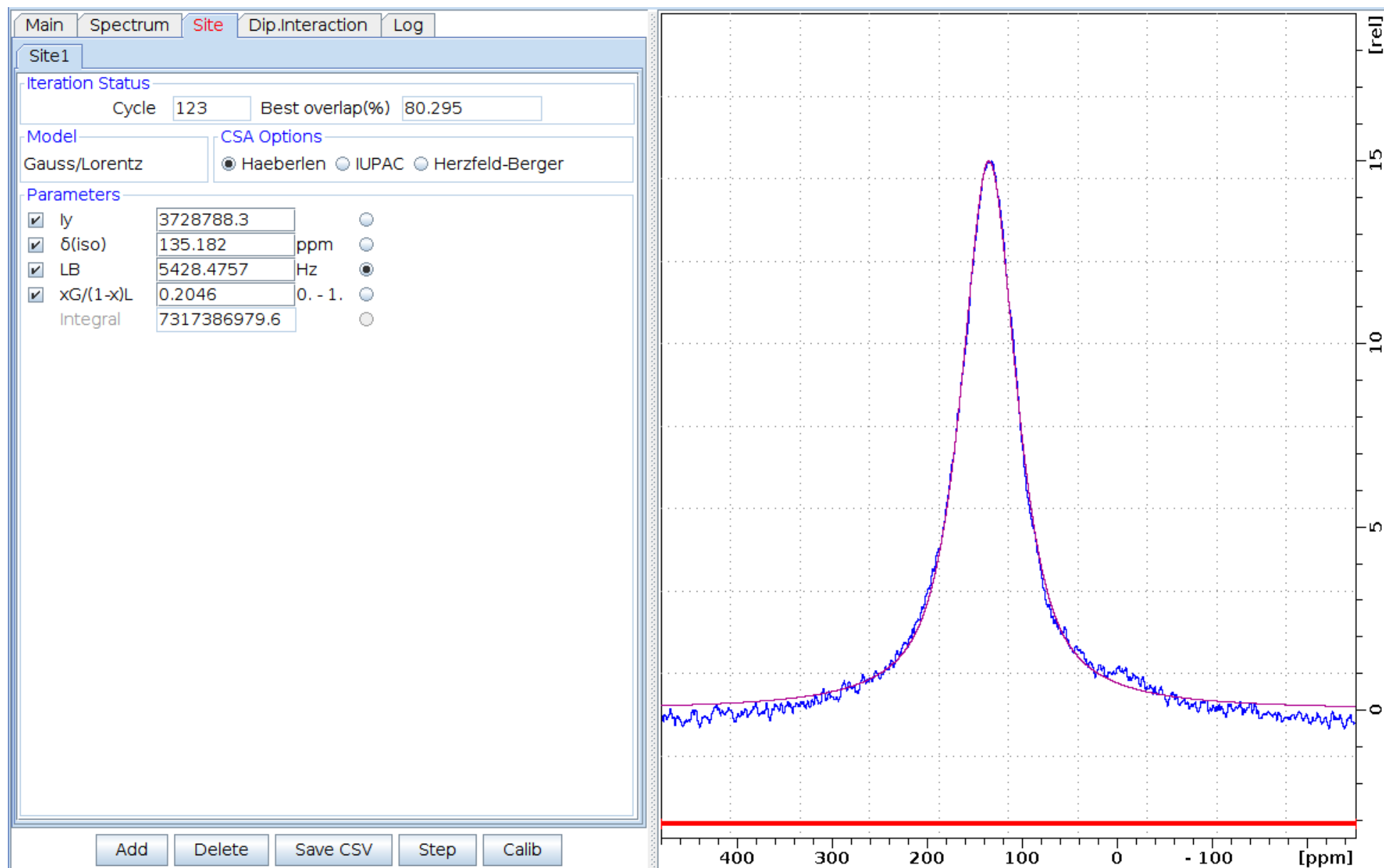
^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{THF}$ complex — Line shape analysis

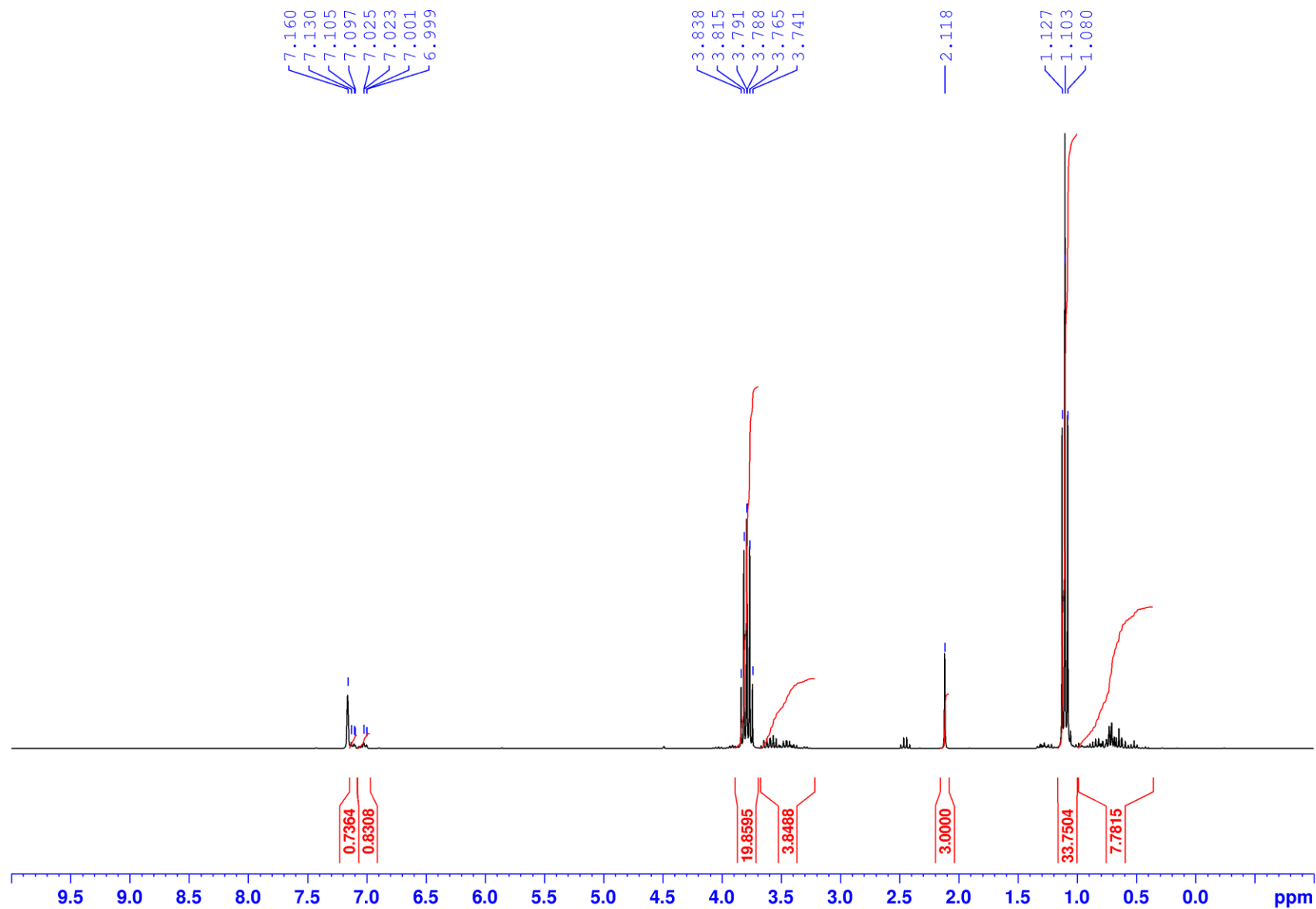
^1H NMR (300.1 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{Me}_2\text{S}$ complex

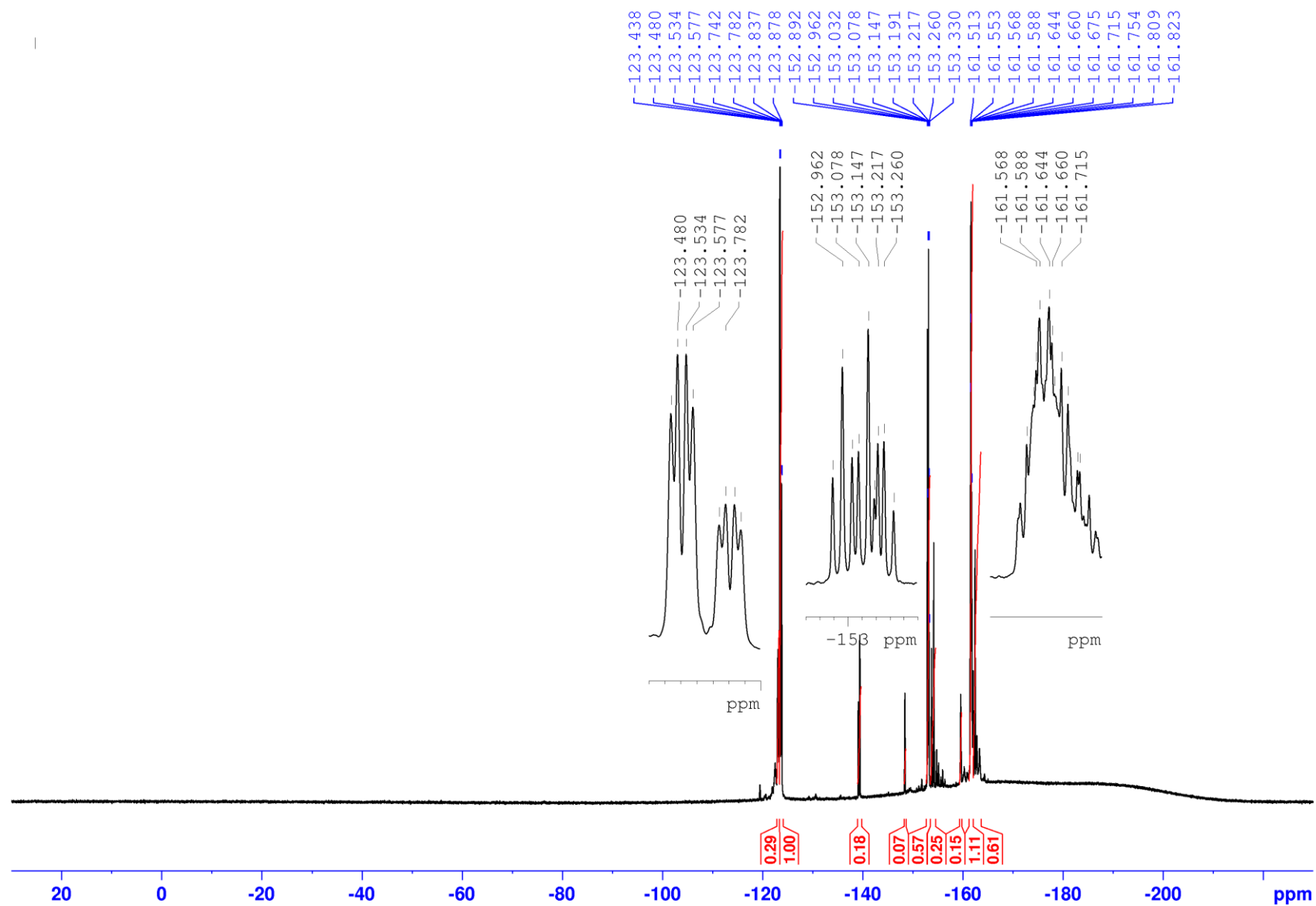
^{19}F NMR (282.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{Me}_2\text{S}$ complex

^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{Me}_2\text{S}$ complex

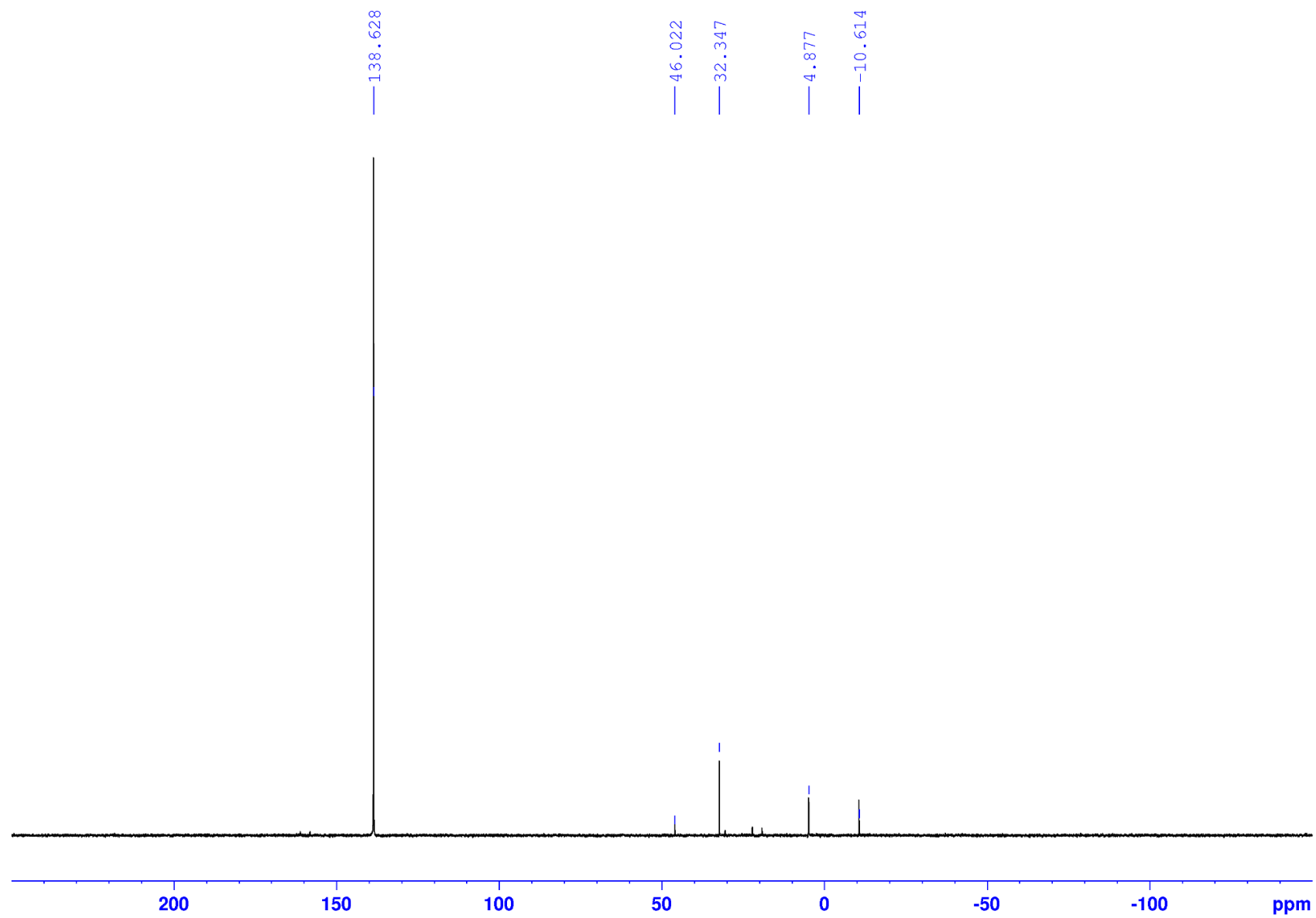


^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{Me}_2\text{S}$ complex — Line shape analysis

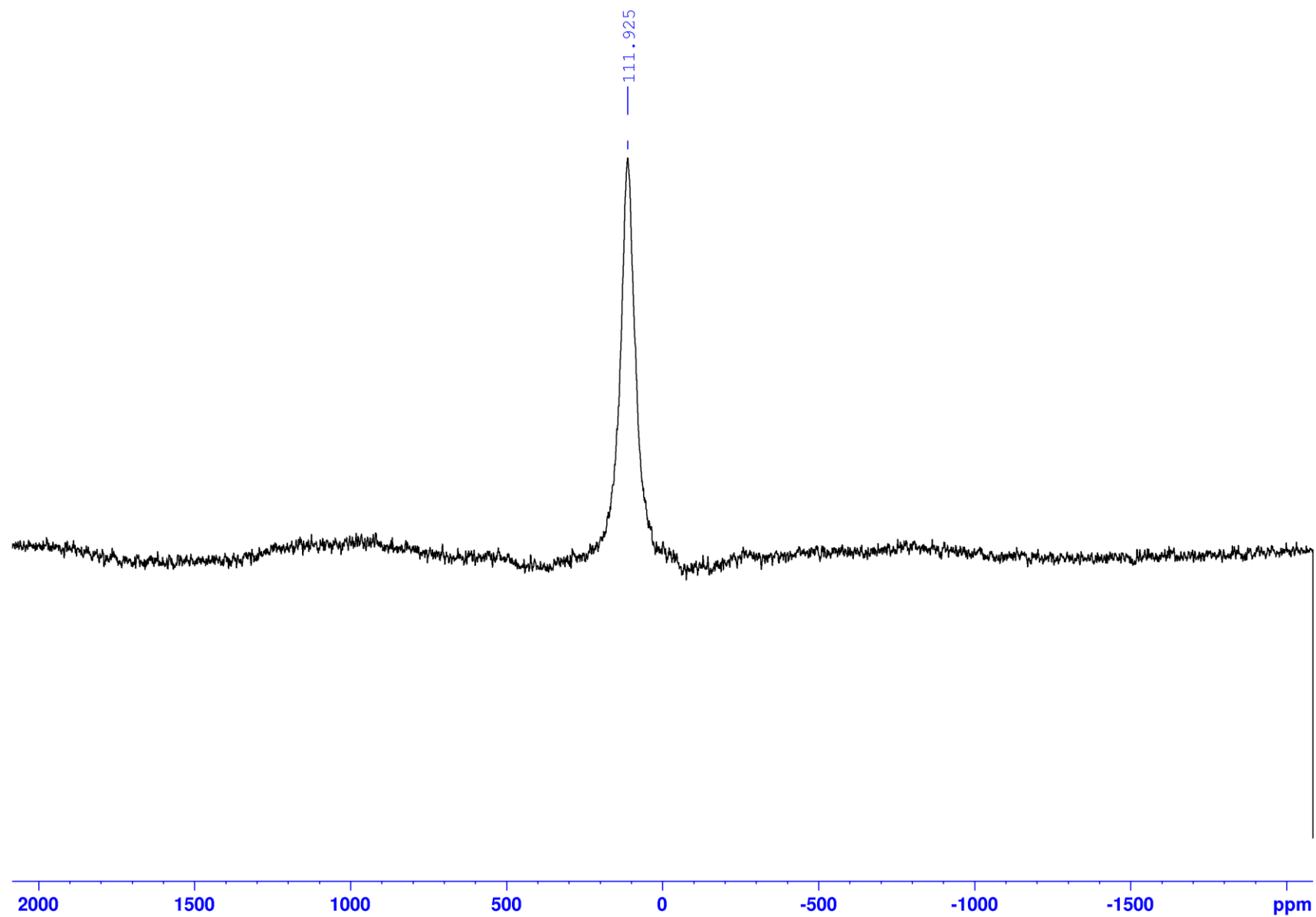
^1H NMR (300.1 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{P}(\text{OEt})_3$ complex

^{19}F NMR (282.4 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{P}(\text{OEt})_3$ complex

$^{31}\text{P}\{^1\text{H}\}$ NMR (121.5 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{P}(\text{OEt})_3$ complex



^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{P}(\text{OEt})_3$ complex



^{27}Al NMR (78.2 MHz, C_6D_6 , 298 K) of $\text{Al}(\text{C}_6\text{F}_5)_3 \cdot \text{P}(\text{OEt})_3$ complex — Line shape analysis