

Synthesis and cytotoxic activity of novel 4-amino-5-cyano-2-sulfonylpyrimidines

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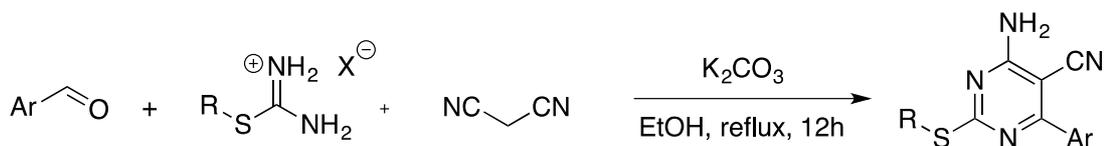
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Chemistry experimental

General methods

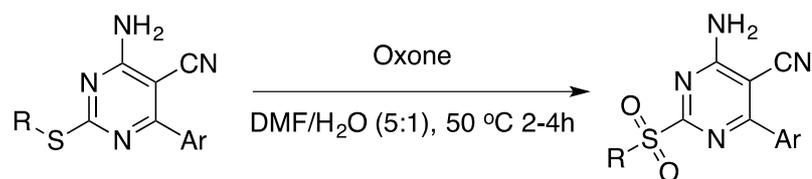
All reagents and solvents were used as purchased. ^1H and ^{13}C NMR spectra were acquired on a Bruker Avance spectrometer (600 and 150 MHz, respectively) in $\text{DMSO-}d_6$, internal standard was TMS. ^{19}F NMR spectra were recorded on the Bruker Avance III 500 spectrometer, operating at 471 MHz. ^{19}F chemical shifts was measured relative to CFCl_3 as an external standard. Elemental analysis was performed on a vario EL cube analyzer. Melting points were determined on a Boetius hot stage and were not corrected. All compounds reported in this communication are at least 95+% pure judged by HPLC UV detection (Agilent Technologies 1220 Infinity LC) and optionally NMR.

Synthesis of 5-cyanopyrimidines, structure type I

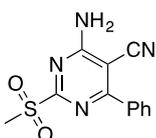


Synthesis of 2-alkylthio-4-amino-6-arylpyrimidine-5-carbonitriles (typical procedure)

A mixture of aromatic aldehyde (10 mmol), *S*-alkylisothioureia salt (hydrochloride, hydroiodide, hydrosulfate or mesylate, 10 mmol), malononitrile (10 mmol), potassium carbonate (20 mmol), and ethanol (100 ml) was refluxed for approximately 12 h. When the reaction was complete (TLC monitoring, hexane/EtOAc 7:3), the mixture was poured into water (400 ml), the solid was filtered off and washed thoroughly with water. The product was dried and recrystallized from 95% ethanol.

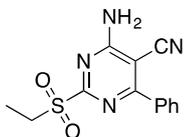


Synthesis of 2-alkylsulfonyl-4-amino-6-arylpyrimidine-5-carbonitriles 4a-k (typical procedure). To a solution of 2-alkylthio-4-amino-6-arylpyrimidine-5-carbonitrile (5 mmol) in DMF/H₂O mixture (5:1, 30 ml), Oxone (15 mmol) was added, and the mixture was stirred and heated to 50 °C for 2-4 h (HPLC monitoring, MeCN/H₂O, 80:20, C-18). When the reaction was complete, the mixture was poured into water (150 ml). The product was isolated by filtration and dried.



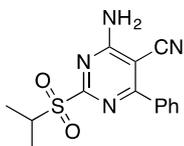
4-Amino-2-methylsulfonyl-6-phenylpyrimidine-5-carbonitrile (**1a**) was prepared using *S*-methylisothiuronium iodide [S1].

Yield 73%, mp 175-176 °C. Found, %: C 52.63; H 3.72; N 20.47. C₁₂H₁₀N₄O₂S. Calculated, %: C 52.55; H 3.67; N 20.43. ¹H NMR (600 MHz, DMSO-*d*₆): 8.69 (s, 2H, NH₂), 7.99 – 7.90 (m, 2H), 7.72 – 7.52 (m, 3H), 3.37 (s, 3H, SO₂Me). ¹³C NMR (150 MHz, DMSO-*d*₆): 169.4, 168.9, 166.5, 165.3, 135.5, 132.2, 129.2, 115.5, 89.1, 39.1.



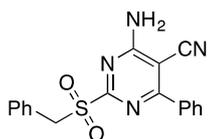
4-Amino-2-ethylsulfonyl-6-phenylpyrimidine-5-carbonitrile (**1b**) was prepared using *S*-ethylisothiuronium bromide [S2]

Yield 74%, mp 185-187 °C. Found, %: C 54.22; H 4.30; N 19.48. C₁₃H₁₂N₄O₂S. Calculated, %: C 54.16; H 4.20; N 19.43. ¹H NMR (600 MHz, DMSO-*d*₆): 8.69 (s, 2H, NH₂), 7.97 – 7.90 (m, 2H), 7.68 – 7.56 (m, 3H), 3.56 (q, *J* = 7.4 Hz, 2H, CH₂), 1.29 (t, *J* = 7.4 Hz, 3H, CH₃). ¹³C NMR (150 MHz, DMSO-*d*₆): 169.4, 165.8, 165.3, 135.5, 132.2, 129.3, 129.1, 115.5, 89.2, 45.3, 7.2.



4-Amino-2-isopropylsulfonyl-6-phenylpyrimidine-5-carbonitrile (**1c**) was prepared using *S*-isopropylisothiuronium iodide [S3]

Yield 89%, mp 198-200 °C. Found, %: C 55.74; H 4.73; N 18.57. C₁₄H₁₄N₄O₂S. Calculated, %: C 55.62; H 4.67; N 18.53. ¹H NMR (600 MHz, DMSO-*d*₆): 8.69 (s, 2H, NH₂), 7.96 – 7.89 (m, 2H), 7.67 – 7.56 (m, 3H), 3.93 (sept, *J* = 6.8 Hz, 1H, CH), 1.30 (d, *J* = 6.9 Hz, 6H, 2CH₃). ¹³C NMR (150 MHz, DMSO-*d*₆): 169.4, 165.3, 165.3, 135.5, 132.2, 129.2, 129.1, 115.5, 89.2, 50.6, 15.1.



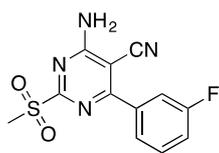
4-Amino-2-benzylsulfonyl-6-phenylpyrimidine-5-carbonitrile (**1d**) was prepared using *S*-benzylisothiuronium chloride [S4]

Yield 91%, mp 204-205 °C. Found, %: C 61.78; H 4.09; N 16.05. C₁₂H₁₀N₄O₂S. Calculated, %: C 61.70; H 4.03; N 15.99. ¹H NMR (600 MHz, DMSO-*d*₆): 8.83 (s, 2H, NH₂), 7.96 (dt, *J* = 7.1, 1.6 Hz, 2H), 7.70 – 7.58 (m, 3H), 7.39 (dq, *J* = 16.8, 3.3, 2.8 Hz, 5H), 4.95 (s, 2H, CH₂). ¹³C NMR (150 MHz, DMSO-*d*₆): 169.4, 166.0, 165.2, 135.4, 132.3, 132.0, 129.3, 129.2, 129.0, 128.9, 115.4, 89.1, 56.8.



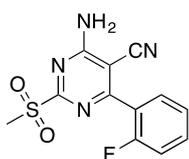
4-Amino-6-(4-fluorophenyl)-2-(methylsulfonyl)pyrimidine-5-carbonitrile (**1e**) was prepared using *S*-methylisothiuronium iodide [S1].

Yield 80%, mp 171-172 °C. Found, %: C 49.36; H 3.19; N 19.25. C₁₂H₉FN₄O₂S. Calculated, %: C 49.31; H 3.10; N 19.17. ¹H NMR (600 MHz, DMSO-*d*₆): 8.69 (s, 2H, NH₂), 8.09 – 7.98 (m, 2H), 7.45 (t, *J* = 8.9 Hz, 2H), 3.37 (s, 3H, SO₂Me). ¹³C NMR (150 MHz, DMSO-*d*₆): 168.2, 166.5, 165.3, 163.6 (d, *J* = 247.6 Hz), 132.1, 132.0, 132.0, 131.9, 116.3 (d, *J* = 22.1 Hz), 89.0, 39.1. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -107.98.



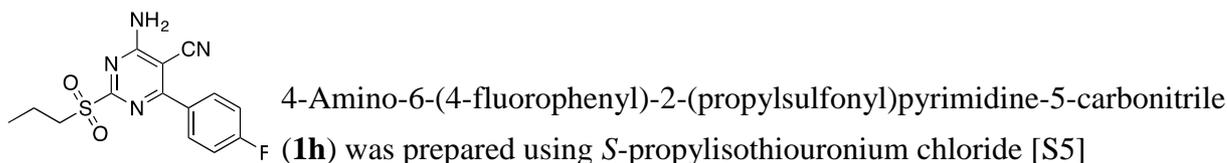
4-Amino-6-(3-fluorophenyl)-2-(methylsulfonyl)pyrimidine-5-carbonitrile (**1f**) was prepared using *S*-methylisothiuronium iodide [S1]

Yield 73%, mp 188-189 °C. Found, %: C 49.42; H 3.16; N 19.21. C₁₂H₉FN₄O₂S. Calculated, %: C 49.31; H 3.10; N 19.17. ¹H NMR (600 MHz, DMSO-*d*₆): 8.74 (s, 2H, NH₂), 7.85 – 7.43 (m, 4H), 3.38 (s, 3H, SO₂Me). ¹³C NMR (150 MHz, DMSO-*d*₆): 168.0, 166.5, 165.2, 163.3 (d, *J* = 244.5 Hz), 161.3, 137.7, 131.4 (d, *J* = 8.3 Hz), 125.5 (d, *J* = 2.9 Hz), 119.1 (d, *J* = 21.1 Hz), 116.1 (d, *J* = 23.5 Hz), 115.2, 89.5, 39.1. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -112.10.

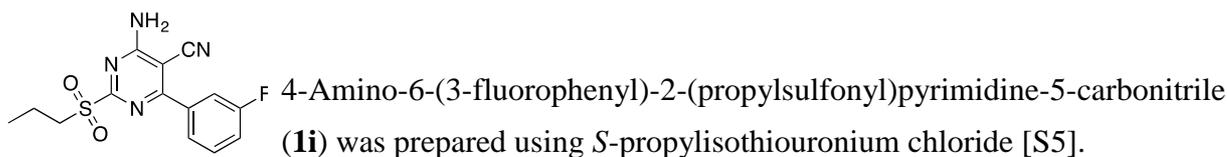


4-Amino-6-(2-fluorophenyl)-2-(methylsulfonyl)pyrimidine-5-carbonitrile (**1g**) was prepared using *S*-methylisothiuronium iodide [S1]

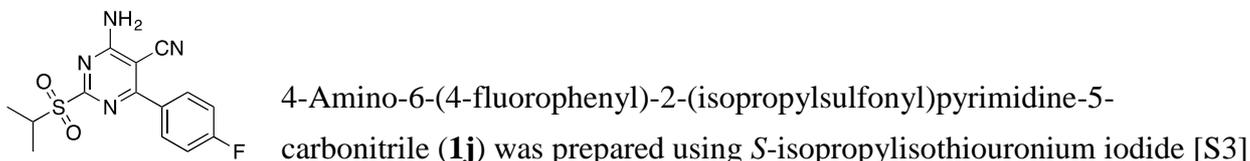
Yield 87%, mp 182-184 °C. Found, %: C 49.39; H 3.17; N 19.22. C₁₂H₉FN₄O₂S. Calculated, %: C 49.31; H 3.10; N 19.17. ¹H NMR (600 MHz, DMSO-*d*₆): 8.80 (s, 2H, NH₂), 7.73 – 7.65 (m, 2H), 7.51 – 7.38 (m, 2H), 3.34 (s, 3H, SO₂Me). ¹³C NMR (150 MHz, DMSO-*d*₆): 166.7, 166.2, 164.4, 160.3 (d, *J* = 248.2 Hz), 158.3, 134.0 (d, *J* = 8.5 Hz), 131.7, 125.5, 124.0, 123.9, 116.7 (d, *J* = 21.2 Hz), 114.4, 92.1, 39.2. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -113.41.



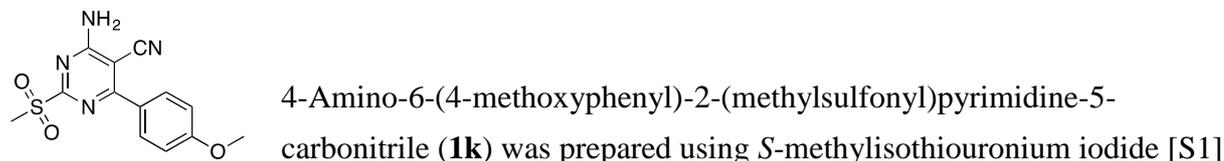
Yield 86%, mp 170-171 °C. Found, %: C 52.58; H 4.15; N 17.55. C₁₄H₁₃FN₄O₂S. Calculated, %: C 52.49; H 4.09; N 17.49. ¹H NMR (600 MHz, DMSO-*d*₆): 8.70 (s, 2H, NH₂), 8.06 – 7.98 (m, 2H), 7.46 (t, *J* = 8.8 Hz, 2H), 3.57 – 3.49 (m, 2H, CH₂), 1.77 (h, *J* = 7.5 Hz, 2H, CH₂), 1.01 (t, *J* = 7.4 Hz, 3H, CH₃). ¹³C NMR (150 MHz, DMSO-*d*₆): 168.2, 166.0, 165.5, 165.3, 163.5 (d, *J* = 247.7 Hz), 132.0 (d, *J* = 9.5 Hz), 116.3 (d, *J* = 22.0 Hz), 115.4, 89.0, 52.1, 16.1, 13.3. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -108.03.



Yield 68%, mp 168-170 °C. Found, %: C 52.57; H 4.14; N 17.54. C₁₂H₁₀N₄O₂S. Calculated, %: C 52.49; H 4.09; N 17.49. ¹H NMR (600 MHz, DMSO-*d*₆): 8.75 (s, 2H, NH₂), 7.80 – 7.76 (m, 1H), 7.75 – 7.71 (m, 1H), 7.70 – 7.63 (m, 1H), 7.53 – 7.47 (m, 1H), 3.61 – 3.49 (m, 2H, CH₂), 1.85 – 1.70 (m, 2H, CH₂), 1.02 (t, *J* = 7.5 Hz, 3H, CH₃). ¹³C NMR (150 MHz, DMSO-*d*₆): 168.0, 166.1, 165.2, 162.3 (d, *J* = 241.2 Hz), 137.7, 131.4 (d, *J* = 8.2 Hz), 125.5 (d, *J* = 2.7 Hz), 119.0 (d, *J* = 21.0 Hz), 116.1 (d, *J* = 23.5 Hz), 115.2, 89.6, 52.1, 16.1, 13.3. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -112.15.

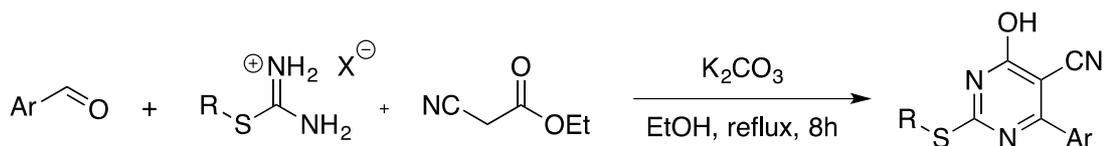


Yield 85%, mp 225-227 °C. Found, %: C 52.61; H 4.16; N 17.58. C₁₄H₁₃FN₄O₂. Calculated, %: C 52.49; H 4.09; N 17.49. ¹H NMR (600 MHz, DMSO-*d*₆): 8.70 (s, 2H, NH₂), 8.07 – 7.95 (m, 2H), 7.51 – 7.38 (m, 2H), 3.92 (p, *J* = 6.9 Hz, 1H, CH), 1.29 (d, *J* = 6.9 Hz, 6H, 2CH₃). ¹³C NMR (150 MHz, DMSO-*d*₆): 168.2, 165.3, 163.5 (d, *J* = 247.7 Hz), 132.0 (d, *J* = 9.2 Hz), 116.3 (d, *J* = 22.1 Hz), 89.1, 50.6, 15.0.



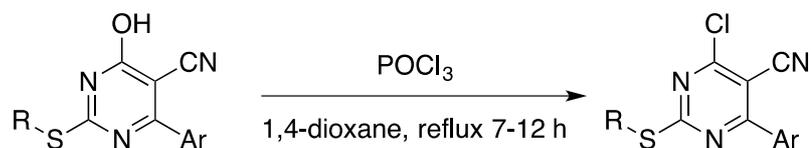
Yield 66%, mp 146-147 °C. Found, %: C 51.37; H 4.07; N 18.46. C₁₃H₁₂N₄O₃S. Calculated, %: C 51.31; H 3.97; N 18.41. ¹H NMR (600 MHz, DMSO-*d*₆): 8.58 (s, 2H, NH₂), 8.00 (d, *J* = 8.9 Hz, 2H), 7.15 (d, *J* = 8.9 Hz, 2H), 3.87 (s, 3H, OMe), 3.37 (s, 3H, SO₂Me). ¹³C NMR (150 MHz, DMSO-*d*₆): 168.4, 166.4, 165.5, 162.7, 131.3, 127.5, 115.9, 114.6, 87.6, 56.0, 39.1.

Synthesis of 5-cyanopyrimidines, structure type II



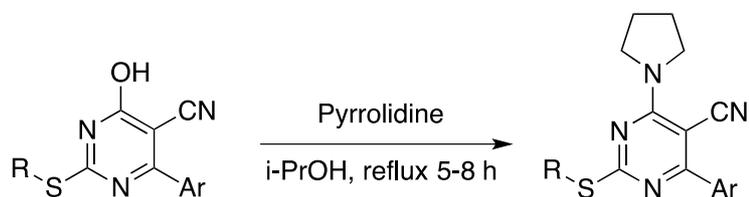
Synthesis of 2-alkylthio-6-aryl-4-hydroxypyrimidine-5-carbonitriles (typical procedure)

A mixture of aromatic aldehyde (10 mmol), *S*-alkylisothioureia salt (hydrochloride, hydroiodide, hydrosulfate or mesylate, 10 mmol), ethyl cyanoacetate (10 mmol), potassium carbonate (20 mmol), and ethanol (100 ml) was refluxed for approximately 8 h. When the reaction was complete (TLC monitoring, hexane/EtOAc 7:3), the mixture was poured into water (400 ml), acidified (5% solution HCl). The product was isolated by filtration, dried, and recrystallized from 95 % ethanol.

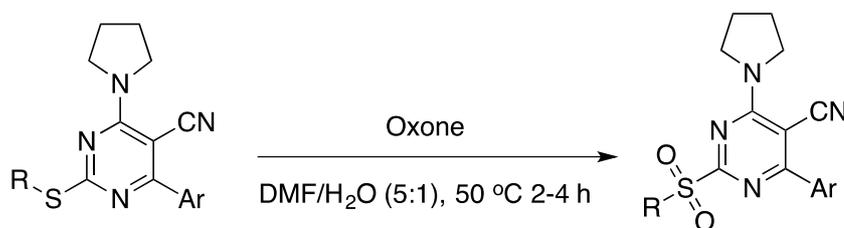


Synthesis of 2-alkylthio-6-aryl-4-chloropyrimidine-5-carbonitriles (typical procedure)

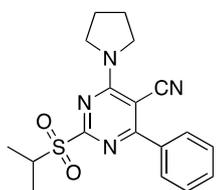
A mixture of 2-alkylthio-6-aryl-4-hydroxypyrimidine-5-carbonitrile (5 mmol), 1,4-dioxane (70 ml) and POCl₃ (50 mmol) was refluxed for 7-12 h (TLC monitoring, hexane/EtOAc 7:3) and the concentrated under vacuum. The residue was dissolved in EtOAc (100 ml) and washed with cold water, saturated NaHCO₃ solution, water, and brine. The solution was dried over MgSO₄ and concentrated under vacuum. The residue was recrystallized from 95% ethanol.



Synthesis of 2-alkylthio-4-aryl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitriles (typical procedure). A mixture of 2-alkylthio-6-aryl-4-chloropyrimidine-5-carbonitriles (3 mmol), pyrrolidine (6 mmol) and i-PrOH (50 ml) was refluxed for 5-8 h (TLC monitoring, hexane/EtOAc 7:3). When the reaction was complete, the reaction mixture was poured into water (200 ml). The product was isolated by filtration, dried, and recrystallized from 95% ethanol.

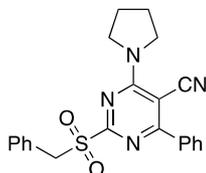


Synthesis of 2-alkylsulfonyl-4-aryl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitriles (typical procedure). To a solution of 2-alkylthio-4-aryl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (2 mmol) in DMF/H₂O mixture (5:1, 30 ml), Oxone (6 mmol) was added, and the mixture was stirred and heated to 50 °C for 2-4 h (HPLC monitoring, MeCN/H₂O, 80:20, C-18). When the reaction was complete, the mixture was poured into water (120 ml). The product was isolated by filtration and dried.



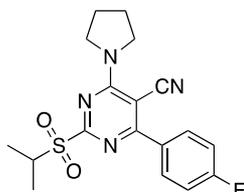
2-Isopropylsulfonyl-4-phenyl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (**2a**) was prepared using *S*-isopropylisothiuronium iodide.

Yield 55%, mp 137-139 °C. Found, %: C 60.73; H 5.72; N 15.80. C₁₈H₂₀N₄O₂S. Calculated, %: C 60.65; H 5.66; N 15.72. ¹H NMR (600 MHz, DMSO-*d*₆): 7.86 (td, *J* = 7.9, 1.5 Hz, 2H), 7.67 – 7.52 (m, 3H), 3.99 (p, *J* = 6.9 Hz, 1H, CH), 3.71 (s, 4H), 1.35 – 1.30 (m, 10H). ¹³C NMR (150 MHz, DMSO-*d*₆): 171.5, 170.7, 164.1, 159.6, 136.0, 132.0, 131.8, 129.7, 129.0, 128.9, 117.5, 89.2, 52.9, 50.6, 17.6, 15.2, 14.0.



2-Benzylsulfonyl-4-phenyl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (**2b**) was prepared using *S*-benzylisothiuronium chloride.

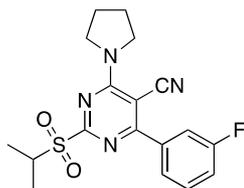
Yield 85%, mp 205-207 °C. Found, %: C 65.41; H 5.07; N 13.93. C₂₂H₂₀N₄O₂S. Calculated, %: C 65.33; H 4.98; N 13.85. ¹H NMR (600 MHz, DMSO-*d*₆): 7.98 – 7.13 (m, 10H), 4.97 (s, 2H, CH₂), 4.14 – 3.65 (m, 4H), 2.12 – 1.76 (m, 4H). ¹³C NMR (150 MHz, DMSO-*d*₆): 172.9, 171.4, 170.8, 164.8, 159.4, 136.3, 135.9, 132.0, 131.8, 131.1, 130.7, 129.1, 128.0, 117.8, 89.1, 59.3, 56.4, 26.5, 24.0.



4-(4-Fluorophenyl)-2-isopropylsulfonyl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (**2c**) was prepared using *S*-isopropylisothiuronium iodide.

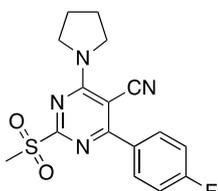
Yield 77%, mp 105-107 °C. Found, %: C 57.85; H 5.18; N 15.05. C₁₈H₁₉FN₄O₂S. Calculated, %: C 57.74; H 5.11; N 14.96. ¹H NMR (600 MHz, DMSO-*d*₆): 7.98 – 7.93 (m, 2H), 7.49 – 7.40 (m, 2H), 3.99 (p, *J* = 6.9 Hz, 1H, CH), 3.79 – 3.64 (m, 4H), 2.11 – 1.88 (m, 4H), 1.32 (d, *J* =

6.9 Hz, 6H, 2CH₃). ¹³C NMR (150 MHz, DMSO-*d*₆): 170.3, 165.4, 164.1, 163.5 (d, *J* = 248.1 Hz), 159.5, 132.4 (d, *J* = 9.2 Hz), 117.4, 116.1, 89.1, 50.6, 49.6, 45.3, 26.5, 24.0, 15.2. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -108.31.



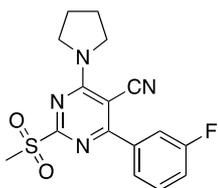
4-(3-Fluorophenyl)-2-isopropylsulfonyl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (**2d**) was prepared using *S*-isopropylisothiuronium iodide.

Yield 56%, mp 138-140 °C. Found, %: C 57.80; H 5.16; N 15.03. C₁₈H₁₉FN₄O₂S. Calculated, %: C 57.74; H 5.11; N 14.96. ¹H NMR (600 MHz, DMSO-*d*₆): 7.75 – 7.43 (m, 3H), 3.99 (p, *J* = 6.9 Hz, 1H, CH), 3.79 – 3.64 (m, 4H), 2.11 – 1.88 (m, 4H), 1.32 (d, *J* = 6.9 Hz, 6H, 2CH₃). ¹³C NMR (150 MHz, DMSO-*d*₆): 170.0, 169.3, 164.1, 163.1 (d, *J* = 244.5 Hz), 161.2, 131.4, 131.2, 125.9, 118.9, 117.7, 89.6, 52.9, 50.6, 26.5, 23.9, 17.7, 15.2. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -112.35.



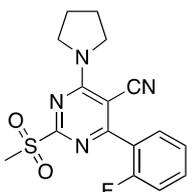
4-(4-Fluorophenyl)-2-methylsulfonyl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (**2e**) was prepared using *S*-methylisothiuronium iodide.

Yield 78%, mp 95-97 °C. Found, %: C 55.58; H 4.47; N 16.23. C₁₆H₁₅FN₄O₂S. Calculated, %: C 55.48; H 4.37; N 16.18. ¹H NMR (600 MHz, DMSO-*d*₆): 8.05 – 7.92 (m, 2H), 7.51 – 7.40 (m, 2H), 4.17 – 3.60 (m, 4H), 3.39 (s, 3H, SO₂Me), 2.20 – 1.76 (m, 4H). ¹⁹F NMR (471 MHz, DMSO-*d*₆): -108.27.



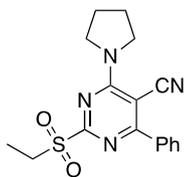
4-(3-Fluorophenyl)-2-methylsulfonyl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (**2f**) was prepared using *S*-methylisothiuronium iodide.

Yield 85%, mp 148-150 °C. Found, %: C 55.48; H 5.45; N 16.25. C₁₆H₁₅FN₄O₂S. Calculated, %: C 55.48; H 4.37; N 16.18. ¹H NMR (600 MHz, DMSO-*d*₆): 7.78 – 7.59 (m, 3H), 7.56 – 7.41 (m, 1H), 4.11 – 3.64 (m, 4H), 3.40 (s, 3H, SO₂Me), 2.12 – 1.86 (m, 4H). ¹³C NMR (150 MHz, DMSO-*d*₆): 174.6, 165.3, 163.2 (d, *J* = 244.5 Hz), 161.2, 159.4, 131.1, 125.9, 117.2, 116.4, 89.5, 87.7, 39.9, 26.5, 24.0. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -112.36.



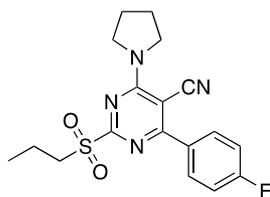
4-(2-Fluorophenyl)-2-methylsulfonyl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (**2g**) was prepared using *S*-methylisothiuronium iodide.

Yield 98%, mp 149-151 °C. Found, %: C 55.57; H 4.45; N 16.29. C₁₂H₁₀N₄O₂S. Calculated, %: C 55.48; H 4.37; N 16.18. ¹H NMR (600 MHz, DMSO-*d*₆): 7.71 – 7.62 (m, 2H), 7.50 – 7.39 (m, 2H), 4.05 – 3.67 (m, 4H), 3.38 (s, 3H, SO₂Me), 2.11 – 1.89 (m, 4H). ¹³C NMR (150 MHz, DMSO-*d*₆): 168.0, 165.5, 160.4 (d, *J* = 248.3 Hz), 158.5, 133.8 (d, *J* = 8.5 Hz), 131.7, 125.4, 124.5, 116.6 (d, *J* = 21.1 Hz), 116.4, 91.8, 50.4, 49.3, 39.1, 26.4, 23.9. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -113.86.



2-Ethylsulfonyl-4-phenyl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (**2h**) was prepared using *S*-ethylisothiuronium bromide.

Yield 73%, mp 158-160 °C. Found, %: C 59.69; H 5.36; N 16.42. C₁₇H₁₈N₄O₂S. Calculated, %: C 59.63; H 5.30; N 16.36. ¹H NMR (600 MHz, DMSO-*d*₆): 7.90 – 7.85 (m, 2H), 7.68 – 7.56 (m, 3H), 4.07-3.67 (m, 4H), 3.59 (q, *J* = 7.3 Hz, 2H, CH₂), 2.12 – 1.85 (m, 4H), 1.31 (t, *J* = 7.4 Hz, 3H, CH₃). ¹³C NMR (150 MHz, DMSO-*d*₆): 171.5, 164.5, 159.6, 136.0, 132.0, 129.7, 129.0, 117.4, 89.1, 50.6, 49.5, 45.3, 26.5, 24.0, 7.4.



4-(4-Fluorophenyl)-2-propylsulfonyl-6-(pyrrolidin-1-yl)pyrimidine-5-carbonitrile (**2i**) was prepared using *S*-propylisothiuronium chloride.

Yield 70%, mp 101-103 °C. Found, %: C 57.81; H 5.21; N 15.04. C₁₈H₁₉FN₄O₂S. Calculated, %: C 57.74; H 5.11; N 14.96. ¹H NMR (600 MHz, DMSO-*d*₆): 8.00 – 7.91 (m, 1H), 7.46 – 7.40 (m, 1H), 4.10 – 3.63 (m, 4H), 3.57 – 3.49 (m, 2H, CH₂), 2.10 – 1.87 (m, 4H), 1.77 (h, *J* = 7.5 Hz, 2H, CH₂), 1.01 (t, *J* = 7.4 Hz, 3H, CH₃). ¹³C NMR (150 MHz, DMSO-*d*₆): 173.9, 169.7, 165.3, 163.3 (d, *J* = 248.3 Hz), 159.4, 132.3, 131.9, 118.7, 117.9, 116.3, 87.1, 54.6, 52.0, 49.4, 32.7, 22.9, 16.0, 13.4. ¹⁹F NMR (471 MHz, DMSO-*d*₆): -108.7.

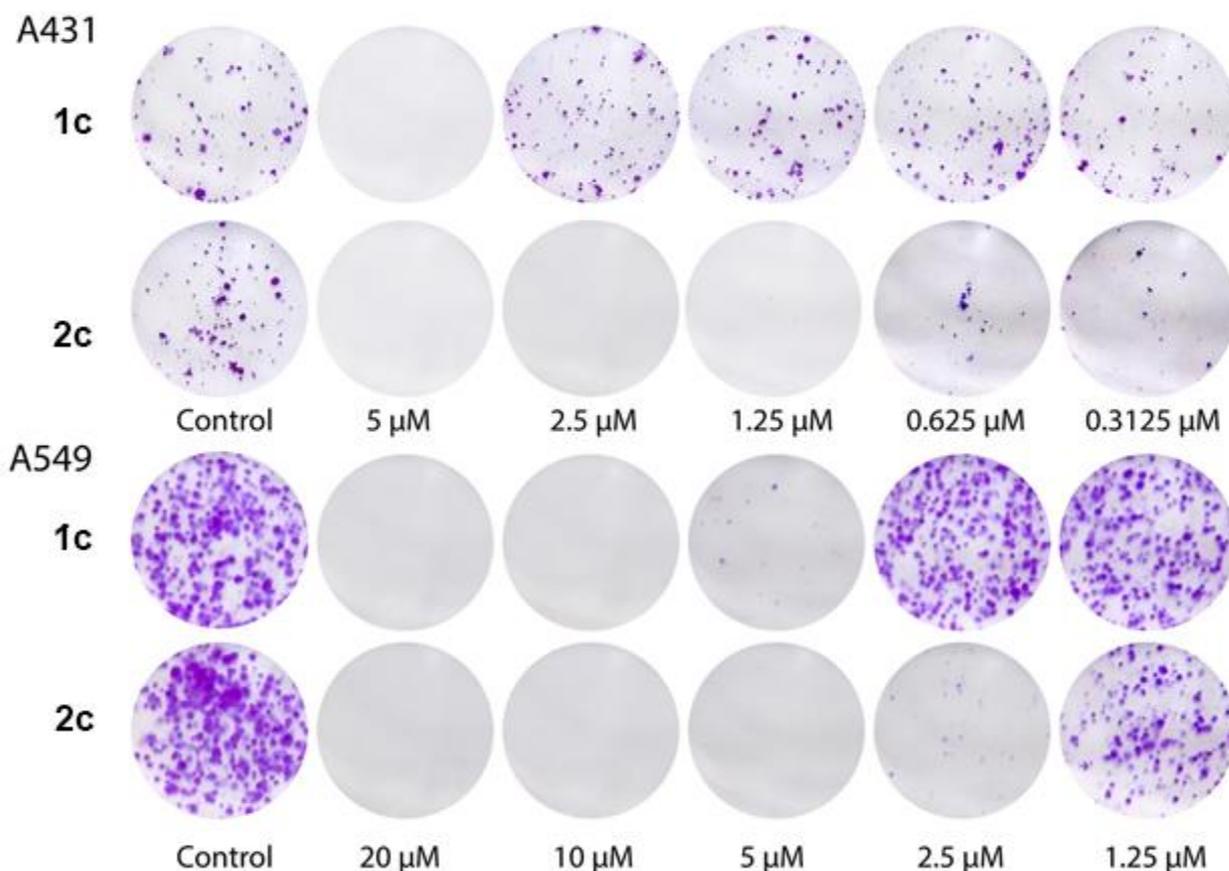
Biological assays

Cell Proliferation Assay [S6]. The human lung adenocarcinoma cell line A549, human epidermoid carcinoma cell line A431, human melanoma cell line A375, human colorectal cancer cell lines HCT-116, human breast cancer cell line MCF-7, human prostate cancer cells LNCaP, human neuroblastoma cell lines SH-SY5Y were used to evaluate the potency of synthesized analogues in cell-based level. All cell lines were purchased from American type culture collection (ATCC). A431, A375, MCF-7 cell line was cultured in DMEM (Gibco), A549, HCT-116 and LnCaP were cultured with RPMI 1640 (Gibco), SH-SY5Y cell line was cultured in

DMEM/F12 medium (Gibco). All culture mediums were supplemented with *L*-glutamine, penicillin/streptomycin (Invitrogen) and 10% fetal bovine serum (Gibco).

All cells were seeded in 96-well tissue culture plates (Eppendorf) in density of 7×10^3 cells/well in 200 μ l of complete growth media for 24 hours. Triplicate wells were treated with test compounds starting at 500.0 μ M concentration and diluted 12 times for 48 hours at various concentrations or DMSO (Sigma) as control with final concentration 0.1%. Plates were incubated at 37°C in 5% CO₂ atmosphere. Following treatment, 40 μ L of MTT (5 mg ml⁻¹) reagent was incubated in each well for 4 hr. After 4 h, medium/MTT mixtures were replaced with DMSO (200 μ l per well), and optical density at a wavelength of 570 nm was measured with a microplate reader (GloMax Multi+, Promega).

Clonogenic Survival Assay [S7]. A549 and A431 cells were placed in 6-well plates at 1000 cells per well. Twenty-four hours later cells were treated with varying concentrations of 5-cyanopyrimidine derivatives and then monitored microscopically for colony growth. Ten days later the cells were fixed with 4% paraformaldehyde solution. Following fixation for 15 min, the plates were washed and stained with a 0.4% solution of crystal violet in 20% methanol for 30 min, washed with PBS, and air-dried. Colonies consist more than 50 cells were counted as positive. Results are normalized to percent clonogenic survival from untreated control cells (Figure S1)



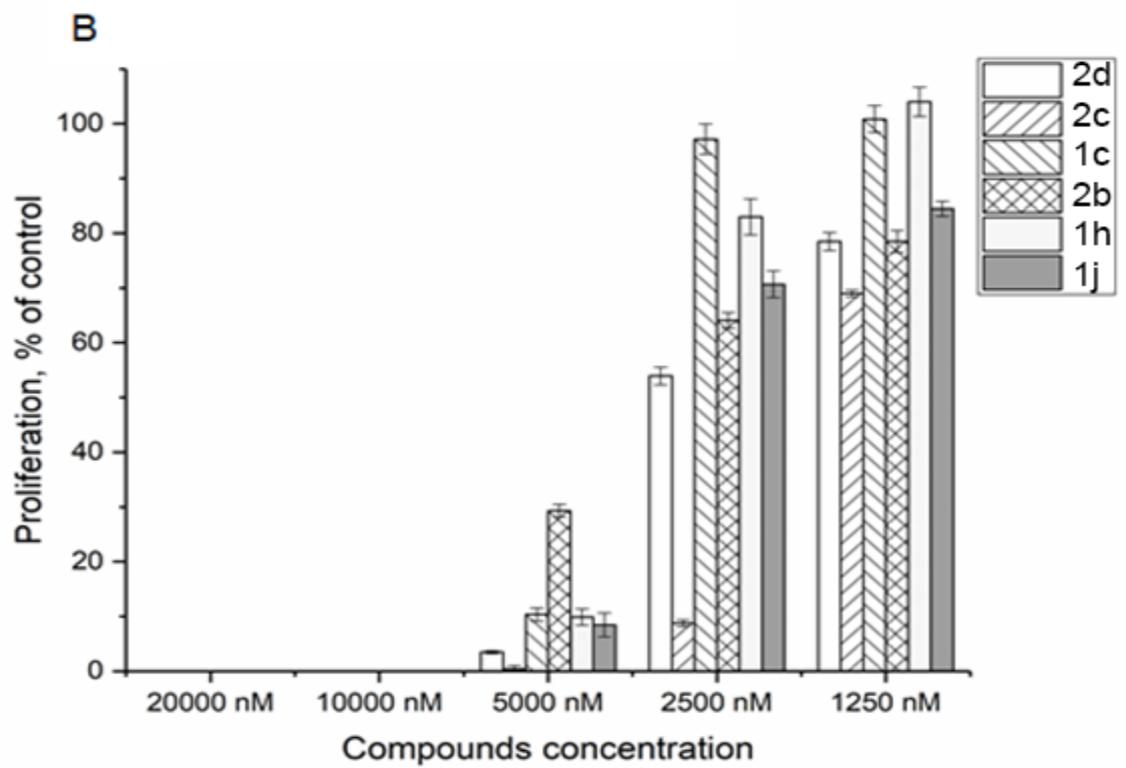
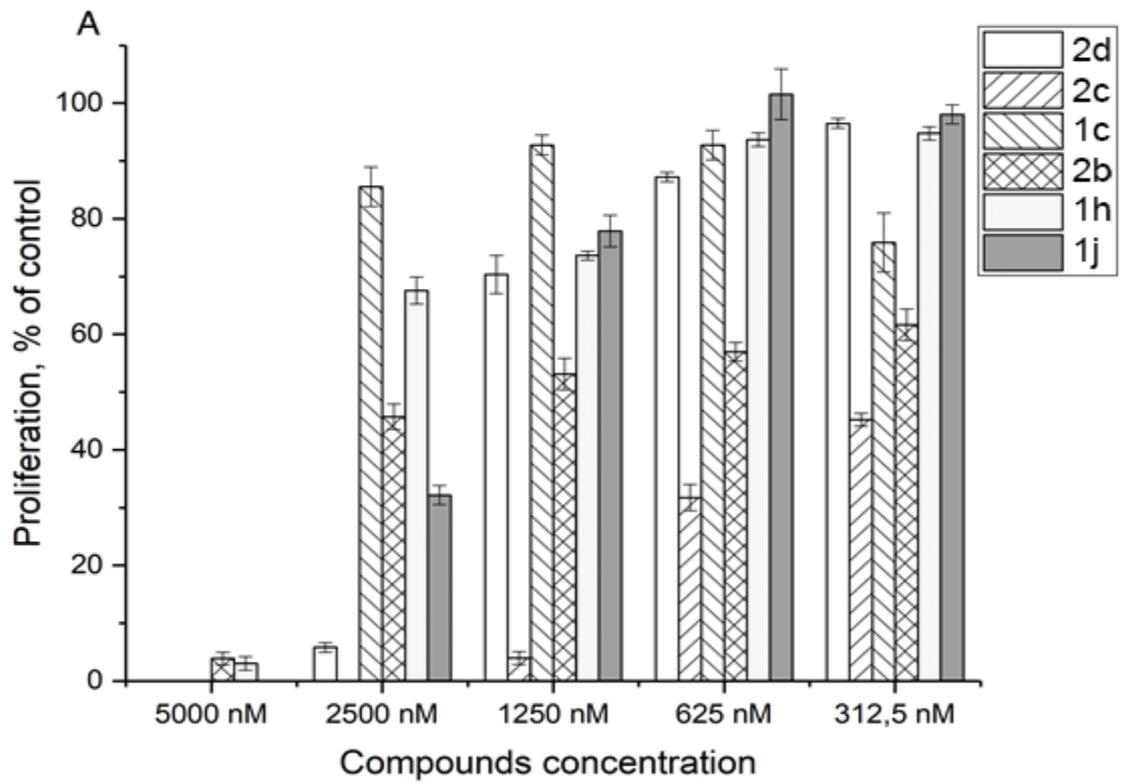


Figure S1. Clonogenic survival assay. (A) A431 cell line, (B) A549 cell line

Apoptosis Assay [S8] The induction of apoptosis A549 and A431 cancer cells by 5-cyanopyrimidine derivatives was assayed by staining with Annexin V and propidium iodide (PI) using the AnnexinV-FITC kit (Invitrogen, USA) in accordance with the manufacturer's protocol. After incubation cells with the 5-cyanopyrimidine derivatives for 24 h, they were centrifuged, the pellet was resuspended in 100 μ l of binding buffer and a solution containing PI and annexin V was added. The cells were incubated at room temperature in the dark for 15 minutes, then 400 μ l of binding buffer was added to stop the reaction. Data analysis (at least 2×10^4 events) was performed on a NovoCyte 2000R flow cytometer (ACEA Biosciences, San Diego, CA, USA) using NovoExpress v.1.2.4 software.

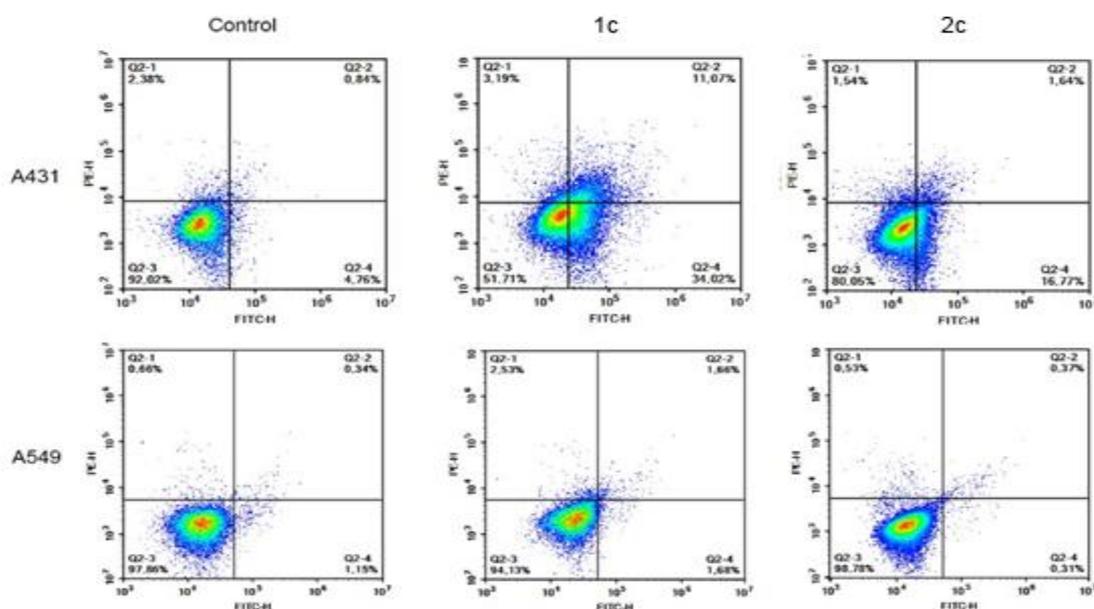


Figure S2. Apoptosis assay on A431 and A549 cell lines

Cell cycle analysis [S9]. A549 and A431 cancer cells were placed in 6-well cultures plates (Nunc, Denmark) at 2×10^5 cells per well and cultivated overnight in complete grow RPMI 1640 media. After media was replaced on RPMI 1640 with 2% FBS and incubate for 24 h.

Tested compound in noncytotoxic concentration were added to A549 cells in RPMI 1640 media with 2% FBS and 20 ng/ml recombinant human EGF (Invitrogen, USA). After 24 h of treatment, the cells were harvested, washed with PBS. The cell pellets were resuspended in 500 μ l of 50 μ g/ml solution of propidium iodide (PI) in buffer (BD Biosciences, Franklin Lakes, NJ, USA) and incubated in the dark at room temperature for 15 min. The PI fluorescence signal was assessed on a NovoCyte 2000R flow cytometer (ACEA Biosciences, San Diego, CA, USA) and the cell cycle distribution was analyzed using NovoExpress v.1.2.4 software.

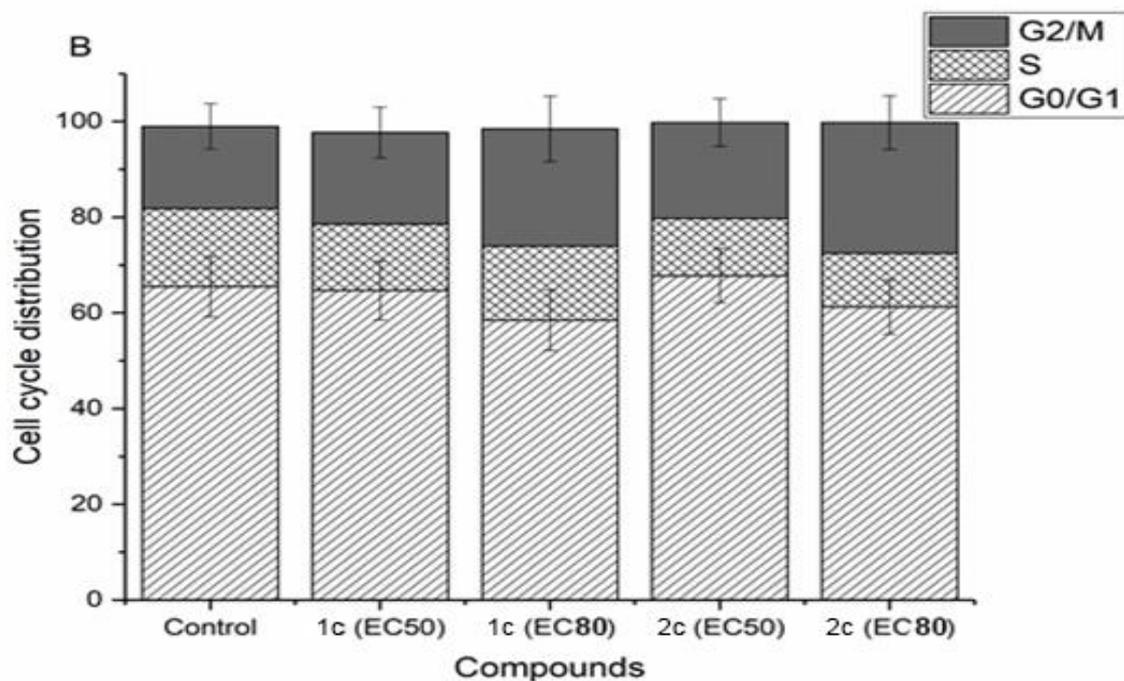
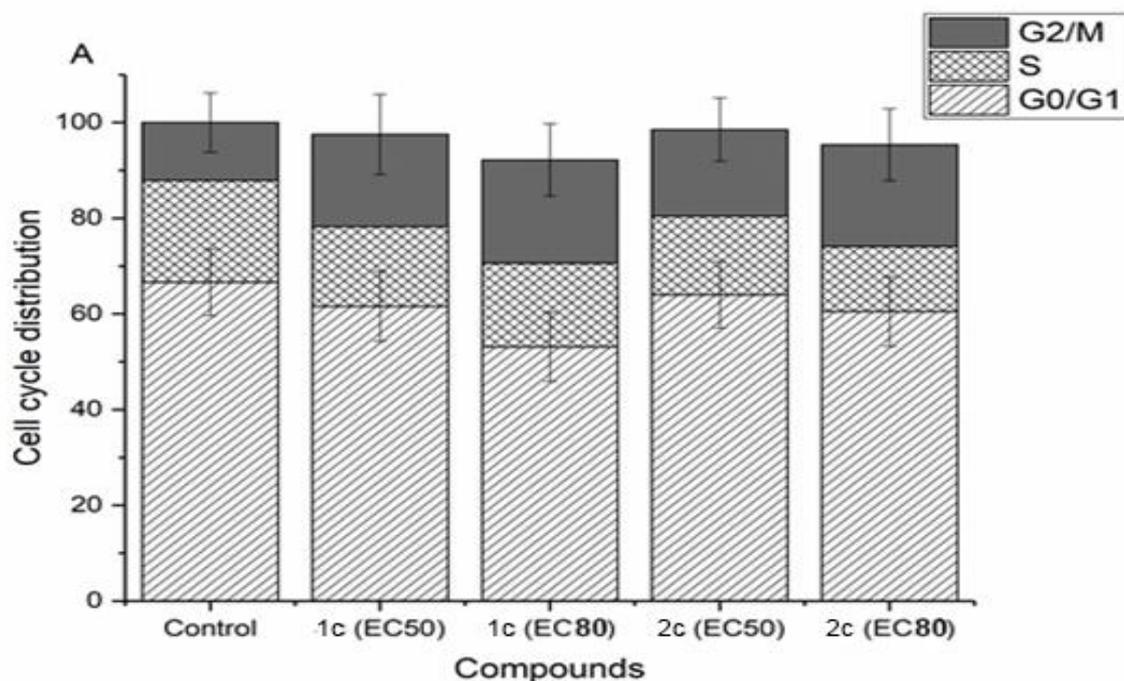


Figure S3. Cell cycle. (A) A549 cell line, (B) A431 cell line

Cell migration assay [S10]. The wound healing assay mimics tumor cell migration *in vivo*. A549 cancer cells were seeded in 24-well cell culture plates and cultured to a confluent monolayer. A 200 μ l pipette tip was used to scratch a wound on the midline of the culture well, and cell layer was then washed twice with RPMI 1640 and were treated with the control RPMI 1640 medium or medium containing 20 ng ml^{-1} EGF and 5-cyanopyrimidines derivate for 48 h. At 0 and 48 h,

images were captured on a microscope Leica DMIL (Leica, Mannheim, Germany) with Canon 60D camera (Canon Inc., Tokyo, Japan) at a magnification of $\times 100$. The migration ability of the cells was assessed by counting the wound closure rate (Figure S4).

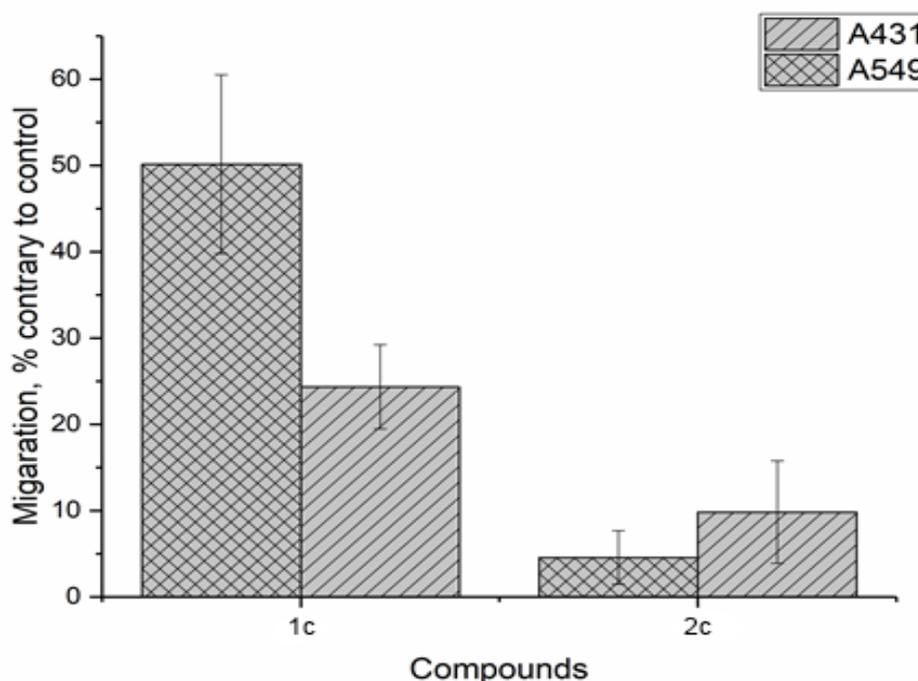


Figure S4. Migration assay at A431 and A549 cell line

Colorimetric detection of EGFR, Akt and ERK1/2 levels. To evaluate EGFR, Akt, ERK1/2 and its phosphorylated forms expression we performed colorimetric detection using Pierce EGFR Colorimetric In-Cell ELISA Kits for EGFR, Akt, ERK1/2 (Thermo scientific). Briefly, A431 cells were placed in 96-well plate at 10,000 cells per well and incubated overnight at 37°C in 5% CO₂. After cells were treated with selected 5-cyanopyrimidine derivatives at IC₅₀ concentration in complete DMED containing 20 ng ml⁻¹ EGF for 24 h. Cells were fixed with 4% formaldehyde for 15 minutes and washed twice with 100µl per well of 1X TBS. Cells were treated with 100µL/well of 1X Permeabilization buffer for 15 minutes and washed once with 1X TBS. Cells were treated of Quenching solution and incubate at room temperature for 20 min, washed once with 1X TBS and blocked with 100 µl per well of Blocking buffer at room temperature for 30 min. Cells were incubated with 50 µl per well of primary antibody overnight at 4°C. Later in well was added diluted HRP conjugate and TMB substrate. After stopping reaction absorbance was measured at 450nm within 30 minutes of stopping the reaction.

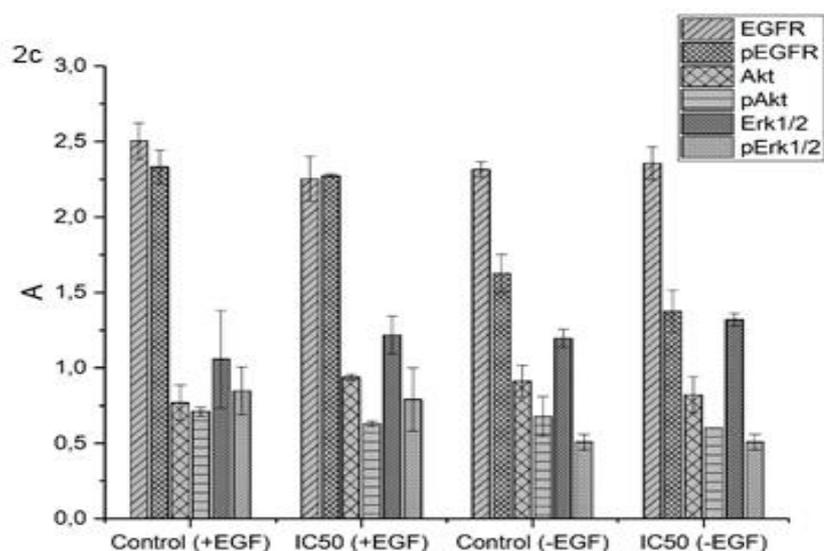
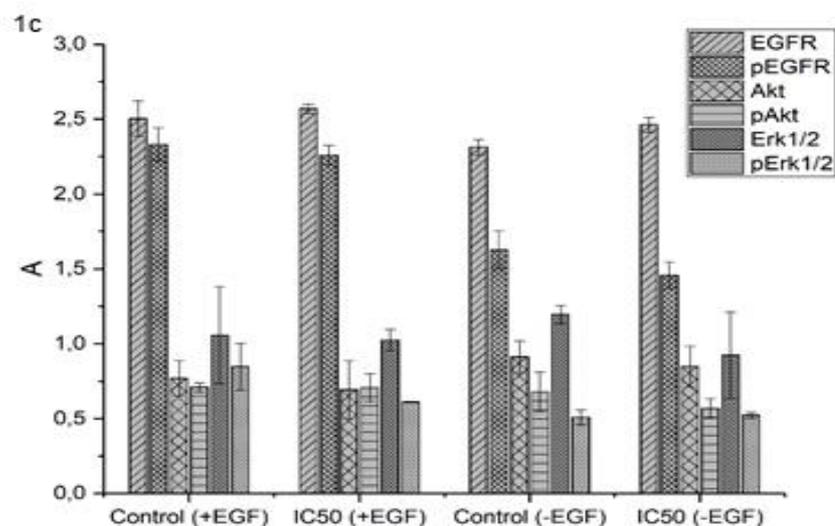


Figure S5. In-Cell ELISA assay for determining EGFR, pEGFR, Akt, pAkt, ERK1/2 and pERK1/2 levels

Generation of Spheroids. Spheroids were generated by plating A549 cancer cells at 1×10^4 cells per well into U-bottom Ultra Low Adherence (ULA) 96-well plates (Corning, USA) at 200 μ l per well for formation of a single spheroid. After the cells were introduced into the plate, compounds **1c** and **2c** were added in a non-cytotoxic concentration and incubated for 72 hours at 37°C, 5% CO₂. Images were captured InCell Analyzer 6000 at magnification 4x and InCell Investigator software (GE Healthcare).

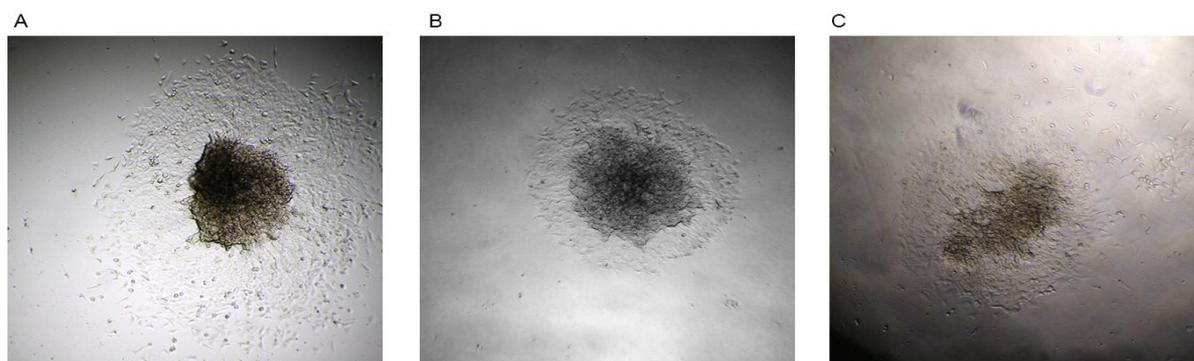


Figure S6 Generation of tumor spheroids in culture (*A* – Control, *B* – **1c**, *C* – **2c**)

References

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NMR spectra

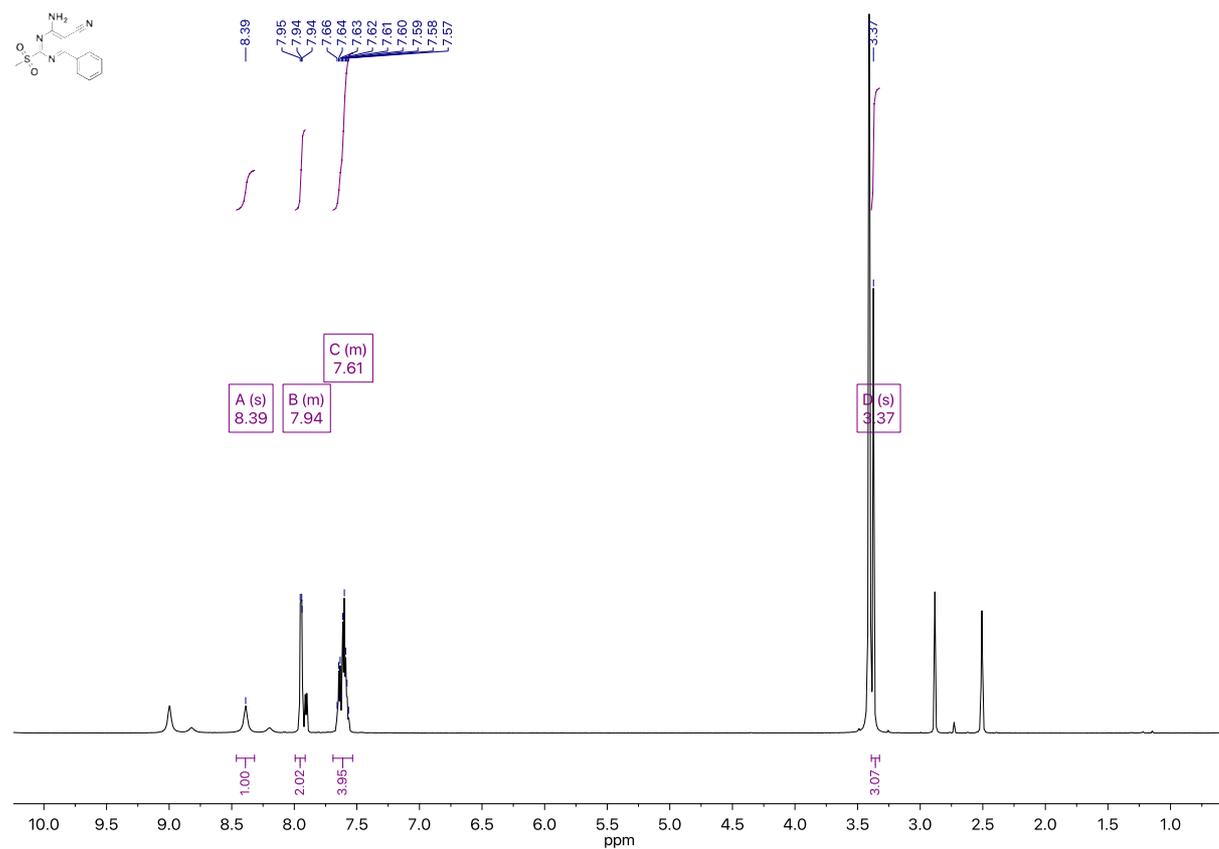


Figure S7. The ¹H NMR spectrum of **1a**.

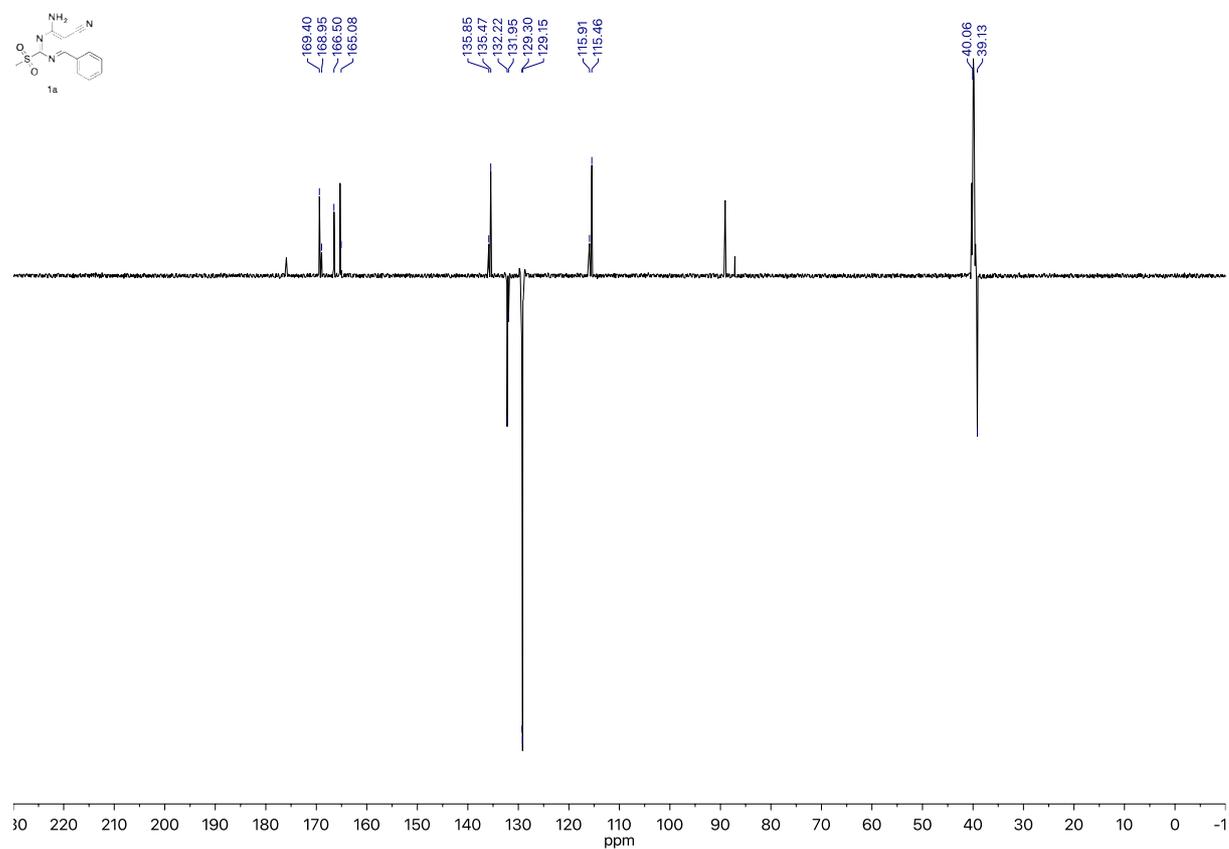


Figure S8. The ¹³C NMR spectrum of **1a**

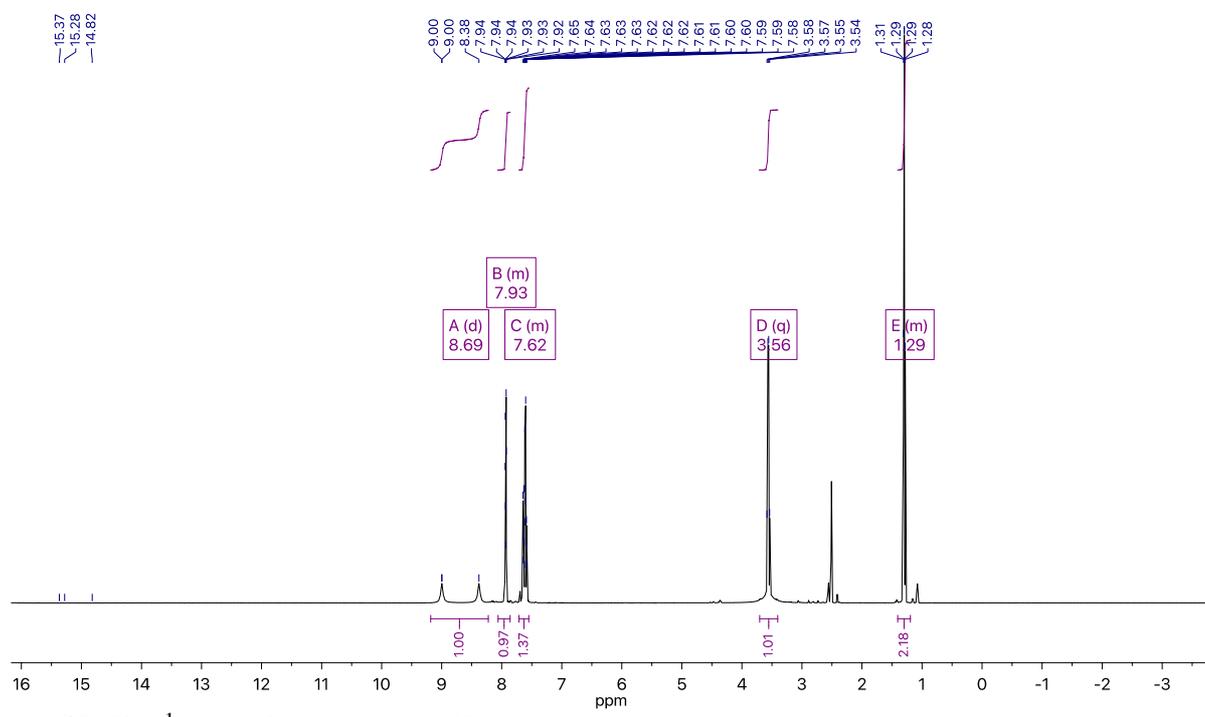
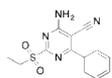


Figure S9. The ¹H NMR spectrum of **1b**.

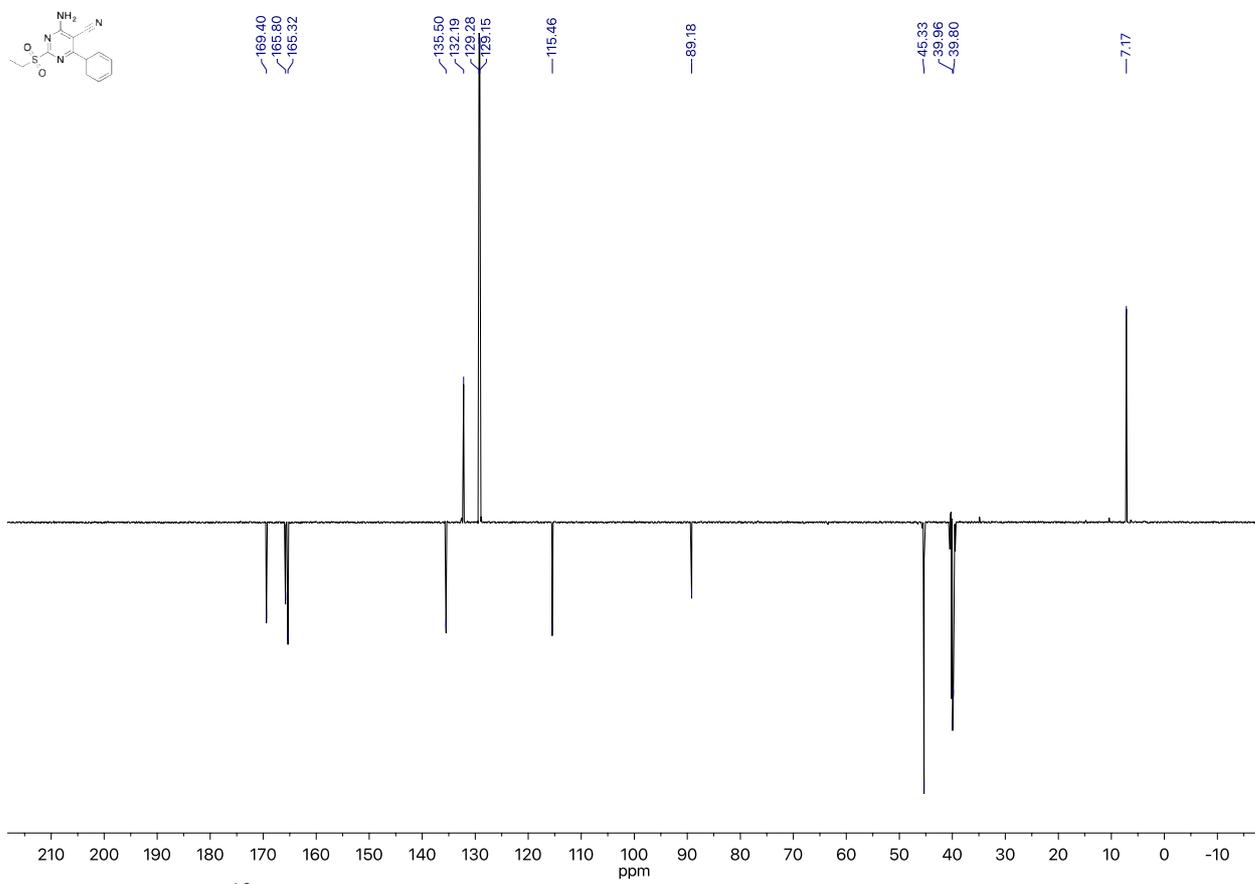
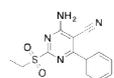


Figure S10. The ¹³C NMR spectrum of **1b**.

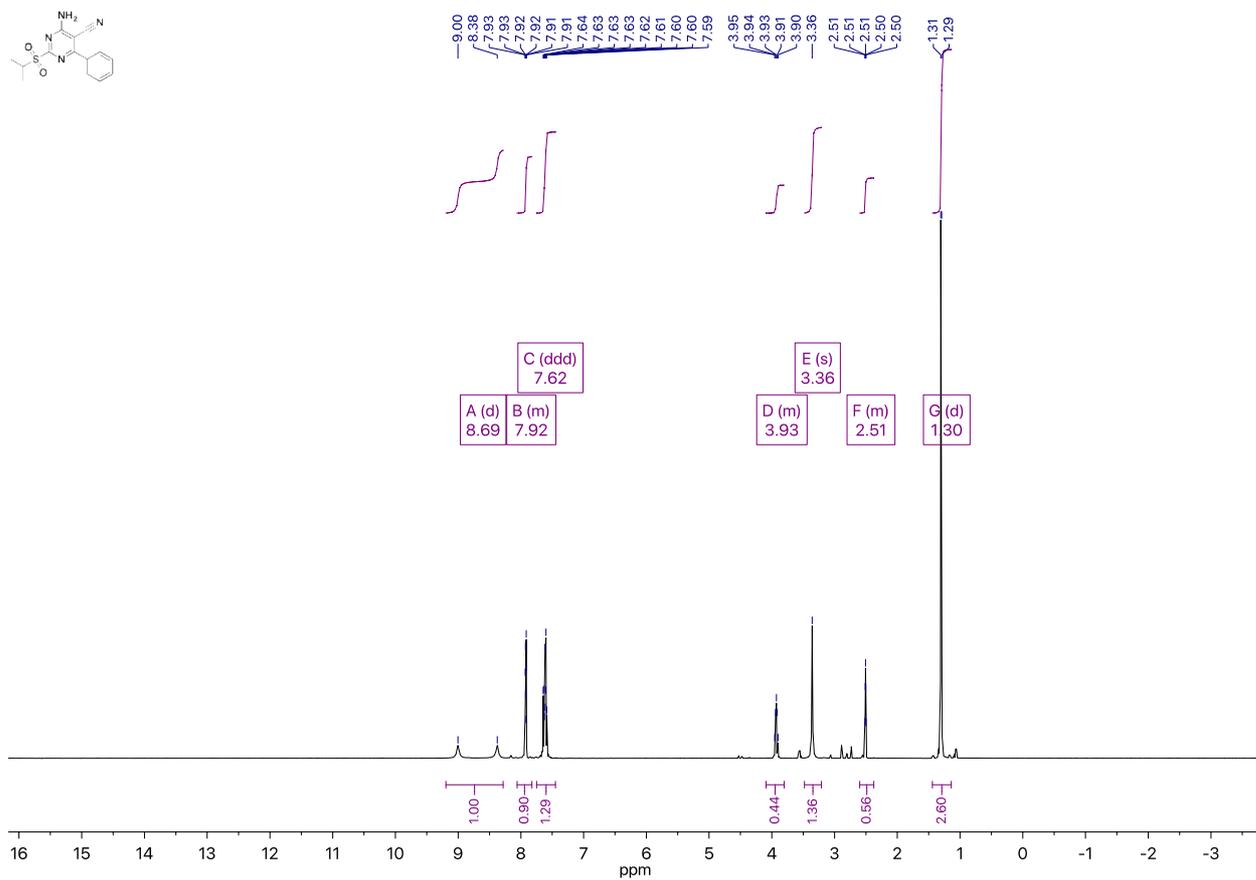


Figure S11. The ¹H NMR spectrum of **1c**.

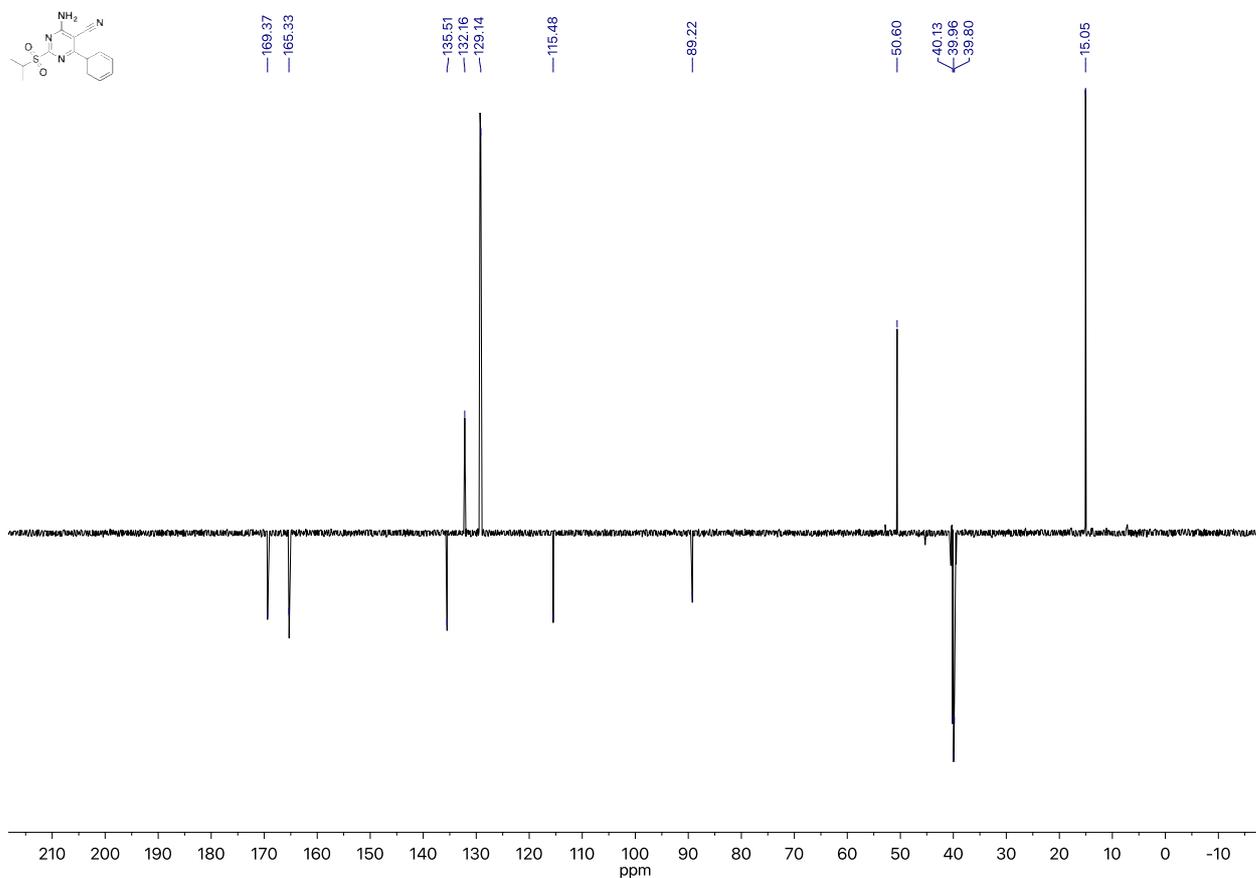


Figure S12. The ¹³C NMR spectrum of **1c**.

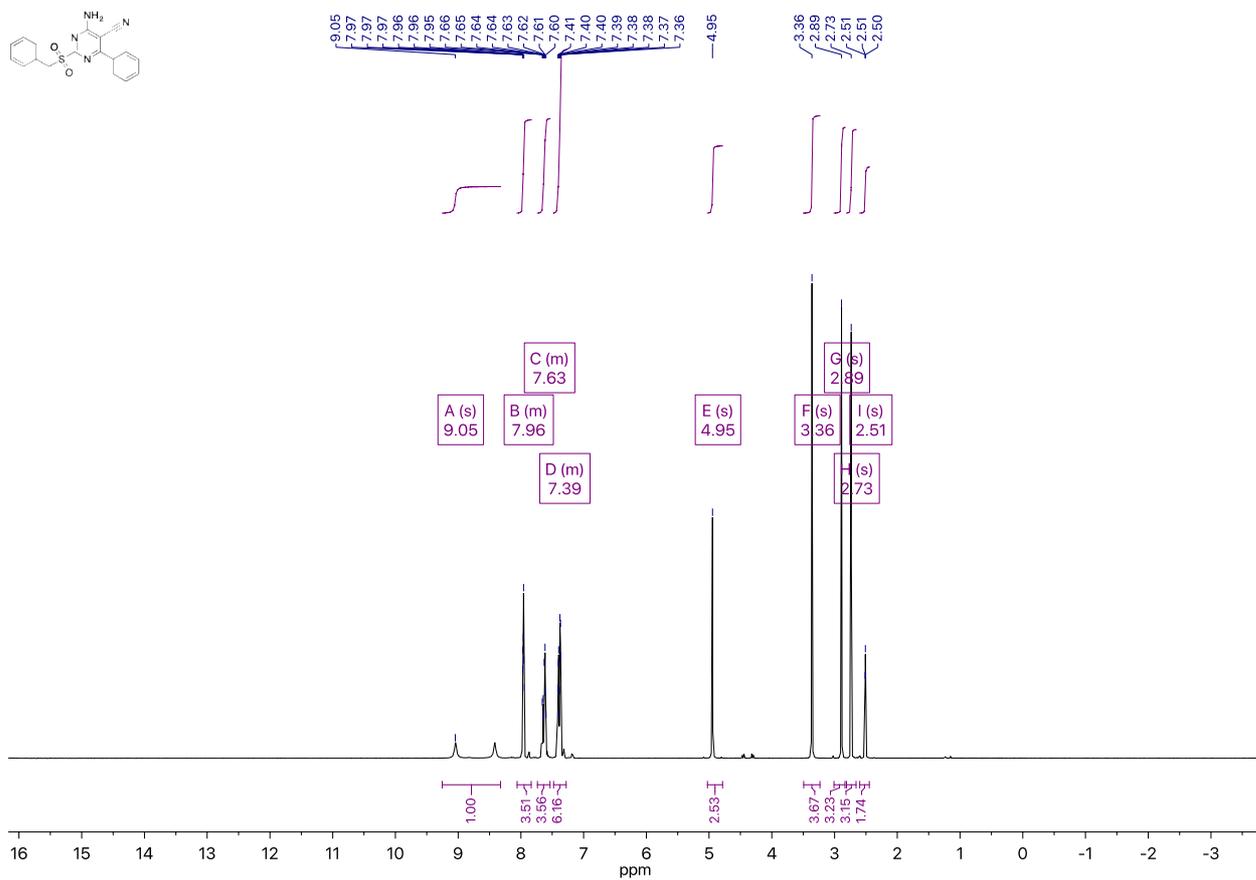


Figure S13. The ¹H NMR spectrum of **1d**.

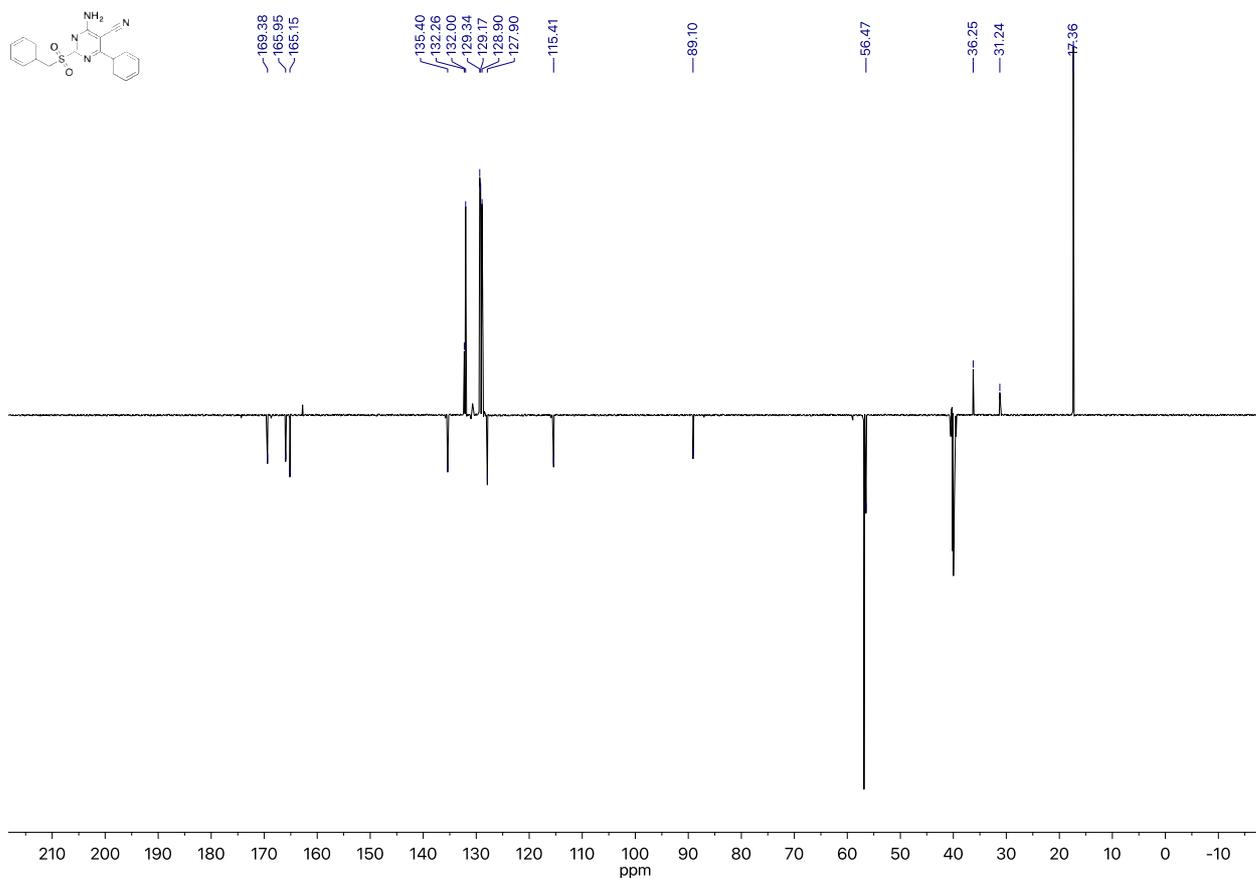


Figure S14. The ¹³C NMR spectrum of **1d**.

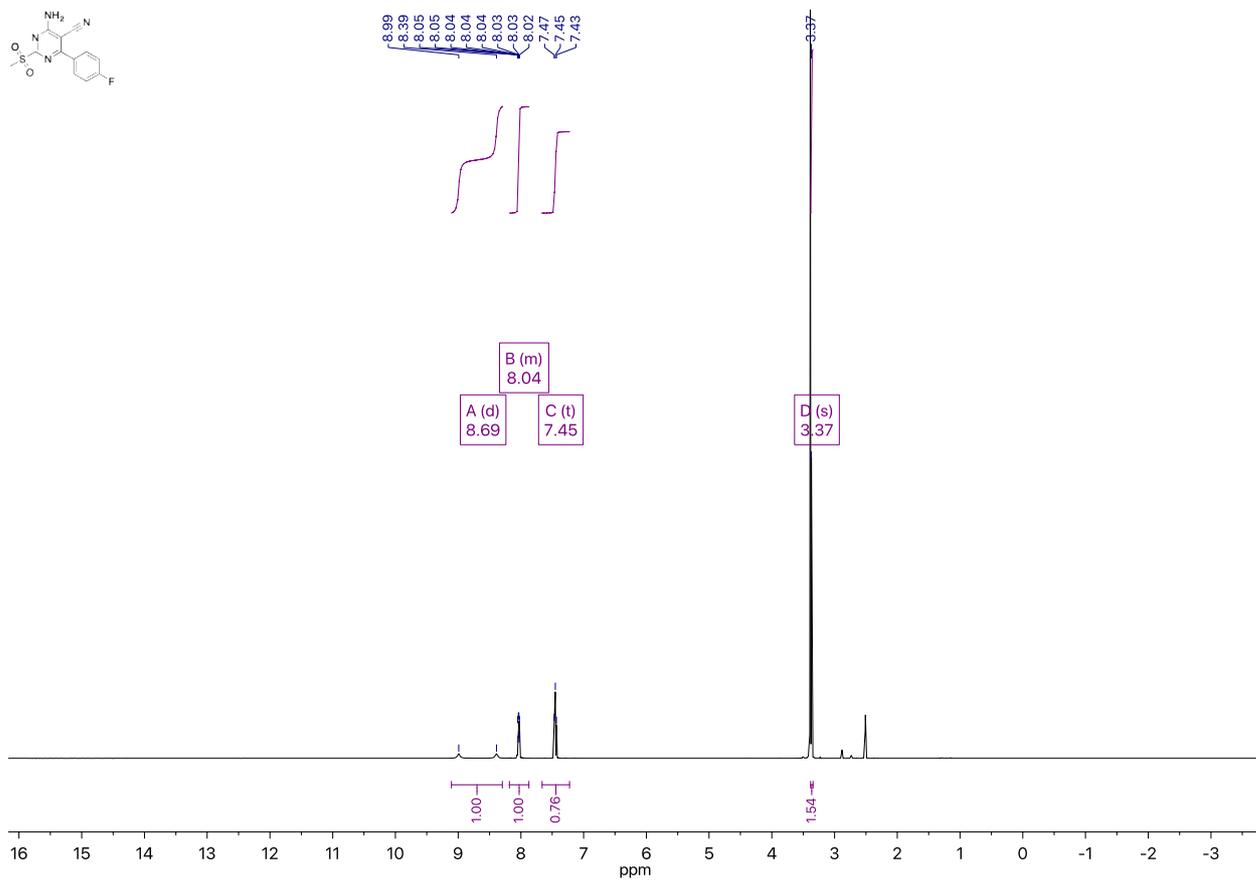


Figure S15. The ¹H NMR spectrum of **1e**.

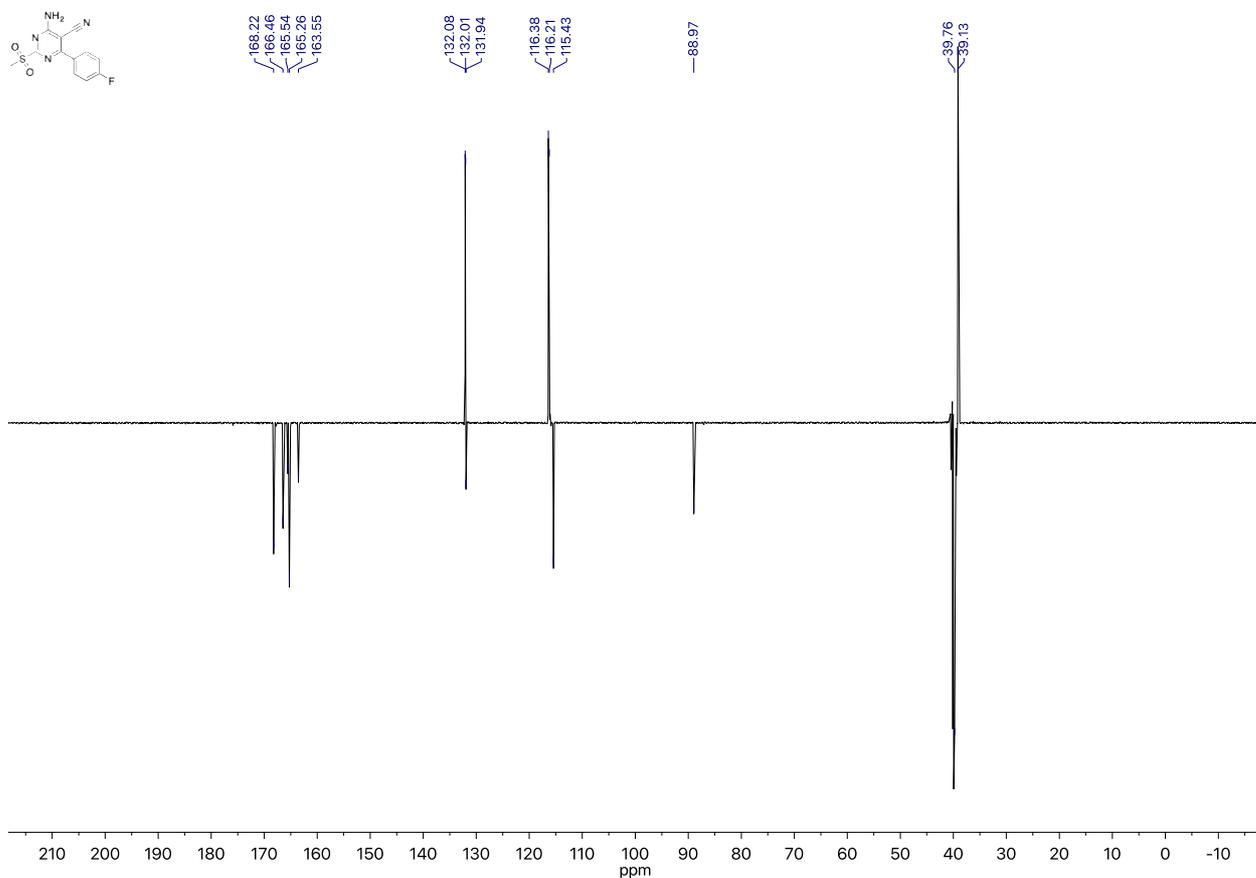


Figure S16. The ¹³C NMR spectrum of **1e**.

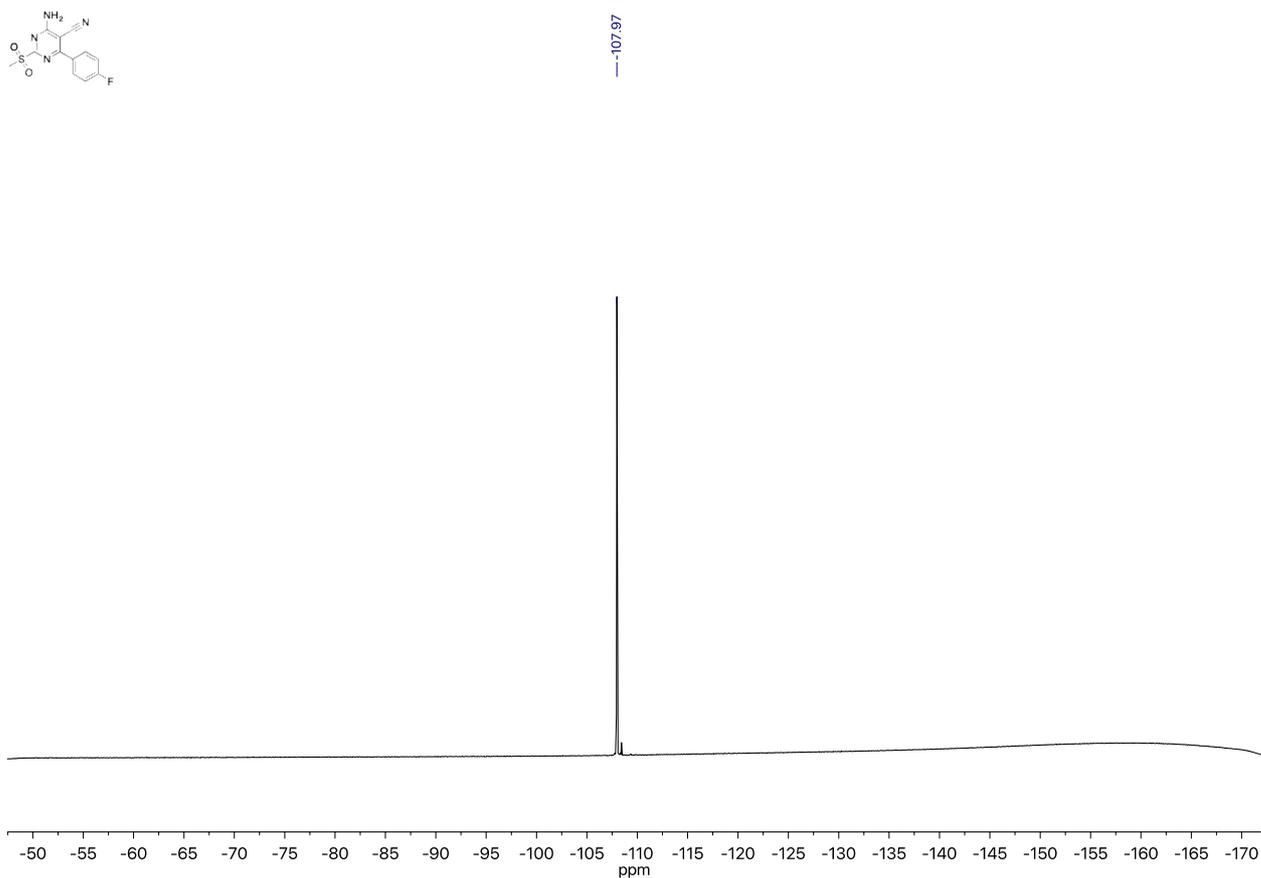


Figure S17. The ^{19}F NMR spectrum of **1e**.

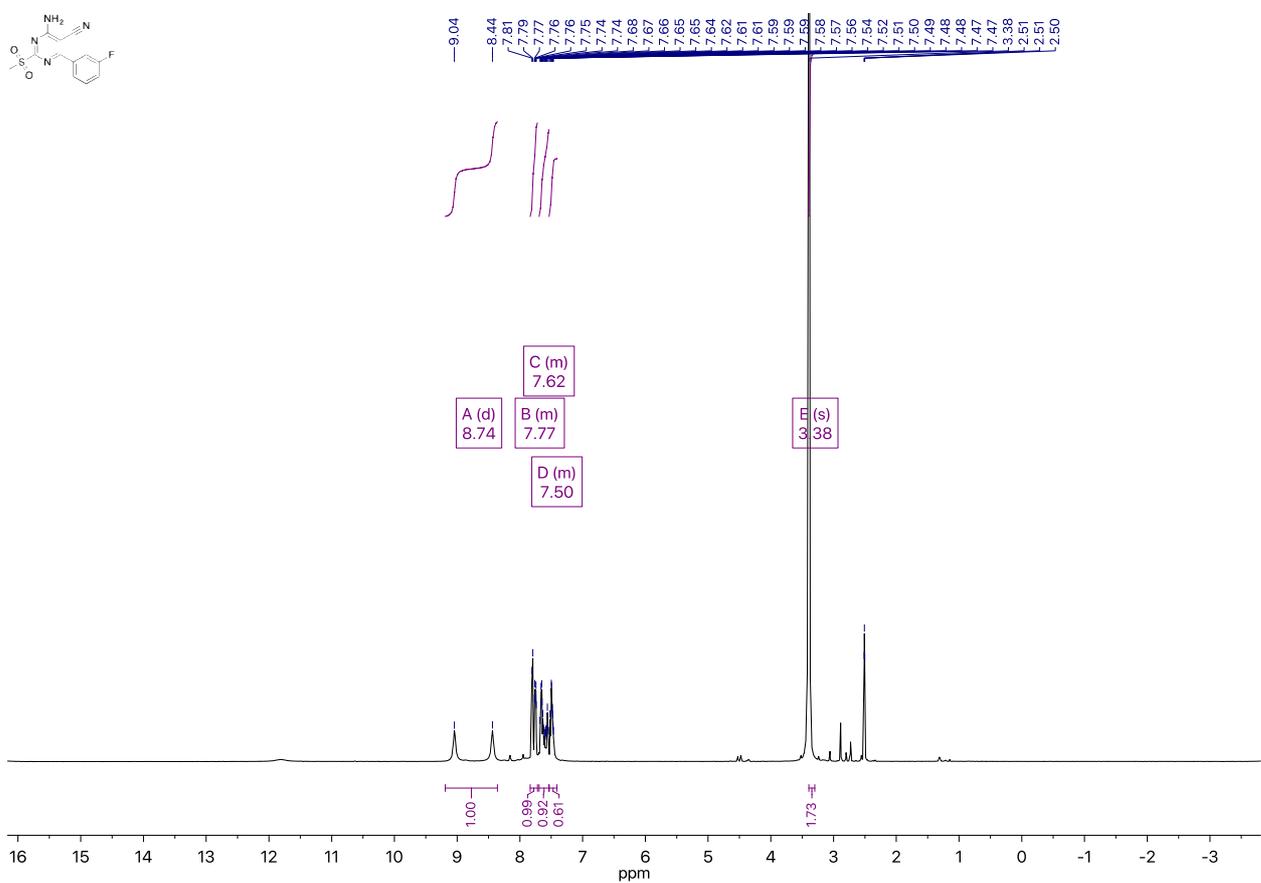


Figure S18. The ^1H NMR spectrum of **1f**.

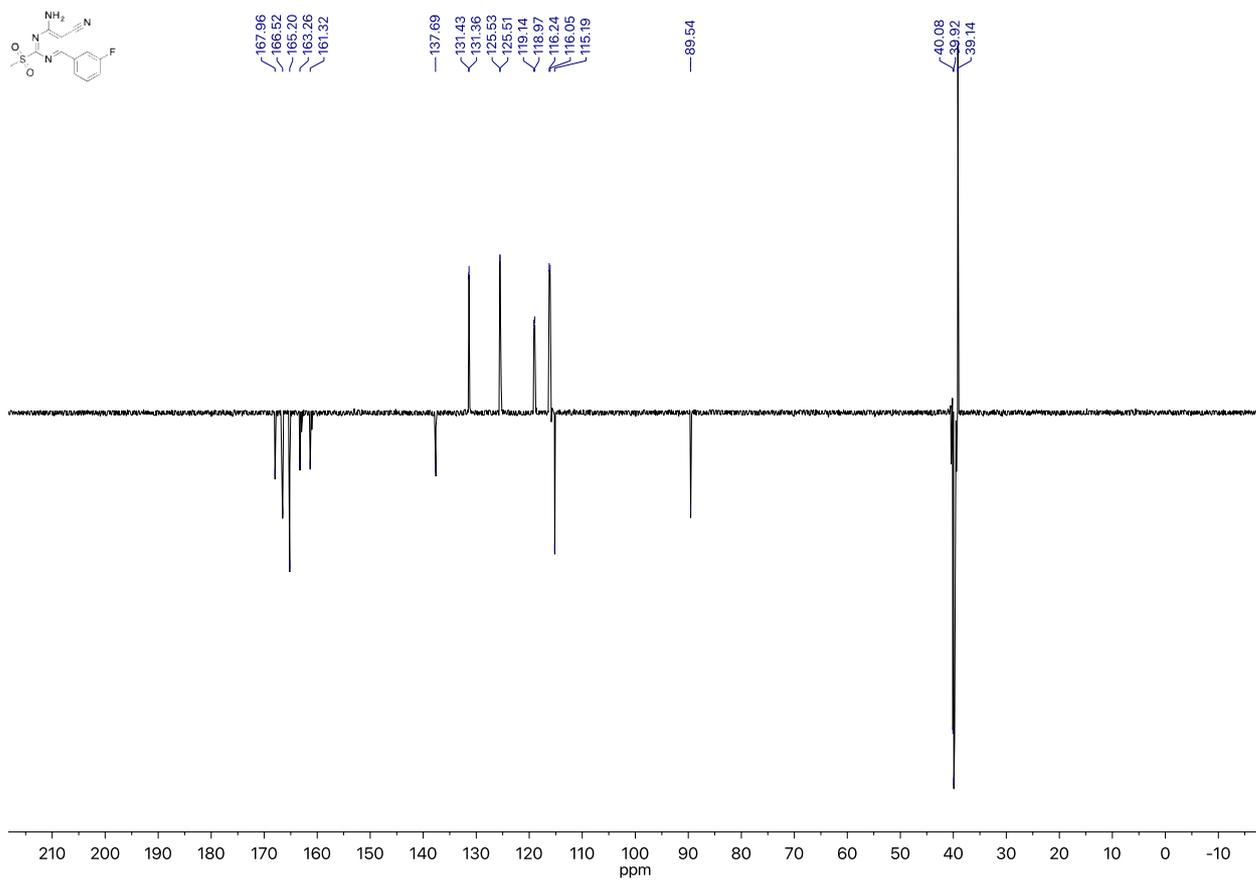


Figure S19. The ¹³C NMR spectrum of **1f**.

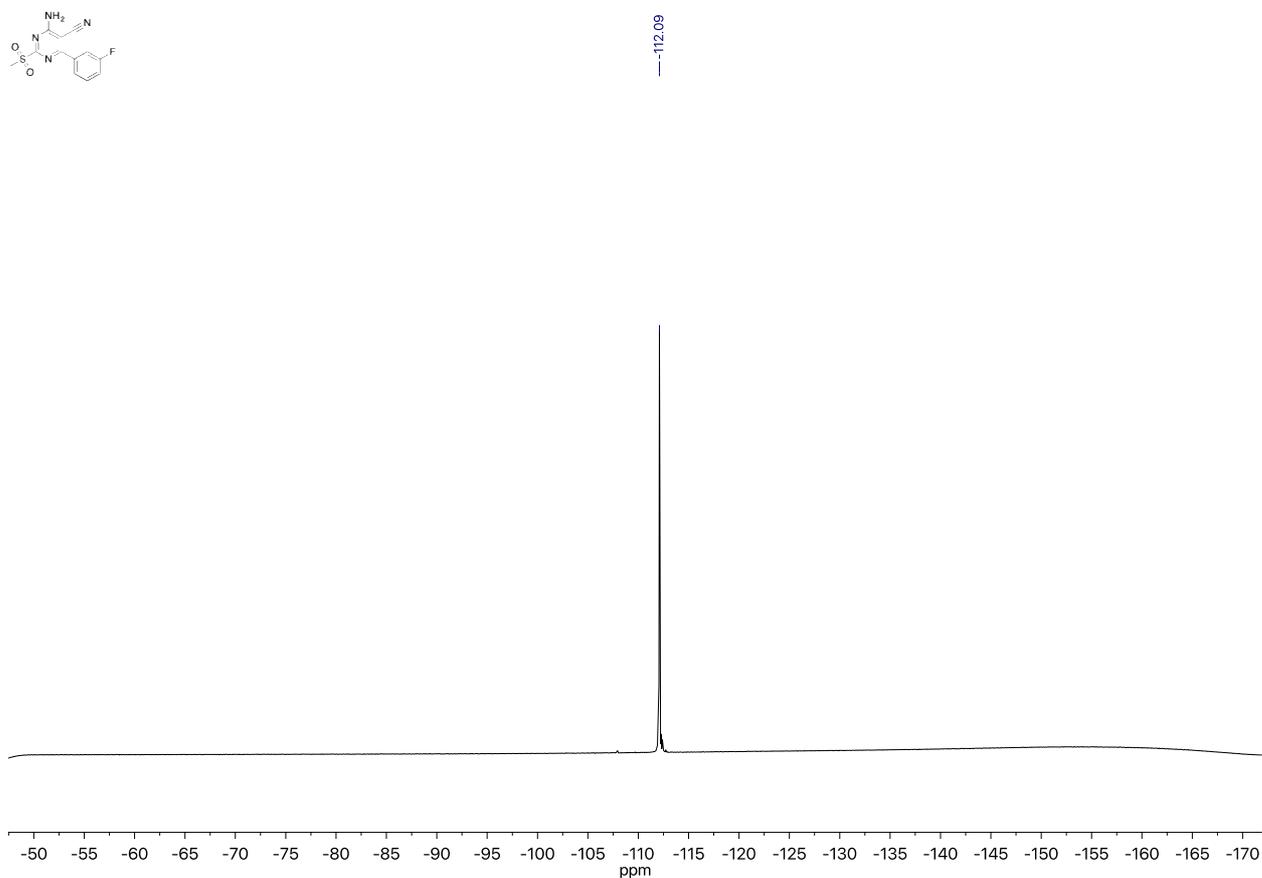
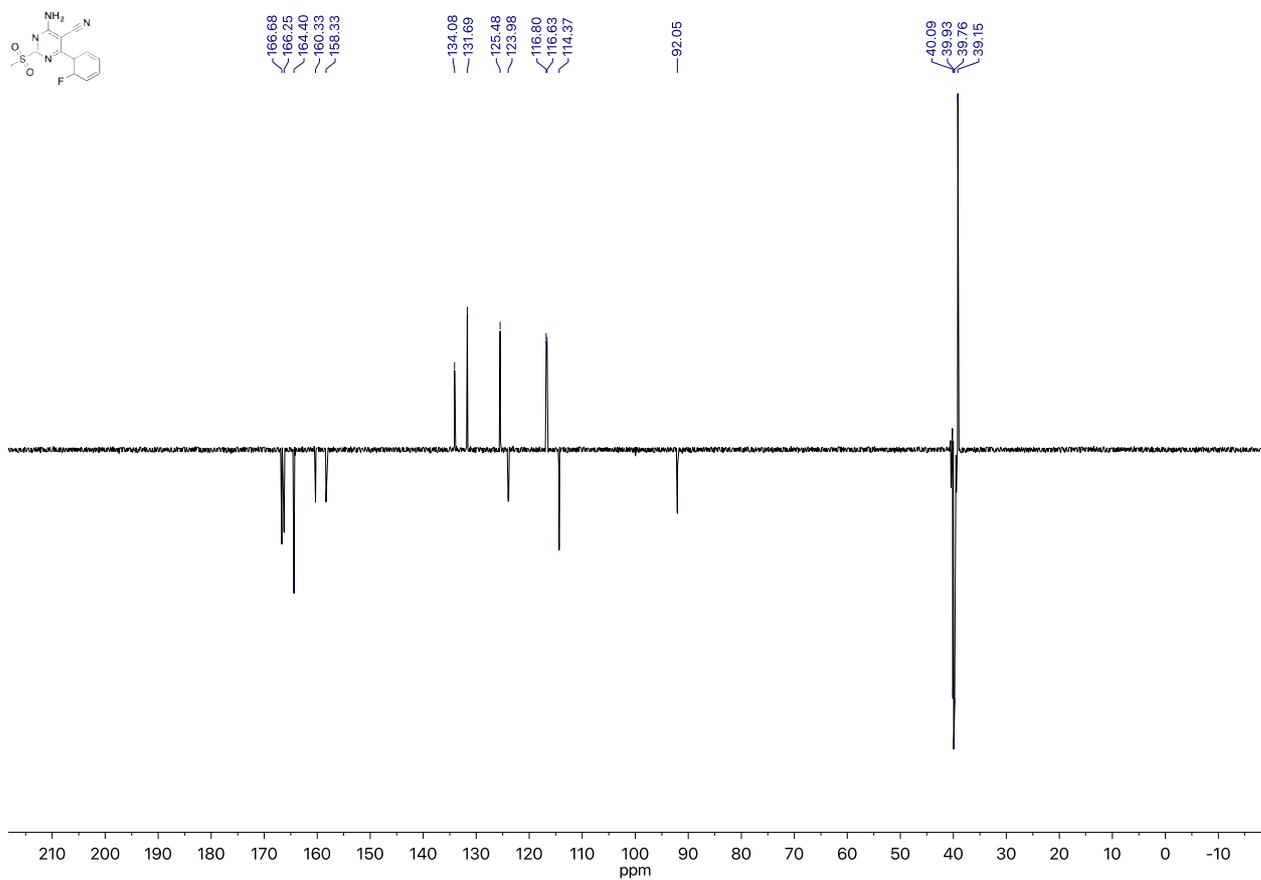
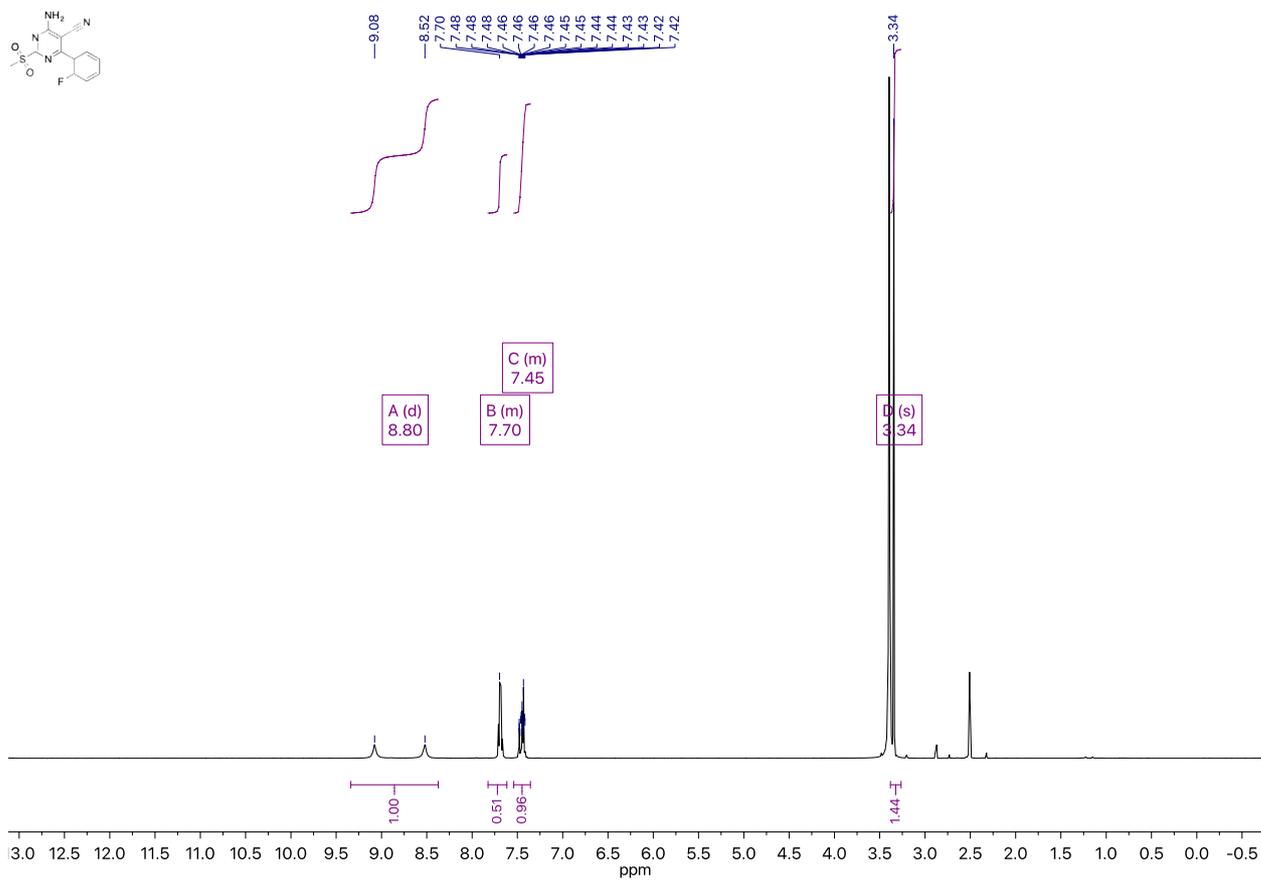


Figure S20. The ¹⁹F NMR spectrum of **1f**.





--113.47

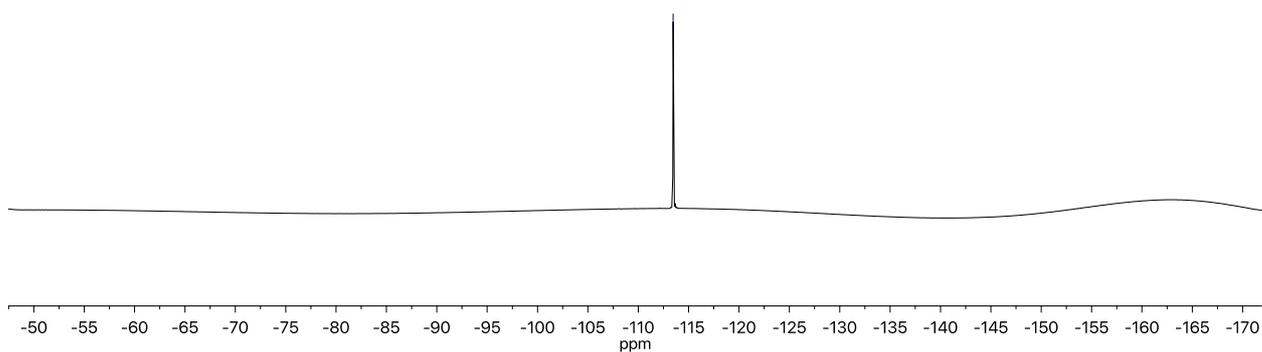


Figure S23. The ^{19}F NMR spectrum of **1g**.

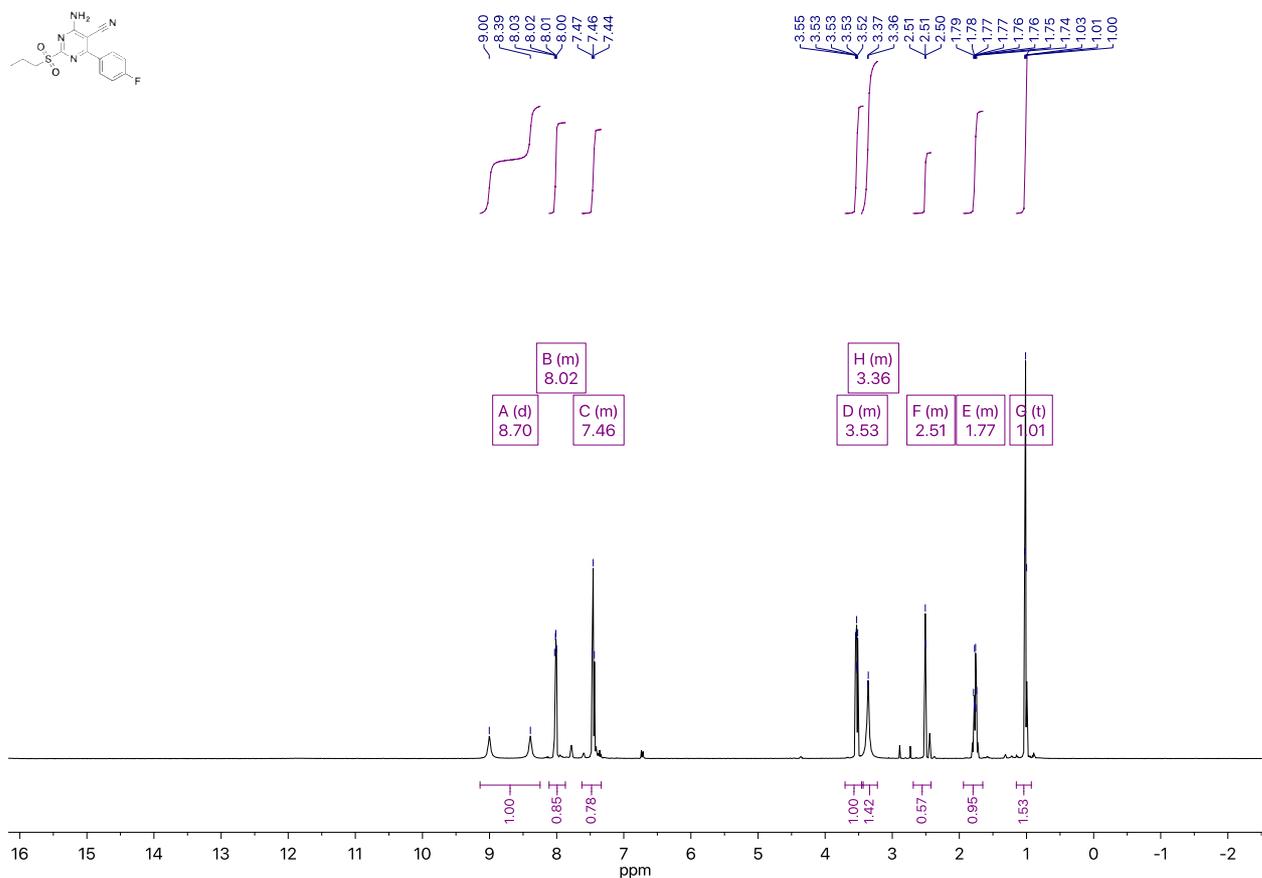
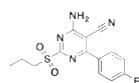


Figure S24. The ^1H NMR spectrum of **1h**.

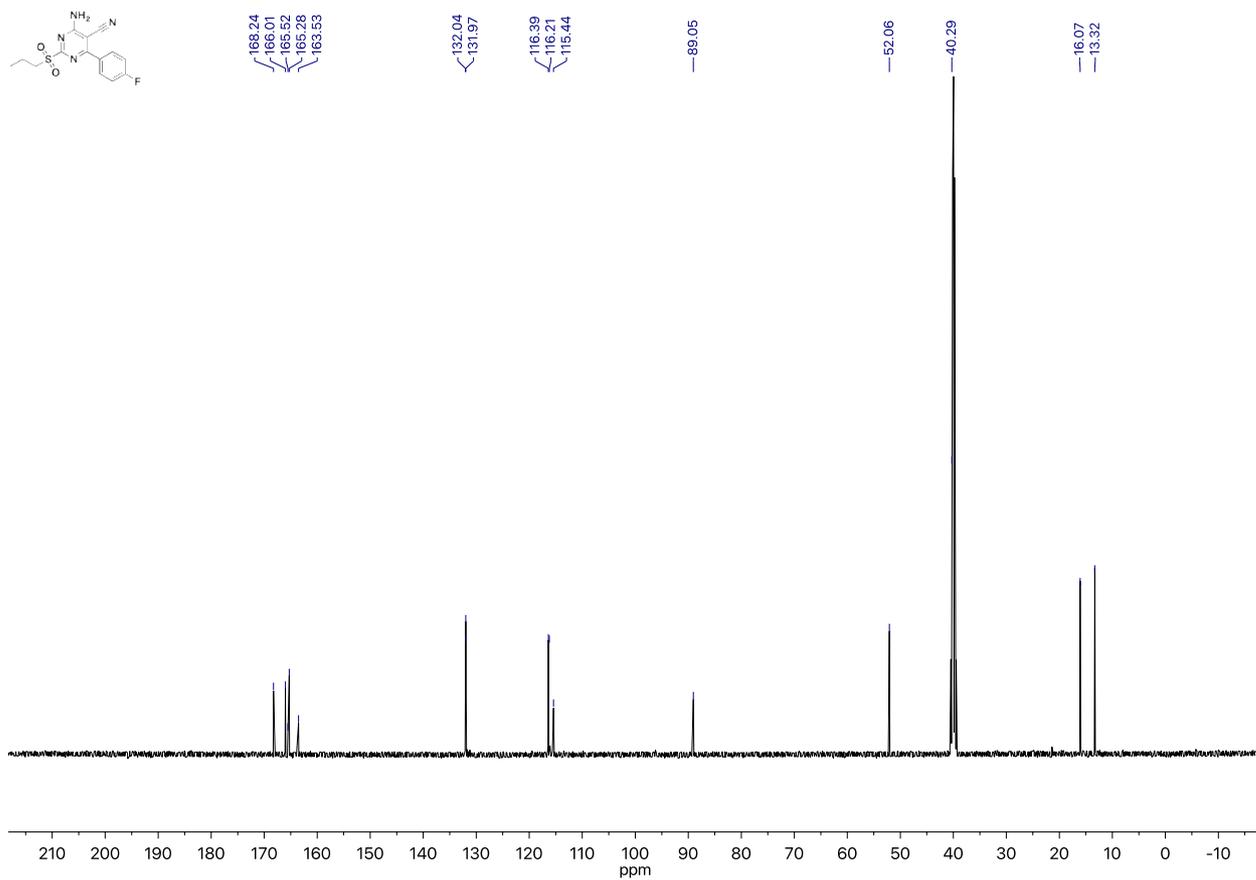


Figure S25. The ^{13}C NMR spectrum of **1h**.

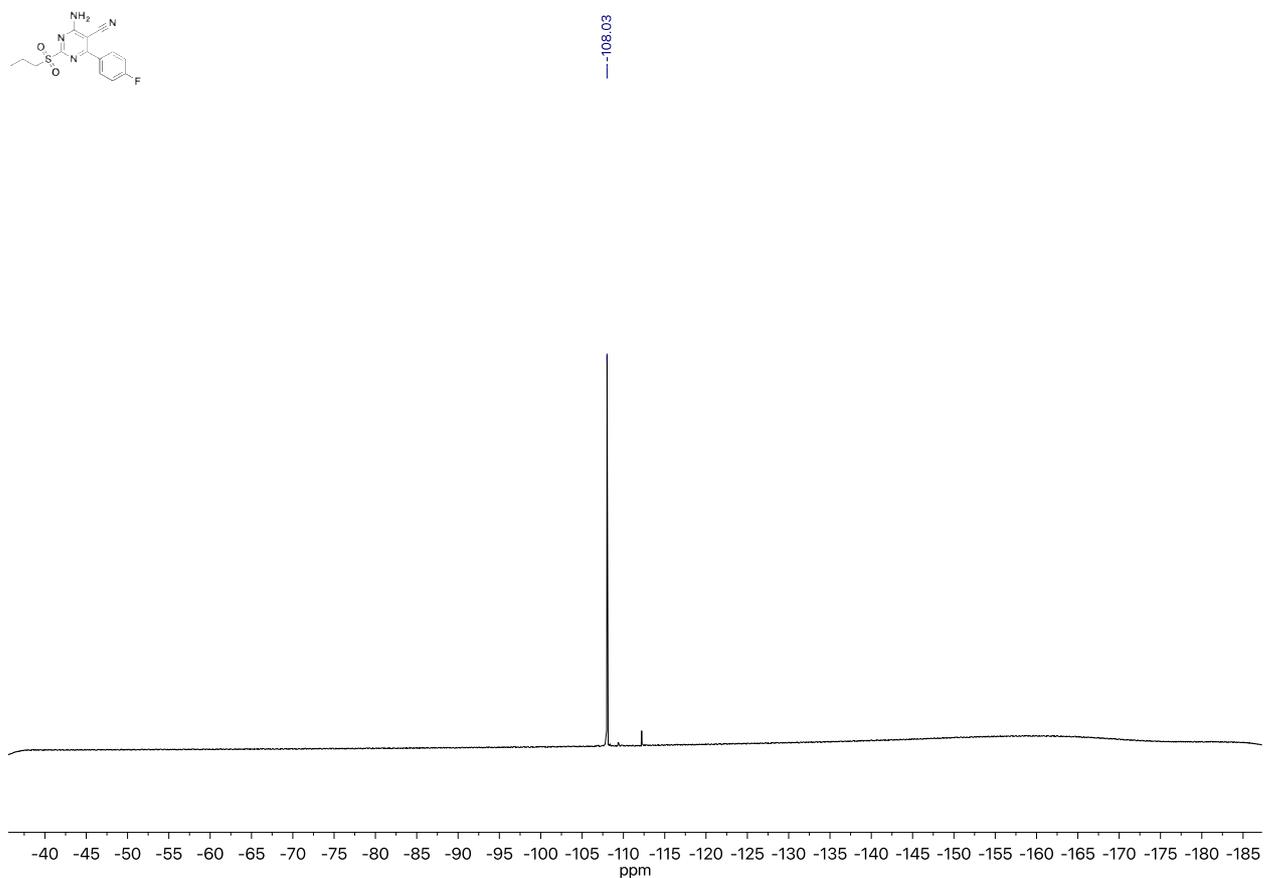


Figure S26. The ^{19}F NMR spectrum of **1h**.

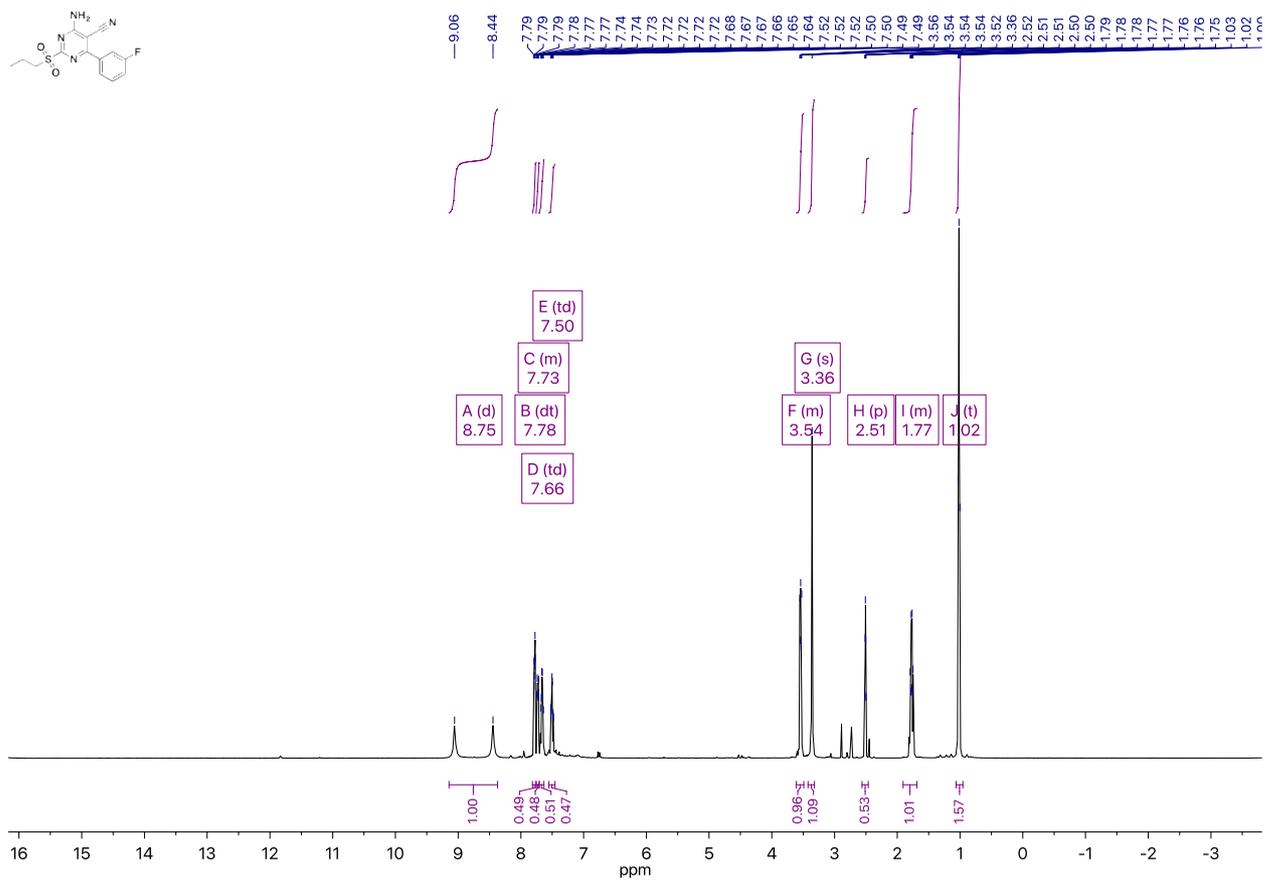


Figure S27. The ¹H NMR spectrum of **1i**.

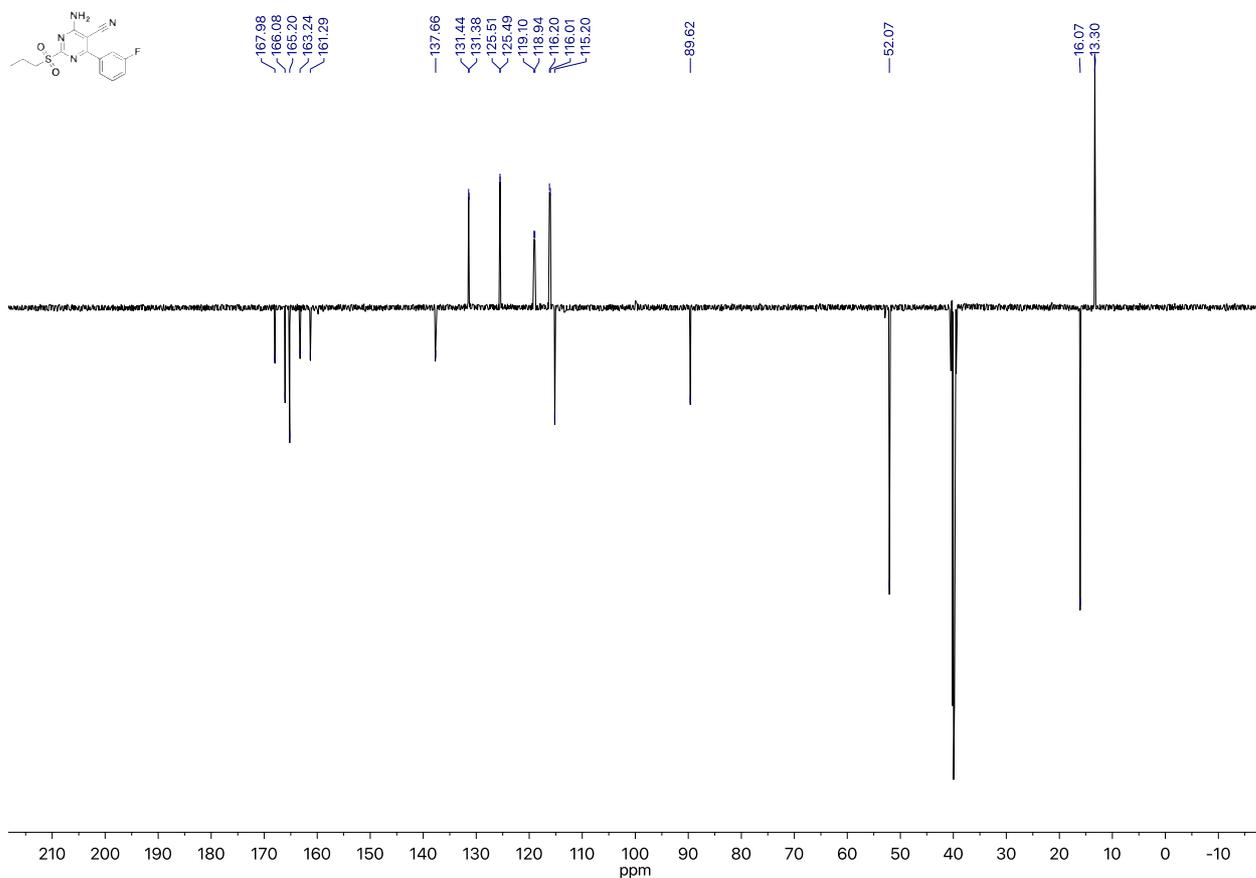


Figure S28. The ¹³C NMR spectrum of **1i**.

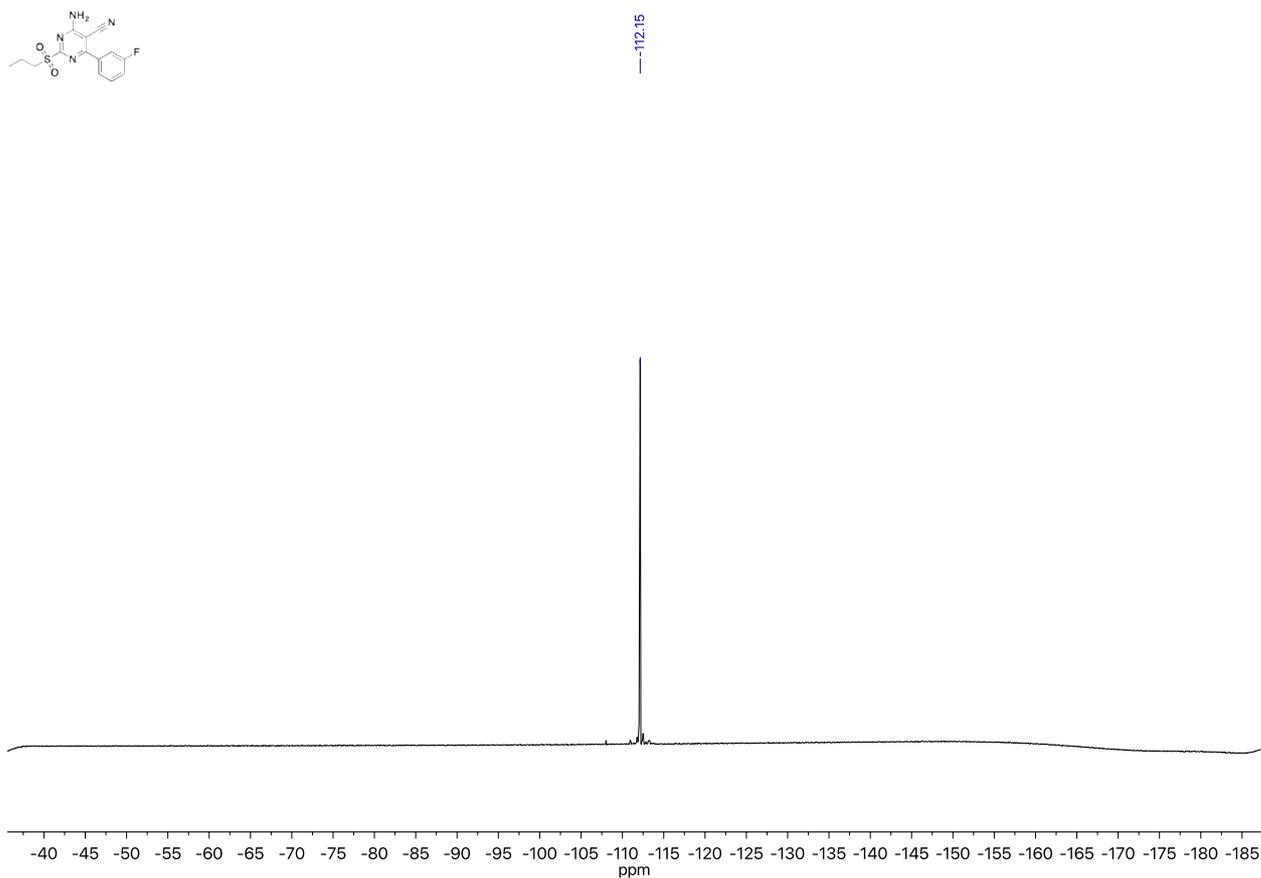


Figure S29. The ^{19}F NMR spectrum of **1i**.

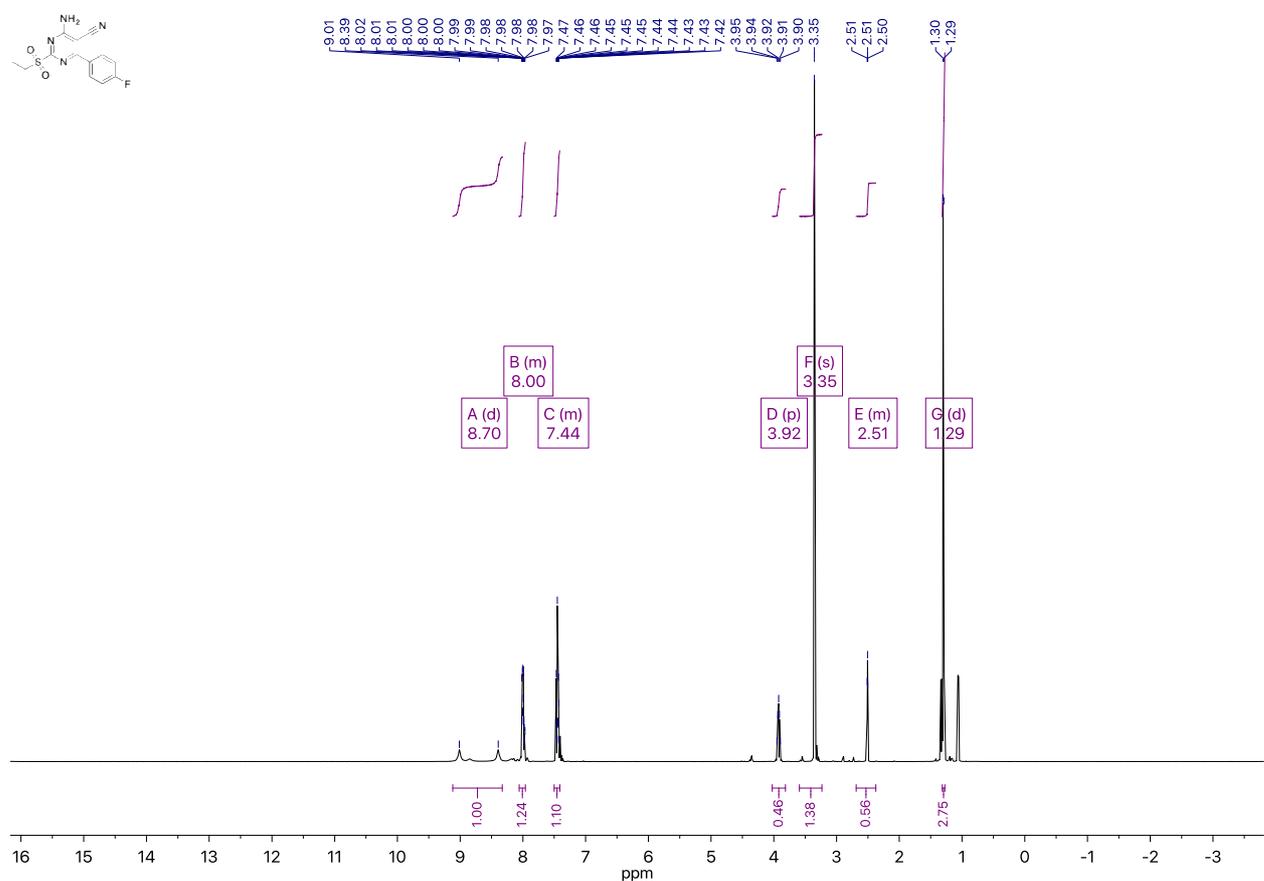


Figure S30. The ^1H NMR spectrum of **1j**.

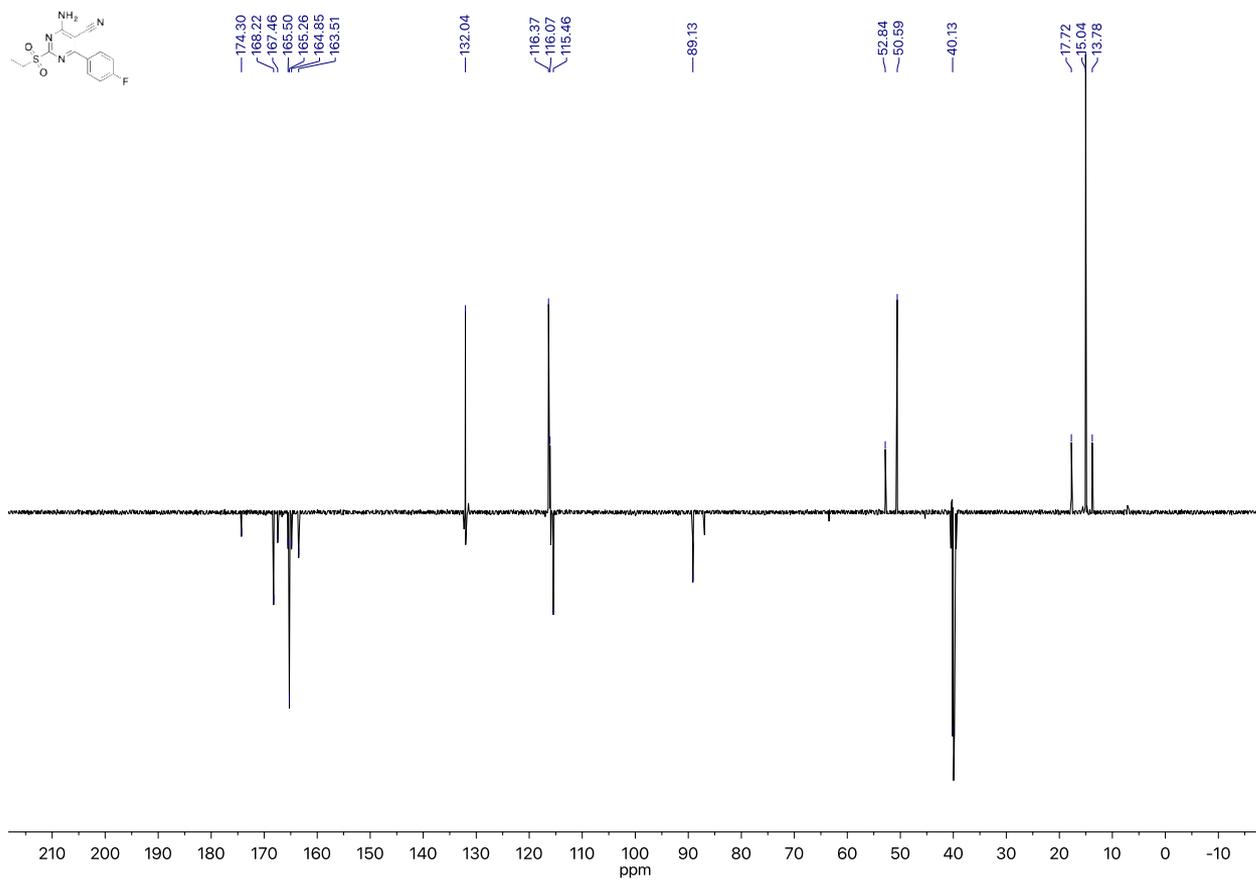


Figure S31. The ¹³C NMR spectrum of **1j**.

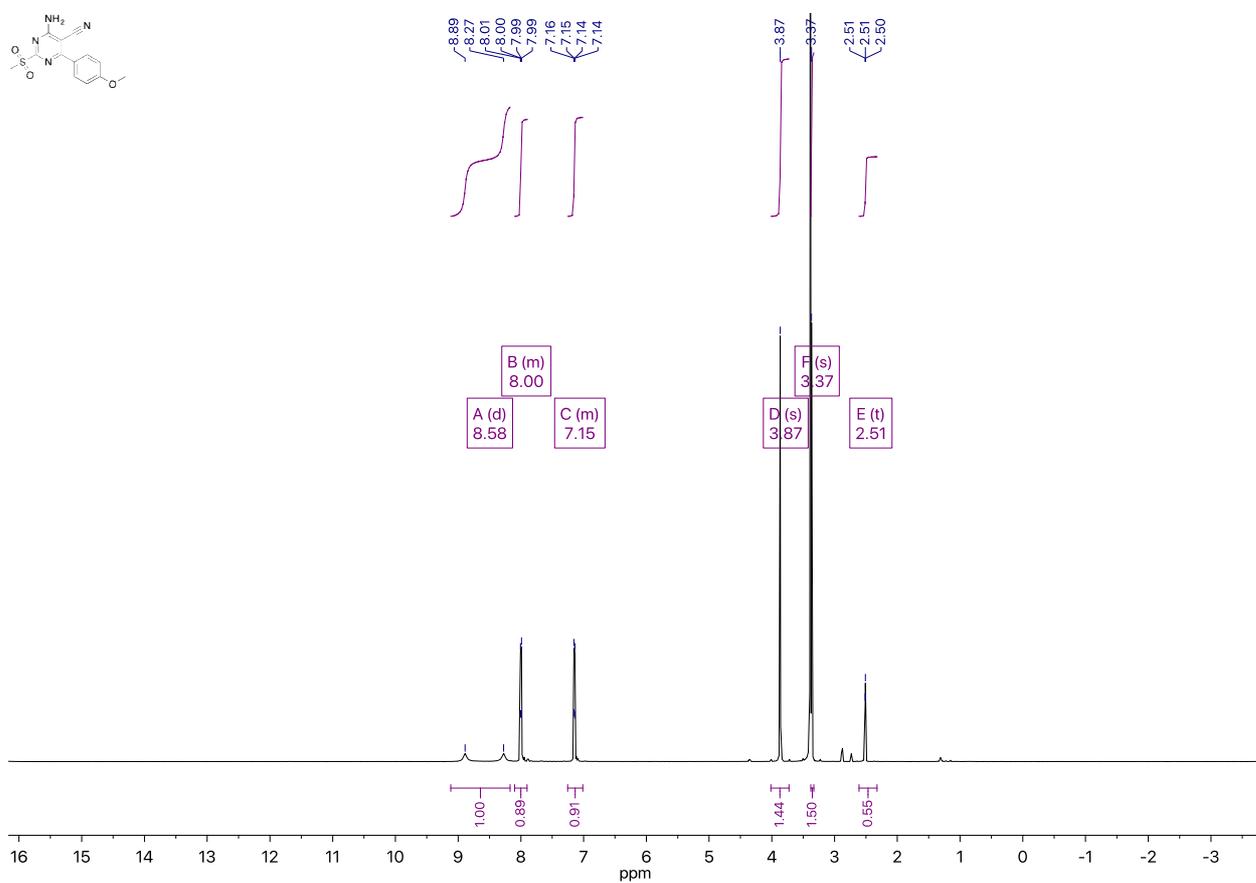


Figure S32. The ¹H NMR spectrum of **1k**.

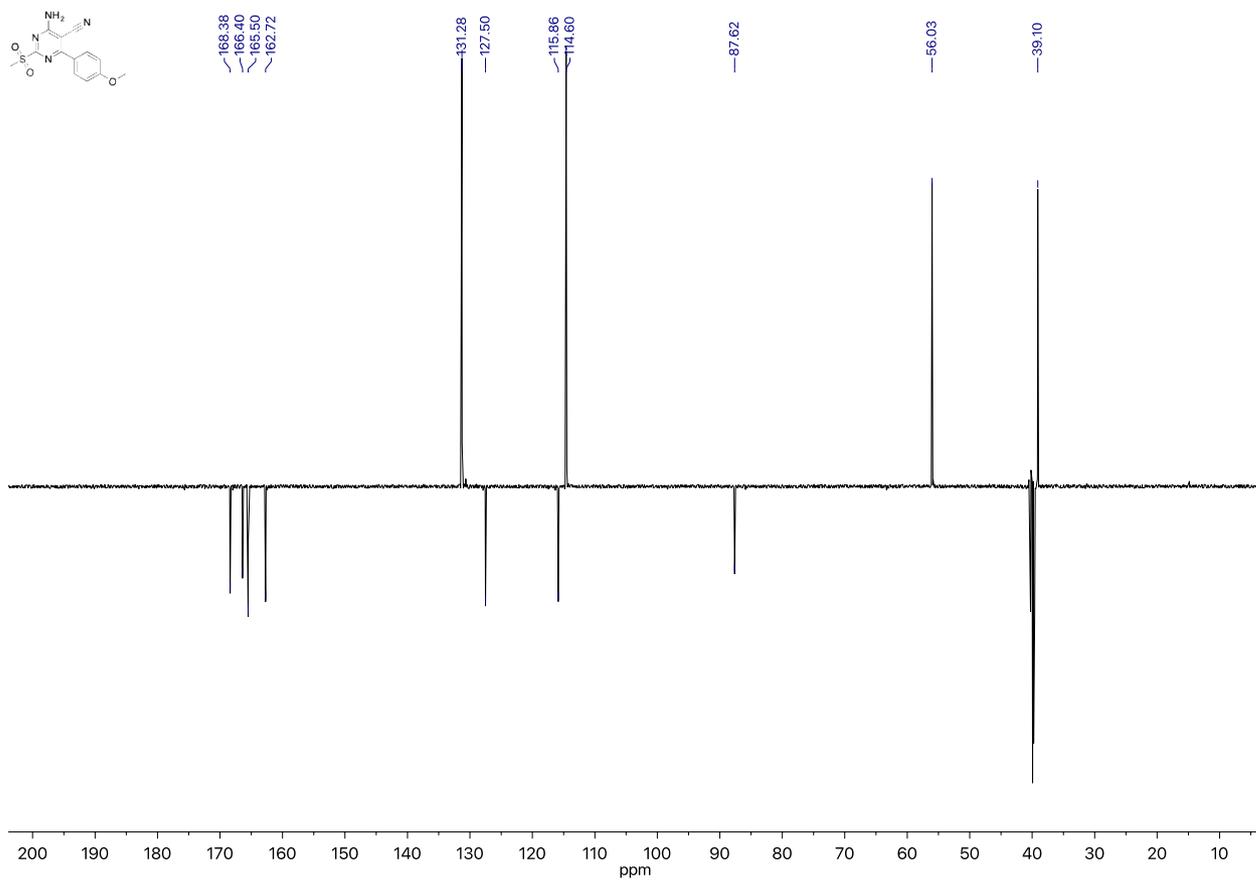


Figure S33. The ¹³C NMR spectrum of **1k**.

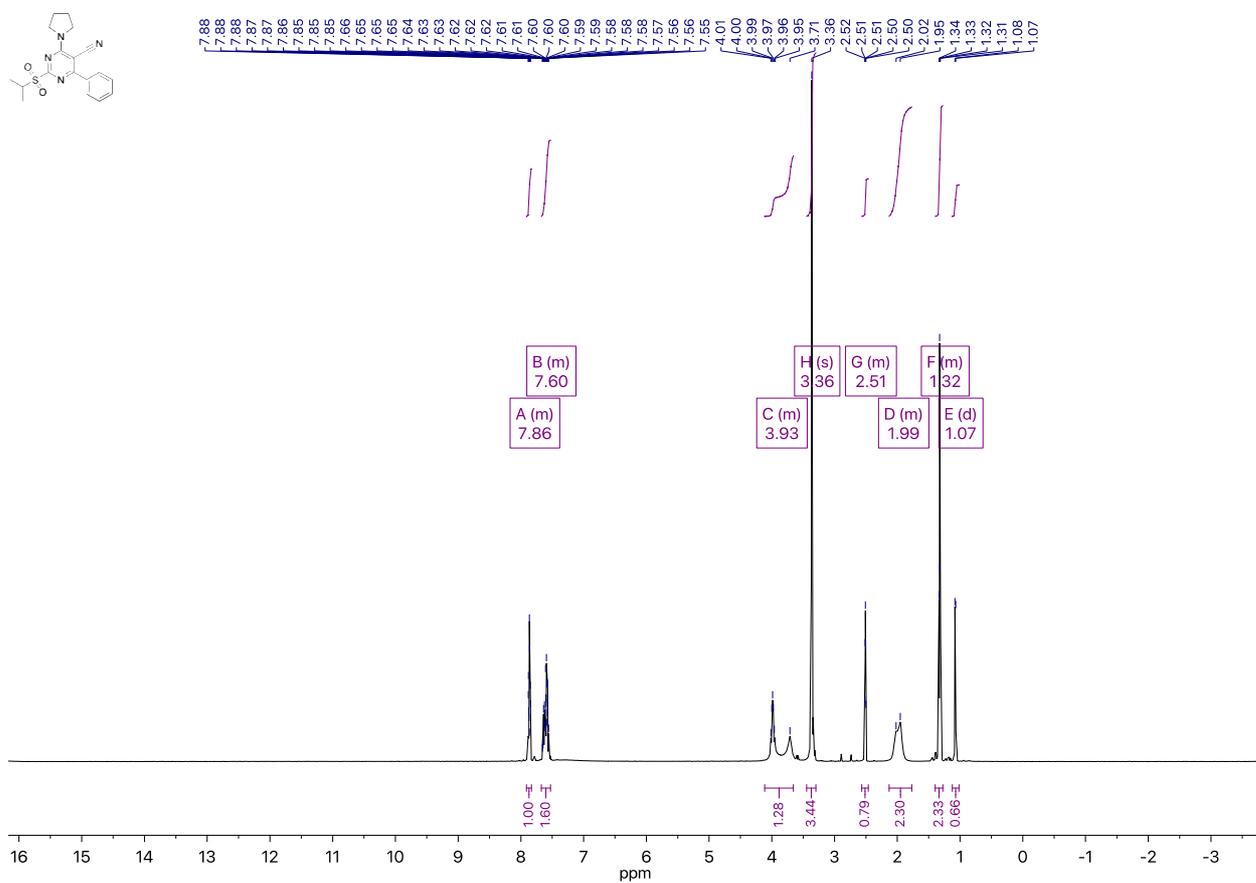


Figure S34. The ¹H NMR spectrum of **2a**.

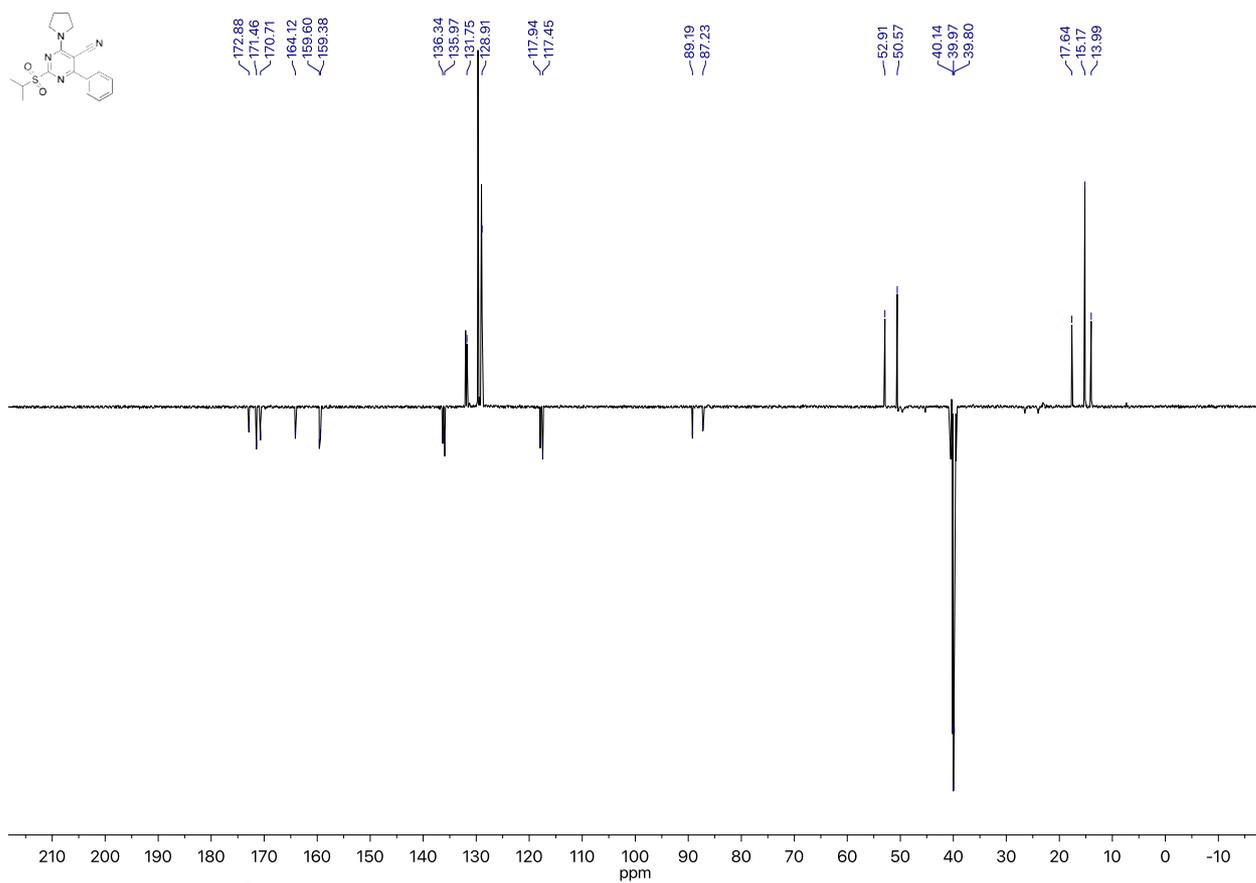


Figure S35. The ¹³C NMR spectrum of **2a**.

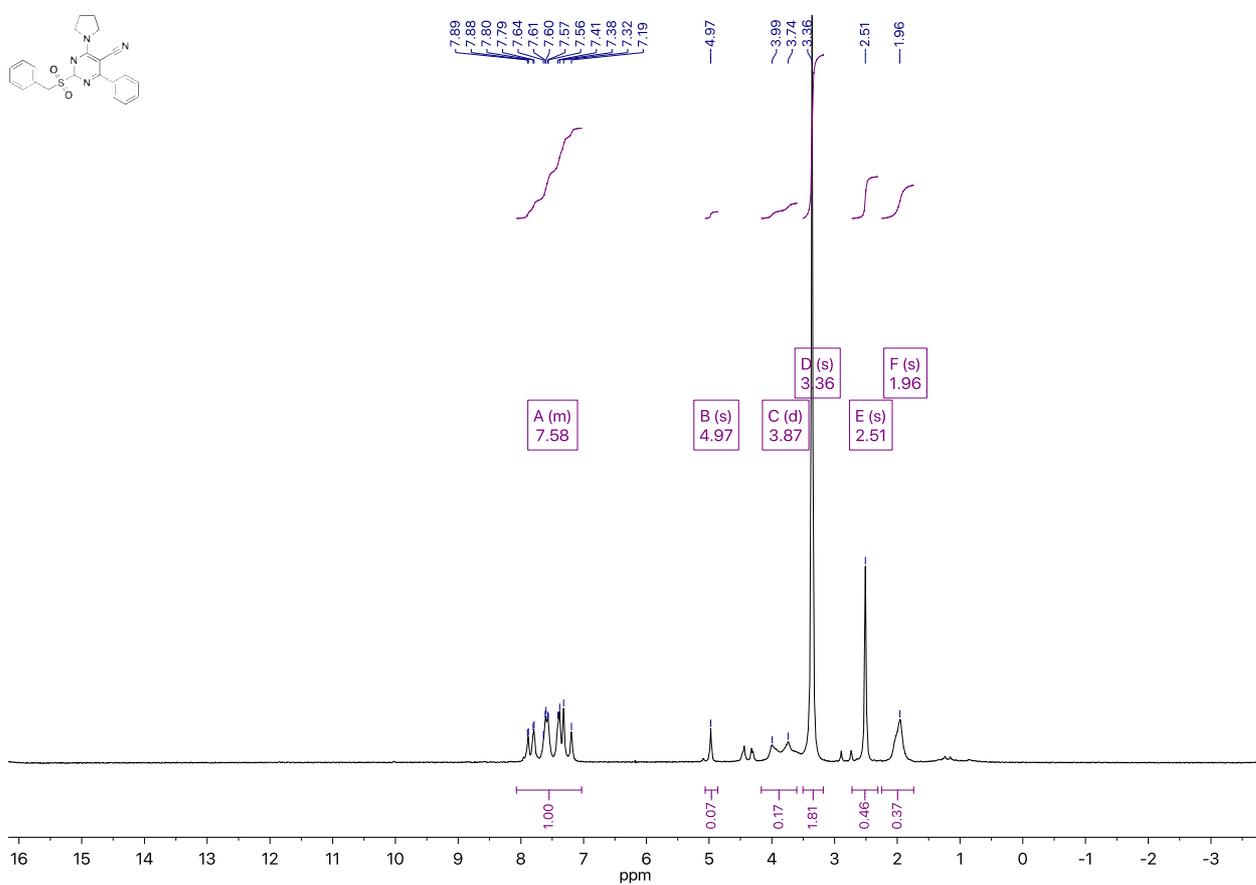


Figure S36. The ¹H NMR spectrum of **2b**.

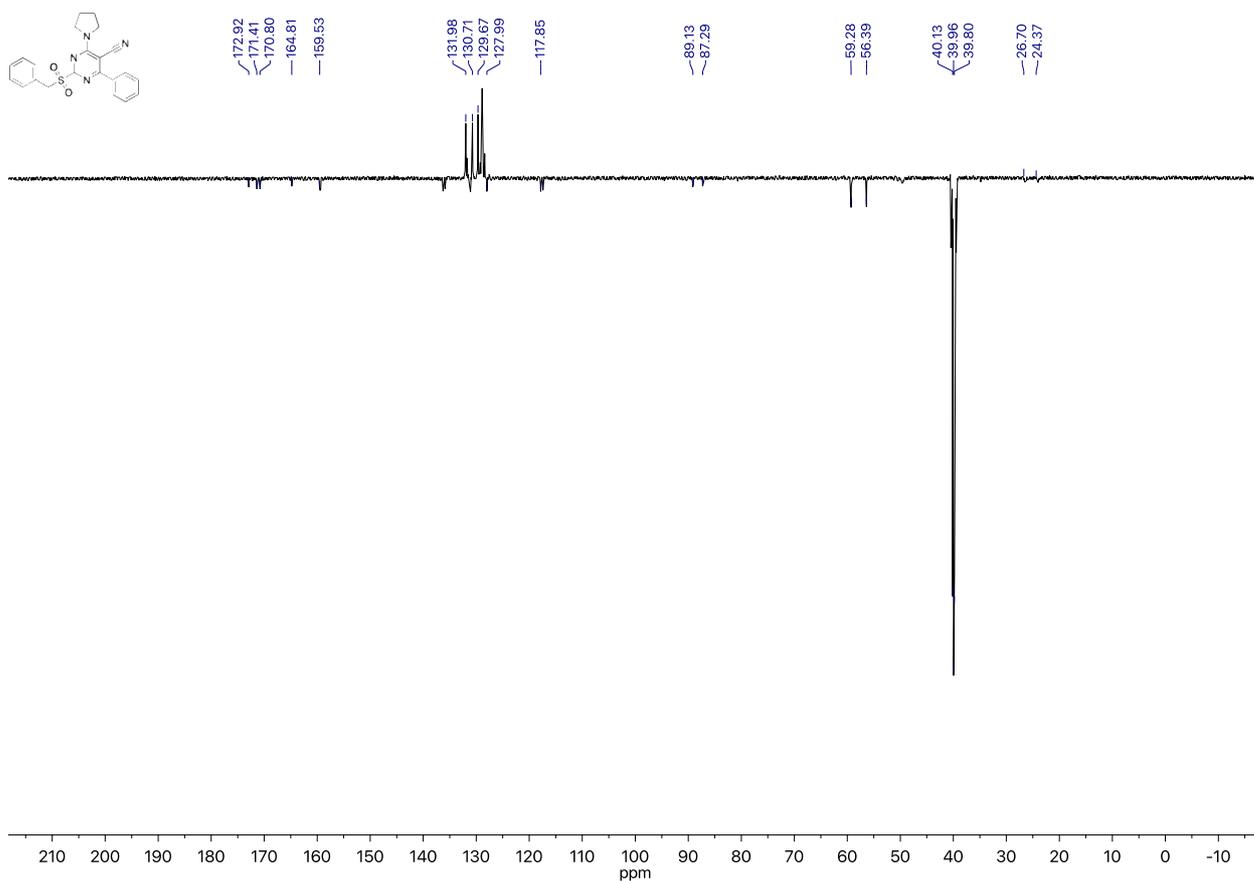


Figure S37. The ¹³C NMR spectrum of **2b**.

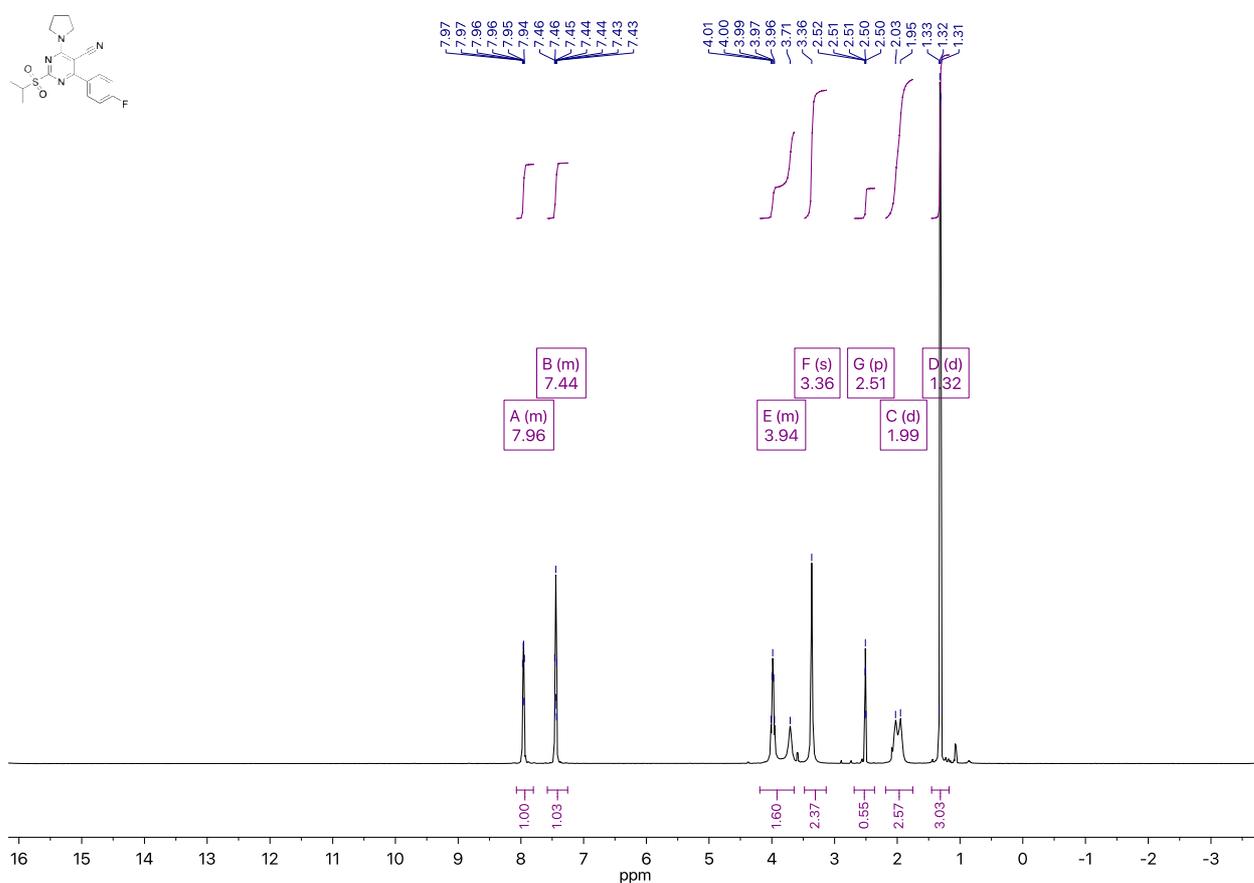


Figure S38. The ¹H NMR spectrum of **2c**.

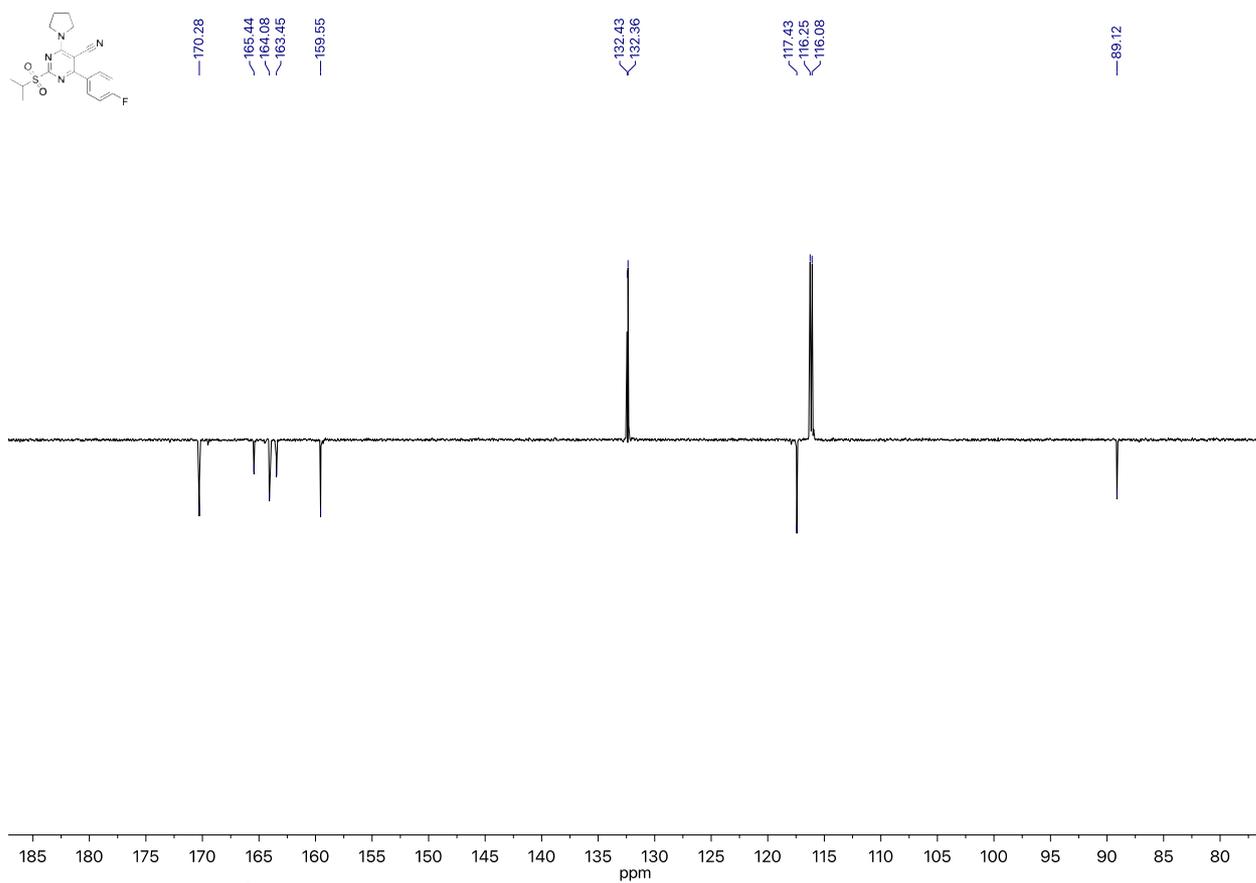


Figure S39. The ^{13}C NMR spectrum of **2c**.

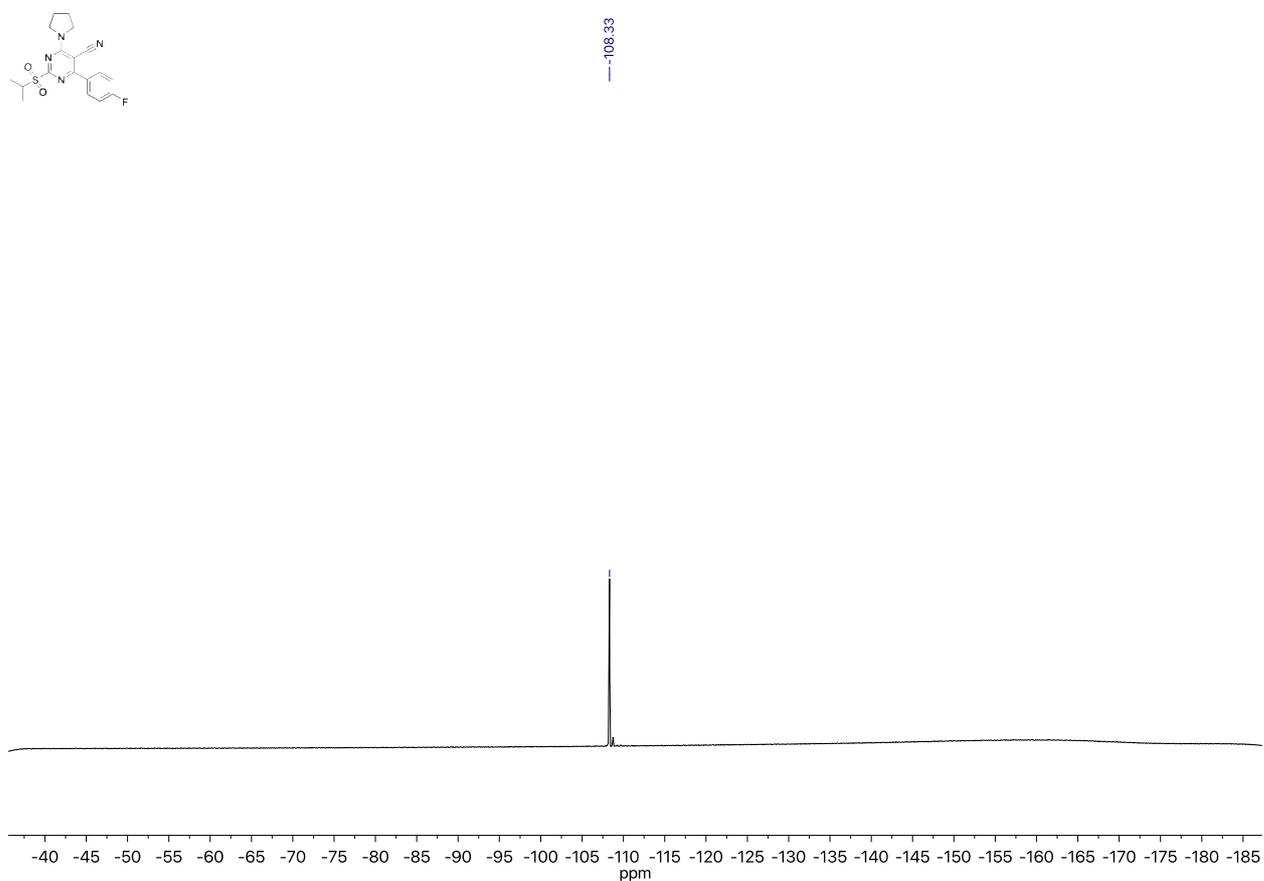


Figure S40. The ^{19}F NMR spectrum of **2c**.

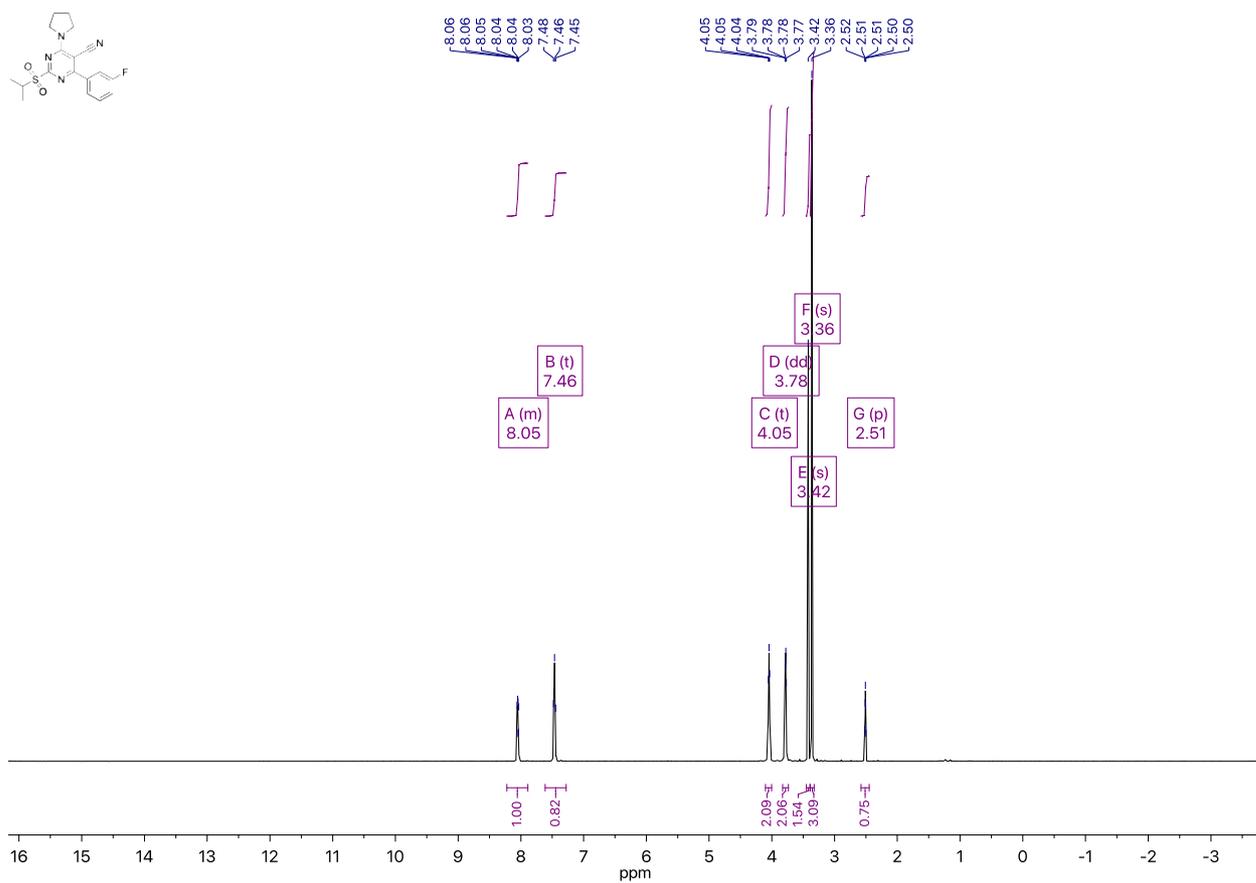


Figure S41. The ^1H NMR spectrum of **2d**.

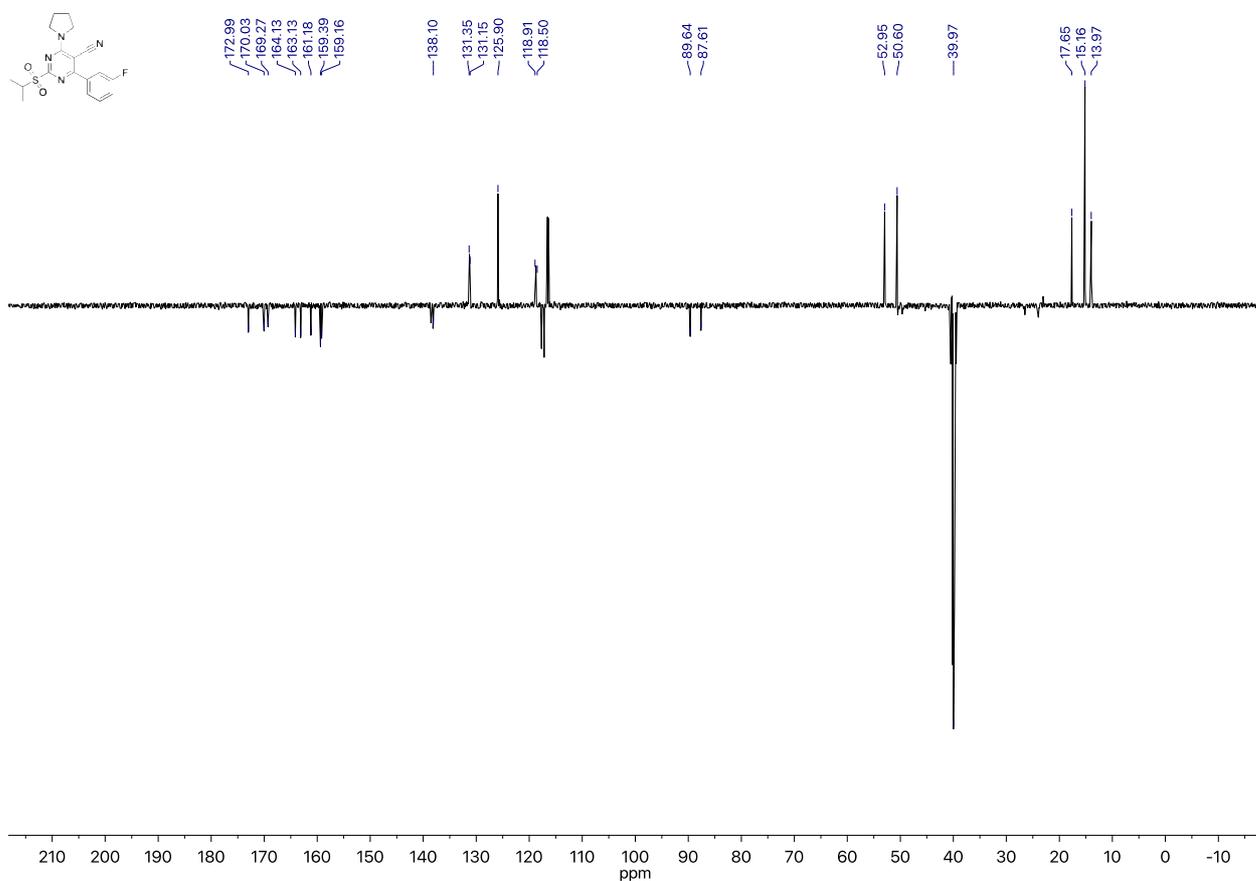


Figure S42. The ^{13}C NMR spectrum of **2d**.

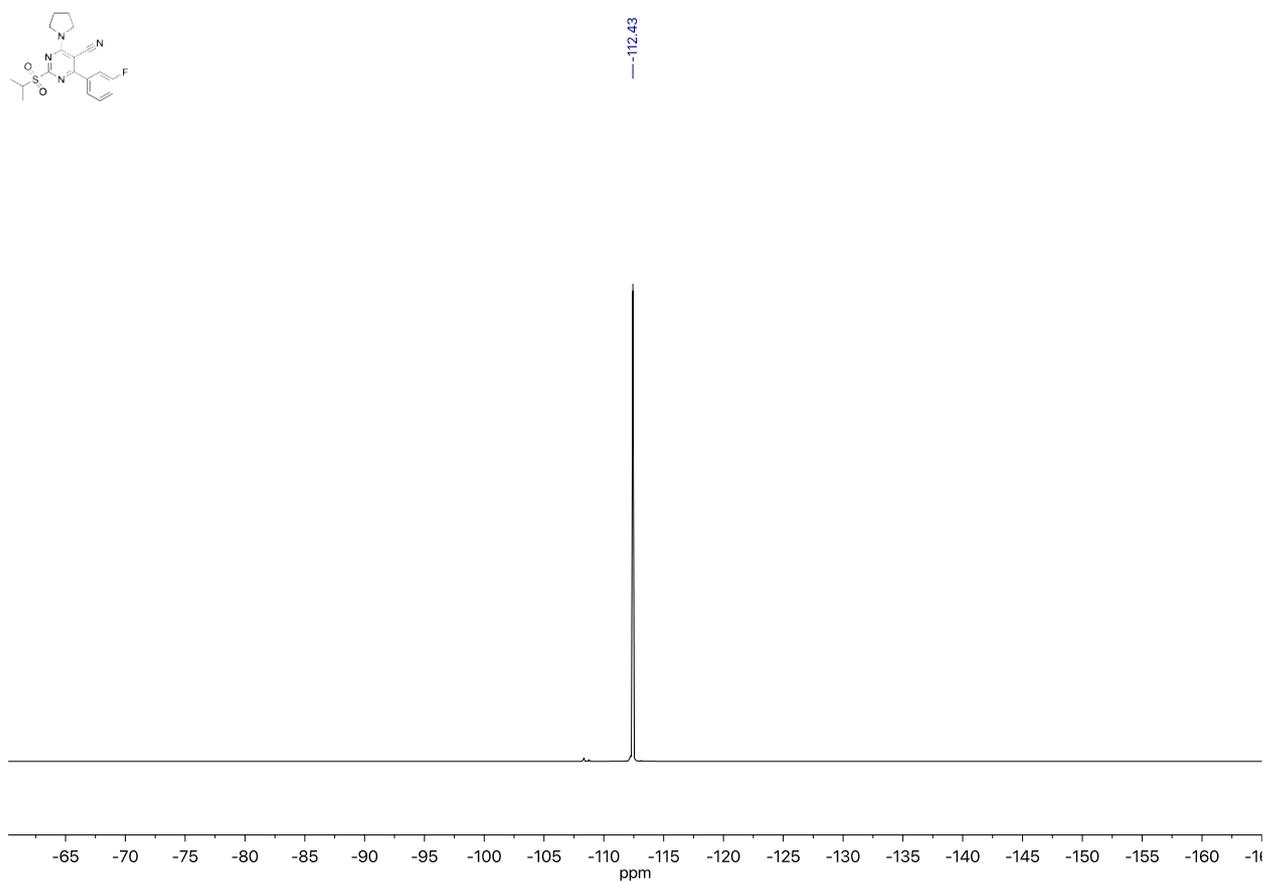


Figure S43. The ^{19}F NMR spectrum of 2d.

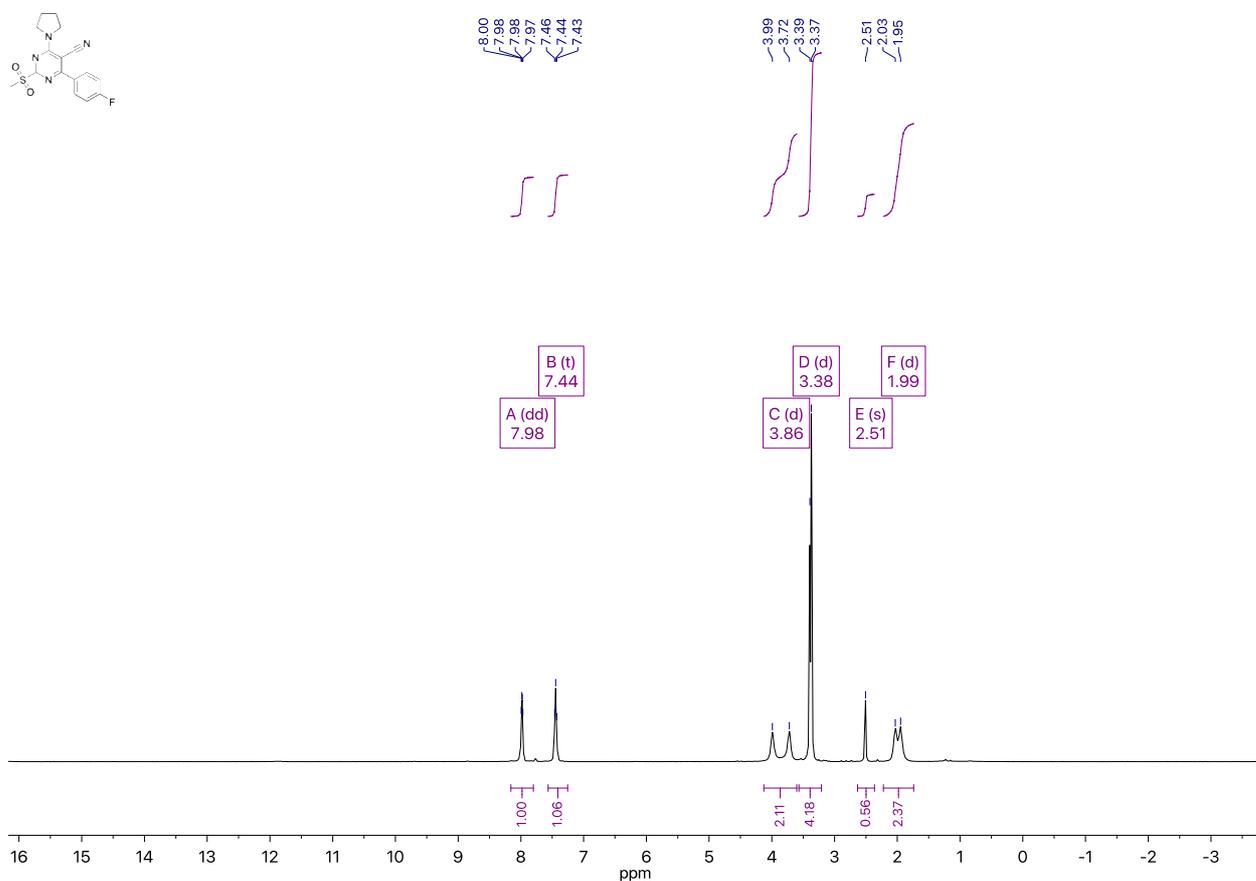


Figure S44. The ^1H NMR spectrum of 2e.

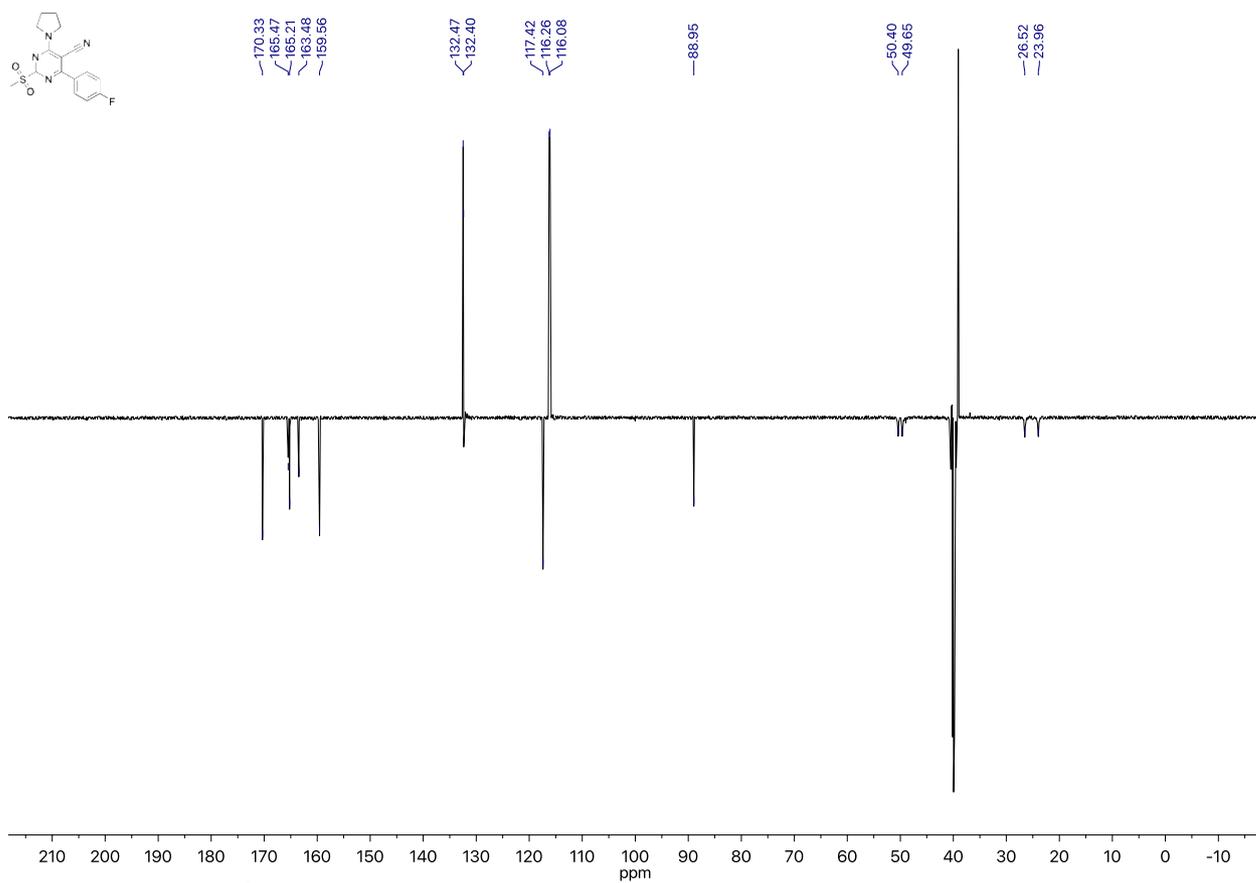


Figure S45. The ^{13}C NMR spectrum of 2e.

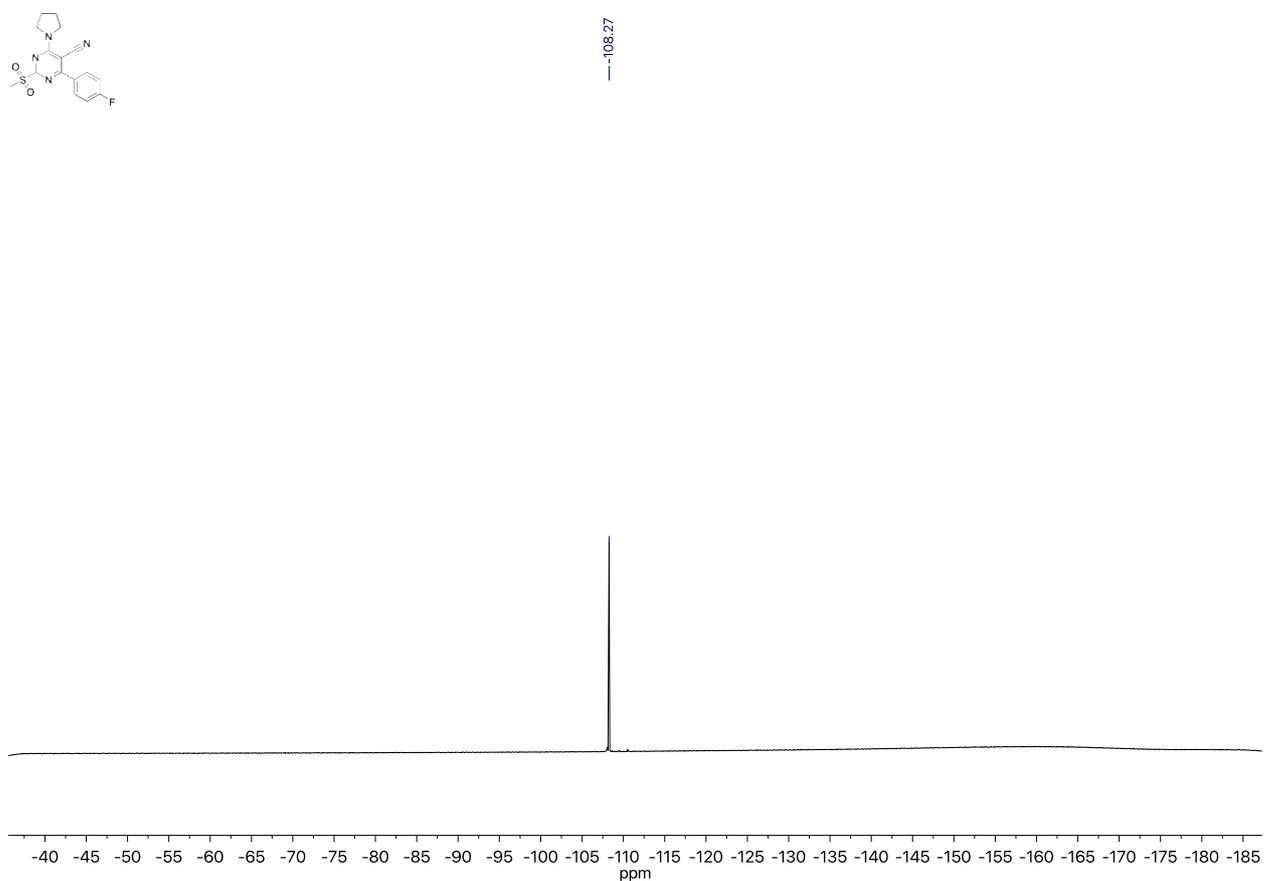


Figure S46. The ^{19}F NMR spectrum of 2e.

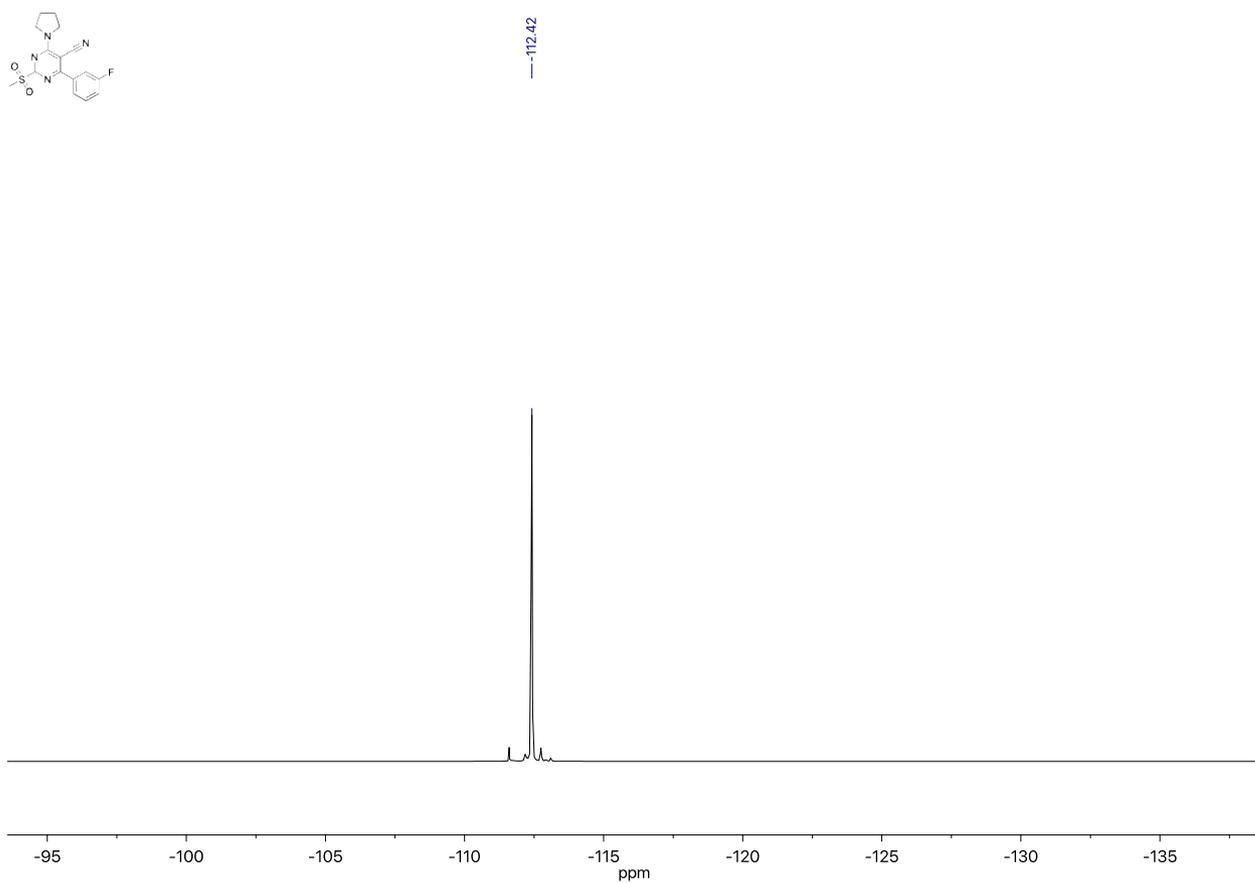


Figure S50. The ^{19}F NMR spectrum of **2f**.

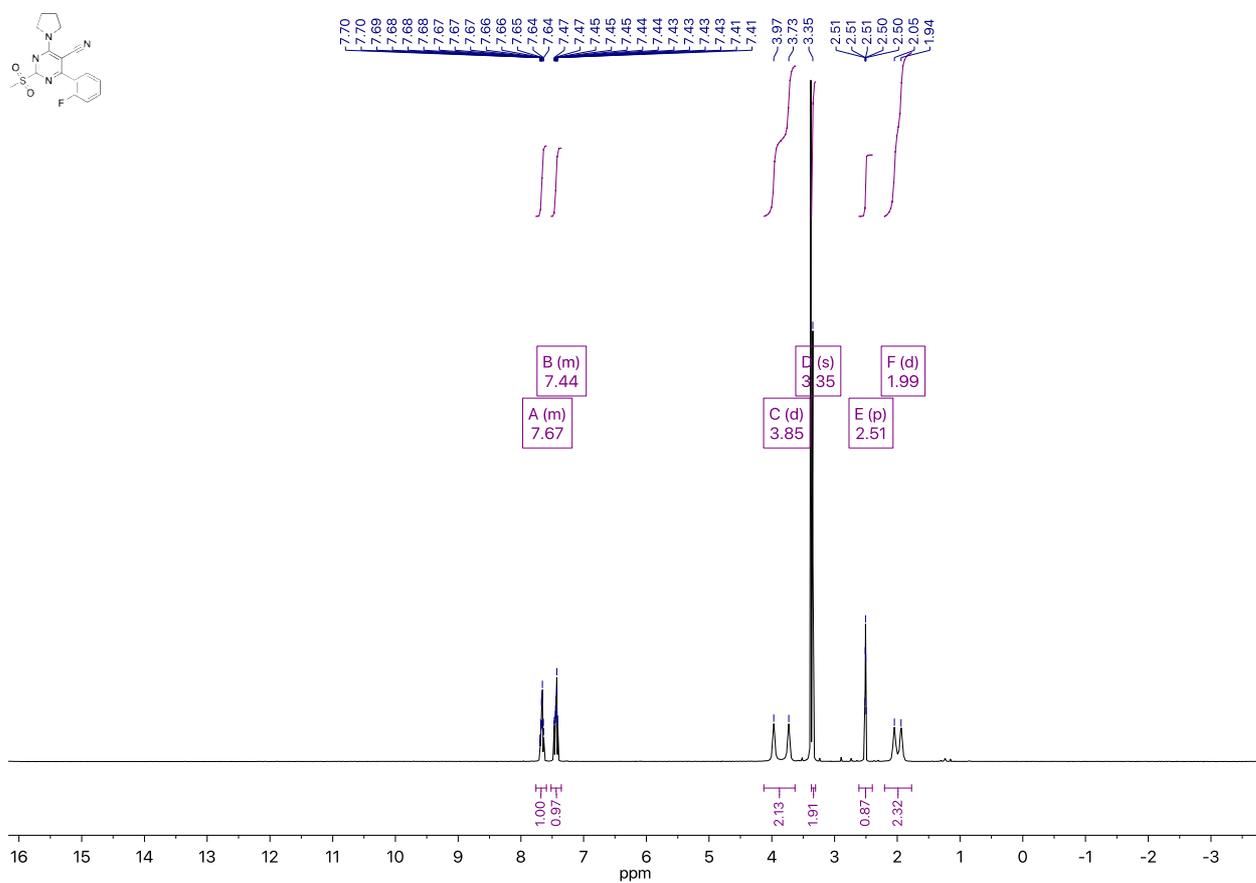


Figure S51. The ^1H NMR spectrum of **2g**.

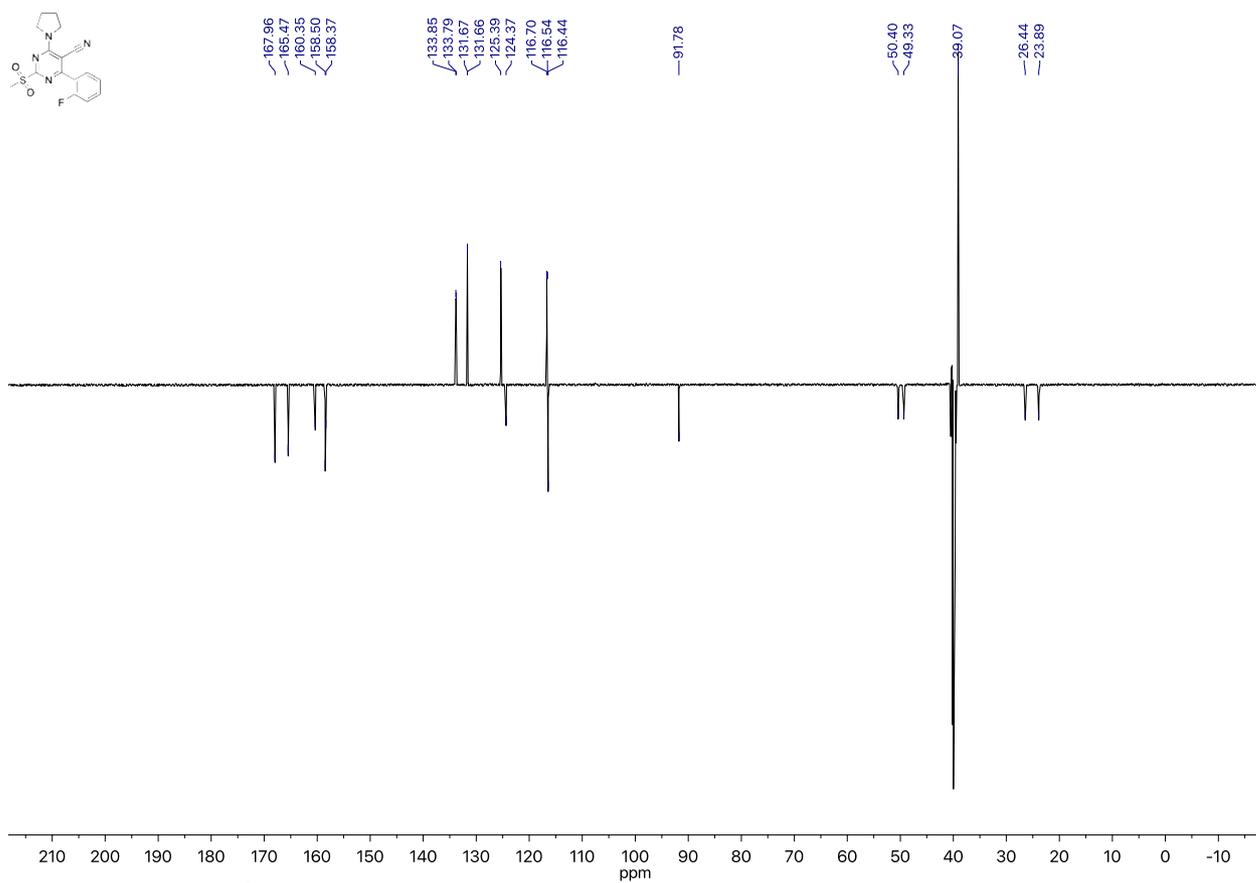


Figure S52. The ¹³C NMR spectrum of **2g**.

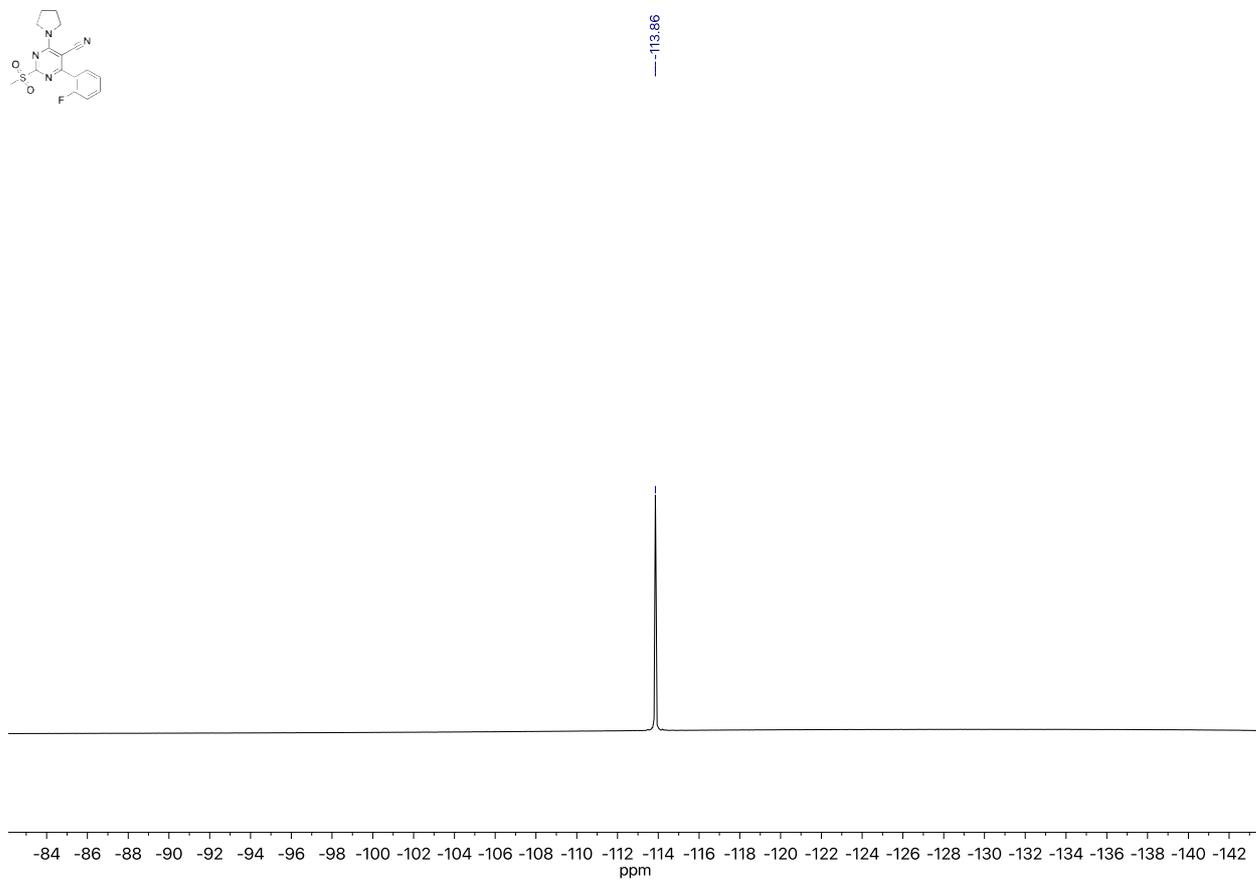


Figure S53. The ¹⁹F NMR spectrum of **2g**.

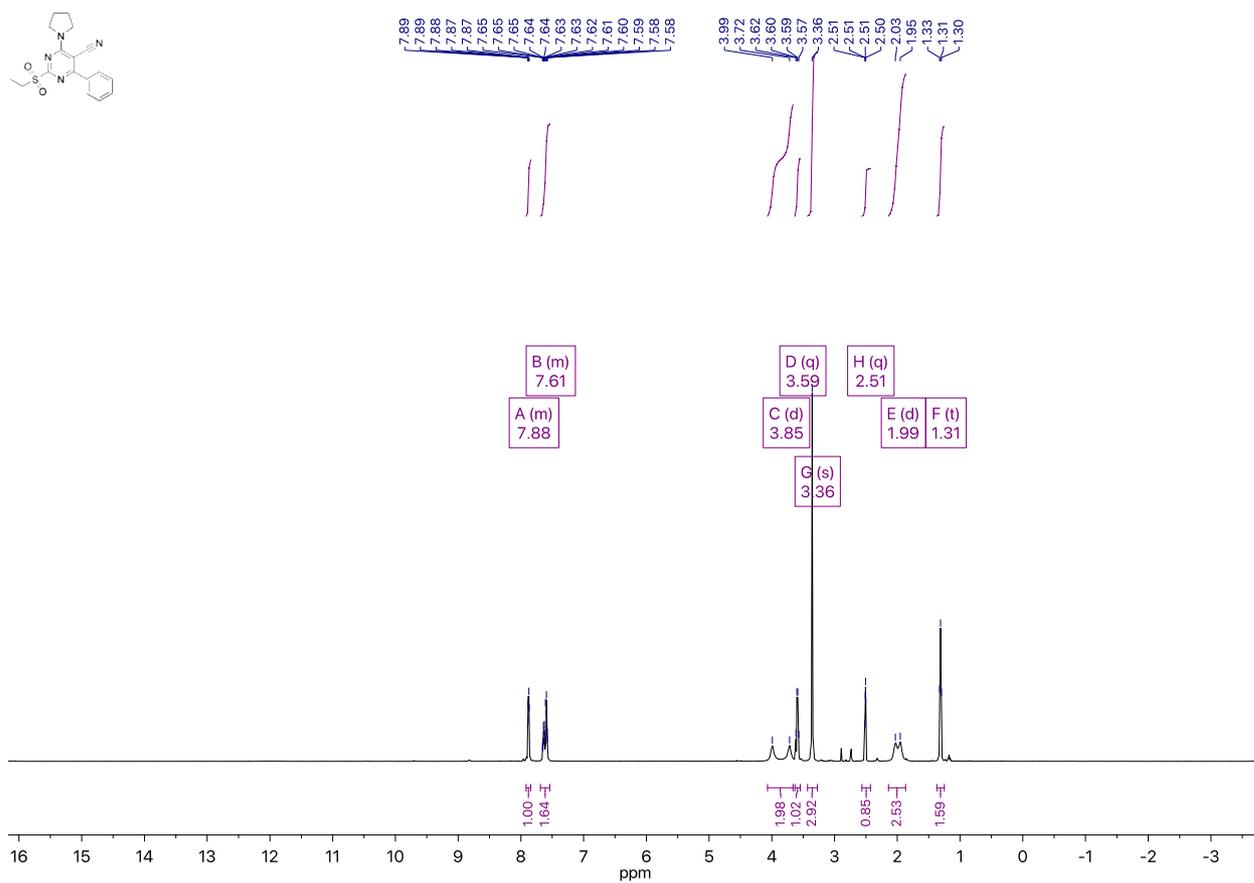


Figure S54. The ¹H NMR spectrum of **2h**.

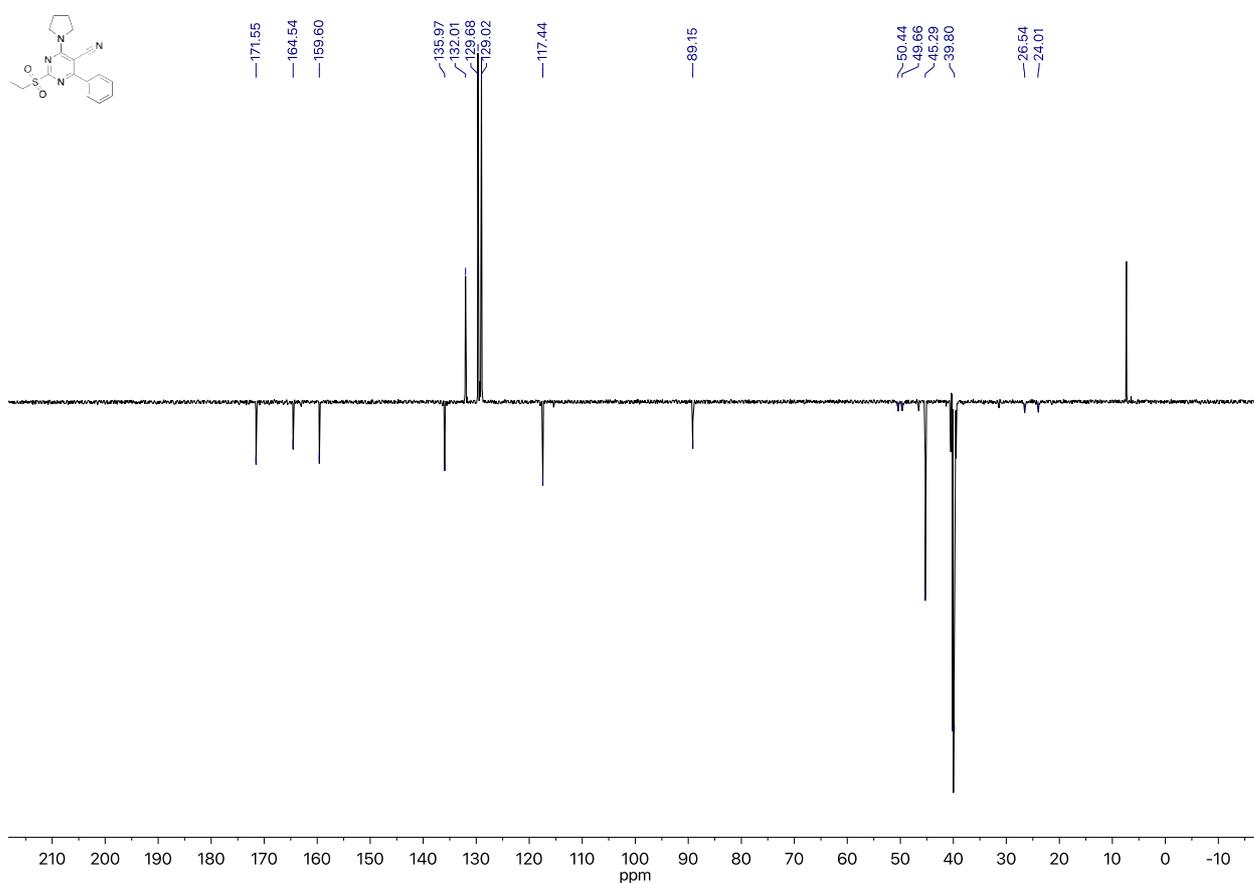


Figure S55. The ¹³C NMR spectrum of **2h**.