

Synthesis of polyester from renewable feedstock: a comparison between microwave and conventional heating

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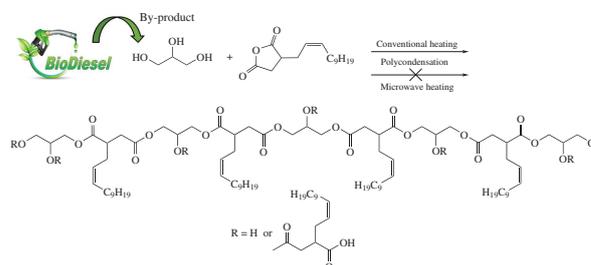
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Synthesis of polyester using (dodec-2-enyl)succinic anhydride and renewable glycerol was carried out to produce a polymer with controlled functionality. Conventional (hot plate) heating was found suitable to obtain oligomers, while the microwave heating was useful only at the first step of the process.



Synthesis of functional polyesters from renewable monomers is of great interest in polymer research^{1–3} due to the large-scale industrial needs for polymers of this class in addition to a common trend of the replacement of petroleum precursors by renewable ones. In this work, we have investigated the opportunity to synthesize the polyester bearing free hydroxyl groups from glycerol by considering the selective reactivity of alcohol groups.⁴ A similar approach was previously reported for the synthesis of poly(glycerol adipate).^{5,6} However, to impart hydrophobicity, the free hydroxyl groups in the latter were esterified with fatty acids, which hinders further functionalization with additional functional groups of interest.

Taking into account the above mentioned limitation, we have selected a more hydrophobic derivative of succinic anhydride, (dodec-2-enyl)succinic anhydride (DDSA), for the polyester synthesis. DDSA has been widely applied for modifications of biopolymers and resins.⁷ The synthesis of alkenylsuccinic anhydride from sunflower oil has been reported, which could make DDSA a potentially renewable monomer.⁸ In order to generate a solvent-free product and to reduce the cost of oligomer synthesis, reactions were performed in bulk.

Microwave radiation as a heat source has been known to decrease the reaction temperature and time^{9,10} to make synthesis processes more commercially viable and eco-friendly and, at the same time, reducing the potential of generation of unwanted by-products.^{11,12} Thus, the microwave assisted synthesis was also implemented in this work. The one-step synthesis of polyester at a high temperature *in vacuo*, unfortunately, resulted in a cross-linked material. Therefore, a step-wise approach was selected for the preparation of oligomers and polymers from DDSA in order to increase the level of control over the final polymeric structure. An additional reason behind adopting this strategy was the acquisition of characterization data on oligomers possessing different molecular weights and containing free hydroxyl units.

Scheme 1 shows the synthetic approach to prepare oligomers of DDSA and glycerol. The G0–Glycerol was obtained from DDSA and glycerol without using any catalyst. It was expected that the reaction of two equivalents of DDSA and one equivalent of glycerol would preferentially result in the esterification of terminal (primary) hydroxyl groups of glycerol and the product would contain one free secondary alcohol group due to a higher steric hindrance and low reactivity of the latter.¹³

Since polyoxyethylenesorbitan monooleate (Tween 80, molecular weight of ~1310 Da) is known as a powerful surfactant,¹⁴ the molecular weight of 1500 Da was initially selected as the target value for the DDSA–glycerol oligomers. Conversion of monomers into oligomers was monitored by ¹H NMR spectroscopy. A shift in the peak observed between 3.0–3.2 ppm to below 3.0 ppm (proton peak associated with anhydride ring, Figure S1, Online Supplementary Materials) and appearance of a signal at 4.0–4.5 ppm indicating the ester bond formation proposed the attachment of DDSA to glycerol *via* an ester bond. The full (100%) conversion of anhydride into acid was observed within 7 h at 120 °C. However, additional signals were also detected in ¹H NMR and mass spectra of the obtained product (5.1, 5.3 and 3.6–3.8 ppm in ¹H NMR, Figure S2), which were related to tri-substituted and partially substituted glycerol (Table S1). Thus, to obtain a more clean product in shorter time, the reaction was performed upon the microwave heating. The same conversion was achieved within 2 h at 80 °C, but the undesired products were still present. Nevertheless, the sample mostly contained the desired product since the intensities of impurity signals were very low.

Subsequently, the synthesis of DDSA–glycerol species with an elongated chain using phosphoric acid as a biologically acceptable catalyst was initially attempted upon microwave heating due to its apparent advantages over the hot plate method. Unfortunately, oligomer G0–Glycerol was reflecting almost 90% of the supplied

- 2 M. Dusselier, P. Van Wouwe, S. De Smet, R. De Clercq, L. Verbelen, P. Van Puyvelde, F. E. Du Prez and B. F. Sels, *ACS Catal.*, 2013, **3**, 1786.
- 3 Z. You, H. Cao, J. Gao, P. H. Shin, B. W. Day and Y. Wang, *Biomaterials*, 2010, **31**, 3129.
- 4 M. Agach, S. Delbaere, S. Marinkovic, B. Estrine and V. Nardello-Rataj, *Polym. Degrad. Stab.*, 2012, **97**, 1956.
- 5 V. Taresco, R. G. Creasey, J. Kennon, G. Mantovani, C. Alexander, J. C. Burley and M. C. Garnett, *Polymer*, 2016, **89**, 41.
- 6 V. Taresco, J. Suksiriworapong, R. Creasey, J. C. Burley, G. Mantovani, C. Alexander, K. Treacher, J. Booth and M. C. Garnett, *J. Polym. Sci., Part A: Polym. Chem.*, 2016, **54**, 3267.
- 7 N. N. Shah, N. Soni and R. S. Singhal, *Int. J. Biol. Macromol.*, 2018, **107**, 2224.
- 8 L. Candy, C. Vaca-Garcia and E. Borredon, *Eur. J. Lipid Sci. Technol.*, 2005, **107**, 3.
- 9 F. Hild, N. T. Nguyen, E. Deng, J. Katrib, G. Dimitrakis, P.-L. Lau and D. J. Irvine, *Macromol. Rapid Commun.*, 2016, **37**, 1295.
- 10 N. Z. Kiss, Z. Rádai, I. Tihanyi, T. Szabó and G. Keglevich, *Mendeleev Commun.*, 2018, **28**, 31.
- 11 I. Bilecka and M. Niederberger, *Nanoscale*, 2010, **2**, 1358.
- 12 D. D. Stöbener, D. Donath, and M. Weinhart, *J. Polym. Sci., Part A: Polym. Chem.*, 2018, **56**, 2496.
- 13 P. Kallinteri, S. Higgins, G. A. Hutcheon, C. B. St. Pourçain and M. C. Garnett, *Biomacromolecules*, 2005, **6**, 1885.
- 14 A. Mohamed and A.-S. M. Mahfoodh, *Colloids Surf., A*, 2006, **287**, 44.
- 15 A. D. Smith, E. H. Lester, K. J. Thurecht, S. W. Kingman, J. El Harfi, G. Dimitrakis, J. P. Robinson and D. J. Irvine, *Ind. Eng. Chem. Res.*, 2010, **49**, 3011.
- 16 D. T. W. Toolan, K. Adlington, A. Isakova, A. Kalamiotis, P. Mokarian-Tabari, G. Dimitrakis, C. Dodds, T. Arnold, N. J. Terrill, W. Bras, D. Hermida Merino, P. D. Topham, D. J. Irvine and J. R. Howse, *Phys. Chem. Chem. Phys.*, 2017, **19**, 20412.
- 17 M. B. Gawande, S. N. Shelke, R. Zboril and R. S. Varma, *Acc. Chem. Res.*, 2014, **47**, 1338.
- 18 P. Lidstrom, J. Tierney, B. Wathey and J. Westman, *Tetrahedron*, 2001, **57**, 9225.

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